

Barley Genetics Newsletter

Volume 47 - 2017



Barley spikes infected with Fusarium head blight

Editorial Committee
P. Bregitzer - U. Lundqvist

Table of Contents

Descriptions of the Barley Genetic Stocks for 2017.....	1
Descriptions of Barley Genetics Stocks Tables 2 and 3 (2017).....	5
Detailed Descriptions of Barley Genetic Stocks.....	50

<<Information about the Barley Genetics Newsletter>>

The Barley Genetics Newsletter is published electronically at
<https://wheat.pw.usda.gov/ggpages/bgn>

The Barley Genetics Newsletter (BGN) was first published in 1971, and in the years since then has served to disseminate to the barley community announcements, memoria, informal research reports, and detailed descriptions of barley genetic stocks.

In recent years, new forms of rapid communication of ideas and data, principally on-line, have reduced the demand for many aspects of BGN. At the 12th International Barley Genetics Symposium, held in Minneapolis, Minnesota, USA June 26-30, 2016, discussion of the fate of BGN resulted in the consensus that a forum for informal communication, memoria, documentation of resources, and any other topic of interest to the community was beneficial.

Thanks to resources provided by GrainGenes, past issues of BGN will be preserved and available in electronic format. GrainGenes will serve also as a mechanism for publishing new submissions from the barley community.

Contributions to the Barley Genetics Newsletter can be sent to Phil Bregitzer, phil.bregitzer@ars.usda.gov. Dr. Bregitzer can be reached also at 208-397-4162 ext. 116.

Acknowledgements

BGN has been made possible by the contributors of research reports, the diligence of the many coordinators, and by the special efforts of leaders in the barley genetics community. Of special note, the compilation of the detailed and extensive new and updated barley genetic stock descriptions published in the last several issues of BGN has been made possible by the time and expertise of Jerry Franckowiak and Udda Lundqvist. Special thanks to the USDA-ARS GrainGenes team.

Descriptions of Barley Genetic Stocks for 2017

Jerome D. Franckowiak¹, and Udda Lundqvist²

**¹Department of Agronomy and Plant Genetics
University of Minnesota Twin Cities
411 Borlaug Hall
1991 Upper Buford Circle
St Paul, MN 55108, USA**

**²Nordic Genetic Resource Center (NordGen)
P.O. Box 41, SE-230 53 Alnarp, Sweden**

**e-mail: jfrancko@umn.edu
udda.lundqvist@nordgen.org**

In this volume of the Barley Genetics Newsletter seventy revised and new Barley Genetic Stock (BGS) descriptions are published (Table 1). The current lists of new and revised BGS descriptions, including those in Table 1, are presented by BGS number order (Table 2) and by locus symbol in alphabetic order (Table 3) in another section of this issue. Information on the description location, recommended locus name, chromosomal location, previous gene symbols, and the primary genetic stock (GSHO number and/or NGB number) are included in these lists. The GSHO stocks are held in the USDA-ARS Barley Genetic Stocks collection at the National Small Grains Collection (U.S. Department of Agriculture – Agricultural Research Service, National Small Grains Germplasm Research Facility, 1691 S 2700 W) Aberdeen, ID 83210, USA. The NGB stocks are held in the Nordic Genetic Resource Center (NordGen), Smedjevägen 3, SE-230 53 Alnarp, Sweden. This information is available through the Internet at the following addresses:

- (1) [www.ars.usda.gov.PacWest/Aberdeen](http://www.ars.usda.gov/PacWest/Aberdeen)**
- (2) [www.ars-grin.gov:7000/npgs/descriptors/barley-genetics](http://www.ars-grin.gov/7000/npgs/descriptors/barley-genetics) (GRIN)**
- (3) <http://wheat.pw.usda.gov/ggpages/bgn/>**
- (4) <http://www.nordgen.org/sesto>**
- (5) <http://www.nordgen.org/bgs>**

Table 1. A listing of new and revised Barley Genetic Stock (BGS) descriptions published in volume 47 of the Barley Genetics Newsletter, giving recommended locus symbols and names, chromosomal locations, and stock source information.

BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
1	brh1	br, ari-i	7HS	Brachytic 1	47:50	25
51	rtt1	rt	2HS	Rattail spike 1	47:53	216
65	Eam1	Ea, Ppd-H1	2HS	Early maturity 1	47:54	1316
67	Vrs1.t	V ^t	2HL	Six-rowed spike 1 (Deficiens 1)	47:57	684
69	Gth1	G	2HL	Toothed lemma1	47:59	309
79	wst7	rb	2HL	White streak 7	47:61	247
82	Zeo1	Ert-r, Knd	2HL	Zeocriton 1	47:63	1613
89	ari-g	lk10	2H	Breviaristatum-g	47:66	1655
97	acr1	acr, ril	2HL	Accordion rachis 1	47:68	1617
102	uzu1	uz, ert-ii, HvBRI1	3HL	Uzu 1 or Semibrachytic 1	47:70	1300
118	Int1	rnt, int-l	3HL	Lower number of tillers 1	47:74	833
124	vrs4	v4, mul, int-e, HvRA2	3HL	Six-rowed spike 4	47:76	775
131	gra-a	gran-a	7H	Granum-a	47:79	1757
155	glf1	gl1, cer-zh	4HL	Glossy leaf 1	47:81	98
157	brh2	br2, ari-l	4HL	Brachytic 2	47:84	573
165	glf3	gl3, cer-j	4HL	Glossy leaf 3	47:86	577
172	lks5	lk5, ari-c	4HL	Short awn 5	47:89	1297
178	int-c	i, v5, HvTB1	4HS	Intermedium spike-c	47:91	776
184	Zeo3	Mo1, Zeo2, cly1.c	2HL	Zeocriton 3	47:95	1611
188	Alp1	Alp, Pht	4HL	Aluminum tolerance 1	47:98	1297
189	Acr2	acr, lax	4HL	Accordion rachis 2	47:101	1071
229	cud2	cud2	1HL	Curly dwarf 2	47:103	1712
238	ari-t	ari-25	1H	Breviaristatum-t	47:104	1611
241	Acr3	acr	1HL	Accordion rachis 3	47:105	1071
258	dsp9	l9, ert-e	6HL	Dense spike 9	47:107	1774
266	ert-e	l9, dsp9	6HL	Erectoides-e	47:109	477
275	lfl1	lfl	6HL	Leafless 1	47:111	609
314	vrs2	v2, int-b	5HL	Six-rowed spike 2	47:113	773
315	vrs3	v3, int-a	1HL	Six-rowed spike 3	47:115	774
320	int-b	vrs2	5HL	Intermedium spike-b	47:118	1764
328	ari-e	lk9, GPert	5HL	Breviaristatum-e	47:120	1653
332	Ert-r	Zeo1	2HL	Erectoides-r	47:123	492
352	gsh2	gs2, cer-b	3HL	Glossy sheath 2	47:125	736
355	gsh5	gs5, cer-s	2HL	Glossy sheath 5	47:128	739
402	cer-g	cer-g	2HL	Eceriferum-g	47:131	428
Table 1. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				

439	cer-zv	cer-zv	4HL	Eceriferum-zv	47:133	1529
473	com1	lax-270, bir1	5HL	Compositum 1	47:135	1702
475	lax-c	lax-21	6HL	Laxatum-c	47:137	1777
476	lax-d	lax-83		Laxatum-d	47:139	
477	lax-f	lax-61		Laxatum-f	47:140	
478	lax-g	lax-25		Laxatum-g	47:141	
479	lax-h	lax-26	3H	Laxatum-h	47:142	
480	lax-i	lax-50		Laxatum-i	47:144	
481	lax-j	lax-49	2H	Laxatum-j	47:145	
482	lax-k	lax-84	2H/3H	Laxatum-k	47:147	
483	lax-l	lax-60	1H	Laxatum-l	47:148	
484	lax-m	lax-80		Laxatum-m	47:149	
485	lax-n	lax-67	2H/4HL /5H	Laxatum-n	47:150	
486	lax-o	lax-79		Laxatum-o	47:152	
487	lax-df	lax-204		Laxatum-df	47:153	
488	lax-ef	lax-225	2H	Laxatum-ef	47:154	
489	lax-ff	lax-216		Laxatum-ff	47:156	
490	lax-gf	lax-217		Laxatum-gf	47:157	
491	lax-hf	lax-244	2H/3HL /5H	Laxatum-hf	47:158	
492	lax-if	lax-246		Laxatum-if	47:160	
493	Lax-jf	lax-253	4HS	Laxatum-jf	47:161	1571
494	lax-kf	lax-295		Laxatum-kf	47:163	
495	lax-lf	lax-274		Laxatum-lf	47:164	
496	lax-mf	lax-302		Laxatum-mf	47:165	
497	lax-nf	lax-322	2HL/4H	Laxatum-nf	47:166	
518	sdw1	denso, Hv20ox ₂	3HL	Semidwarf 1	47:168	2513
525	cer-yl	cer-yl	4HL	Eceriferum-yl	47:171	1545
526	cer-ym	cer-ym	4HL	Eceriferum-ym	47:173	1546
554	ari-m	brh1	7HS	Breviaristatum-m	47:175	1661
614	Zeo2	Zeo, cly1.b, HvAP2	2HL	Zeocriton 2	47:177	637
679	acr4	acr-3	2H/6HL	Accordion rachis 4	47:181	
740	Pbg1	Pbg	7H	Pubescence on glume 1	47:182	
741	stb1	stb		Stubble 1	47:183	
743	twk1	tw	7H	Tweaky spike 1	47:185	
744	acr5	acr-2	7H	Accordion rachis 5	47:187	2153

* Recommended locus symbols are based on utilization of a three letter code for barley genes as approved at the business meeting of the Seventh International Barley Genetics Symposium at Saskatoon, Saskatchewan, Canada, on 05 August 1996.

† Chromosome numbers and arm designations are based on a resolution passed at the business meeting of the Seventh International Barley Genetics Symposium at Saskatoon, Saskatchewan, Canada, on 05 August 1996. The Burnham and Hagberg (1956) designations of barley chromosomes were 1 2 3 4 5 6 and 7 while new designations based on the Triticeae system are 7H 2H 3H 4H 1H 6H and 5H, respectively.

‡ The seed stock associated with each BGS number is held as a GSHO stock number in the Barley Genetics Stock Collection at the USDA-ARS National Grains Germplasm Research Facility, Aberdeen, Idaho, USA.

Descriptions of Barley Genetic Stocks Tables 2 and 3 (2017)

Jerome D. Franckowiak¹ and Udda Lundqvist²

**¹ Department of Agronomy and Plant Genetics
University of Minnesota Twin Cities
411 Borlaug Hall, 1991 Upper Buford Circle
St Paul, MN 55108, USA**

**² Nordic Genetic Resource Centre (NordGen)
Smedjevägen 3, SE-230 53 Alnarp, Sweden**

**e-mail: jfrancko@umn.edu
udda.lundqvist@nordgen.org**

In this section of the Barley Genetics Newsletter volume 47, you will find two updated tables with new and revised barley locus descriptions. The descriptions are listed by BGS numbers (Table 2) and by alphabetic order using the existing and recommended locus symbols (Table 3). As research in barley is proceeding rapidly, it is necessary to update the latest research and findings about specific barley genes.

Table 2. A listing of Barley Genetic Stock (BGS) descriptions in volume 47 and recent issues of the Barley Genetics Newsletter with recommended locus symbols, recommended locus names, chromosome location information, description citation, and stock location information.

Table 3. An alphabetic listing of recently published Barley Genetic Stock (BGS) descriptions for loci in barley (*Hordeum vulgare*), including information on recommended locus names, recommended locus symbols, chromosomal locations, description citation, and original cultivars.

Table 2. A listing of Barley Genetic Stock (BGS) descriptions in recent issues of the Barley Genetics Newsletter with recommended locus symbols, recommended locus names, chromosome location information, description citation, and stock location information.

BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
1	brh1	br, ari-i	7HS	Brachytic 1	47: 50	25
2	fch12	f _c , clo-fc	7HS	Chlorina seedling 12	41: 60	36
3	yvs2	y _c	7HS	Virescent seedling 2	26: 46	41
4	abo8	ac ₂ , alb-m	7HS	Albino seedling 8	26: 47	61
5	fch8	f8	7HS	Chlorina seedling 8	41: 62	40
6	vrs1	v, Int-d	2HL	Six-rowed spike 1	37:192	196
7	nud1	n, h	7HL	Naked caryopsis 1	44: 51	115
9	dsp1	l	7HS	Dense spike 1	43: 50	1232
10	lks2	lk2, lk4	7HL	Short awn 2	45: 80	566
11	ubs4	lks2, ari-d	7HL	Unbranched style 4	45: 84	567
12	des1	lc	7H	Desynapsis 1	42: 58	592
13	des4	des4	7H	Desynapsis 4	44: 54	595
14	des5	des5	7HL	Desynapsis 5	44: 56	596
15	blx1	bl	4HL	Non-blue aleurone xenia 1	26: 60	185
16	wax1	wx, glx	7HS	Waxy endosperm 1	42: 65	908
17	fch4	f4, yv	7HL	Chlorina seedling 4	43: 54	1214
18	fch5	f5, yv2	7HS	Chlorina seedling 5	43: 56	1215
19	blx2	bl2	7HS	Non-blue aleurone xenia 2	26: 65	209
20	Rym2	Ym2	7HL	Reaction to BaYMV 2	26: 66	984
21	Run1	Un	7HS	Reaction to <i>Ustilago nuda</i> 1	26: 67	1324
22	Rsg1	Grb	7H	Reaction to <i>Schizaphis graminum</i> 1	37:199	1317
23	wnd1	wnd	4HL	Winding dwarf 1	42: 74	2499
24	fst3	fs3	7HS	Fragile stem 3	41: 74	1746
25	Xnt1	X _a	7HL	Xantha seedling 1	26: 71	1606
26	snb1	sb	7HS	Subnodal bract 1	26: 72	1217
27	lbi3	lb3	7HL	Long basal rachis internode 3	42: 79	536
28	ert-a	ert-a	7HS	Erectoides-a	41: 76	468
29	ert-d	ert-d	7HS	Erectoides-d	42: 82	475
30	ert-m	ert-m	7HS	Erectoides-m	46: 47	487
31	sex6	ssIIa	7HS	Shrunken endosperm xenia 6	45: 86	2476
32	Rph9	Pa9, Pa12	5HL	Reaction to <i>Puccinia hordei</i> 9	37:201	1601

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
33	ant1	rs, rub-a	7HS	Anthocyanin-less 1	46: 49	1620
34	msg50	msg,,hm	7HL	Male sterile genetic 50	45: 88	2404
35	rsm1	sm	7HS	Reaction to BSMV 1	26: 84	2492
36	xnt4	x _{c2}	7HL	Xantha seedling 4	26: 85	42
37	xnt9	xan,,i	7HL	Xantha seedling 9	26: 86	584
38	smn1	smn	3H/5H	Seminudoides 1	43: 58	1602
39	mss2	mss2	7HS	Midseason stripe 2	44: 59	2409
40	prm1	prm	7HS	Premature ripe 1	44: 60	2429
41	brh7	brh.w	7H	Brachytic 7	42: 98	1687
42	Pyr1	Pyr.g, Pyr.i	3HL	Pyramidatum 1	41: 78	1581
43	mov1	mo5	7HL	Multiovary 1	43: 59	3641
44	brh16	brh.v, ari-o	7HL	Brachytic 16	45: 89	1686
45	sdw4		7HL	Semidwarf 4	46: 51	
48	Rpt4	QRpt7	7HL	Reaction to <i>Pyrenophora teres</i> 4	43: 61	
49	sld8	sld.i	7HS/ 4HL	Slender dwarf 8	43: 63	2484
51	rtt1	rt	2HS	Rattail spike 1	47: 53	216
52	fch15	or	2HS	Chlorina seedling 15	40: 48	49
53	abo2	a2	2HS	Albino seedling 2	26: 89	70
55	fch1	f, lg	2HS	Chlorina seedling 1	40: 49	112
56	wst4	wst4	2HL	White streak 4	44: 61	568
57	eog1	e, lep-e	2HL	Elongated outer glume 1	43: 64	29
58	vrs1	lr, v ^{lr}	2HL	Six-rowed spike 1	26: 94	153
59	gpa1	gp, gp2	2HL	Grandpa 1	45: 91	1379
60	lig1	li, aur-a	2HL	Liguleless 1	45: 93	6
61	trp1	tr	4HL	Triple awned lemma 1	41: 82	210
62	sbk1	sk, cal-a	2HS	Subjacent hood 1	40: 51	267
63	yvs1	y _x , alb-c.7	2HS	Virescent seedling 1	26: 99	68
64	des7	des7	3H	Desynapsis 7	43: 67	598
65	Eam1	Ea, Ppd-H1	2HS	Early maturity 1	47: 54	1316
66	vrs1	V ^d	2HL	Two-rowed spike 1	26:103	346
67	vrs1.t	V ^t	2HL	Deficiens 1	47: 57	684

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
68	Pvc 16	Pc	2HL	Purple veined lemma 1	44: 67	132
69	Gth 1	G	2HL	Toothed lemma 1	47: 59	309
70	Rph1	Pa	2H	Reaction to <i>Puccinia hordei</i> 1	26:107	1313
71	com2	bir2	2HS	Compositum 2	45: 95	1700
74	flo-c	flo-a	6HL	Extra floret-c	45: 97	1743
75	Lks1	Lk	2HL	Awnless 1	41: 84	44
76	Pre2	Re2, P	2HL	Red lemma and pericarp 2	46: 53	234
77	hcm1	h	2HL	Short culm 1	26:113	2492
78	mtt4	mtt,,e, mt	2HL	Mottled leaf 4	41: 86	1231
79	wst7	rb	2HL	White streak 7	47: 61	247
80	ant2	pr, rub	2HL	Anthocyanin-less 2	46: 55	1632
81	gsh7	gs7	1H/2H /5H	Glossy sheath 7	40: 55	1759
82	Zeo1	Knd, Ert-r	2HL	Zeocriton 1	47: 63	1613
83	sld2	sld2	2HS	Slender dwarf 2	44: 74	2491
84	mss1	mss	5H	Midseason stripe 1	44: 75	1404
85	yst4	yst4	2HL	Yellow streak 4	44: 76	2502
86	fch13	f13	5HL	Chlorina seedling 13	44: 77	16
87	fch14	f14	2HL	Chlorina seedling 14	44: 78	1739
88	Rph2	Pa2, A	5HS	Reaction to <i>Puccinia hordei</i> 2	37:212	1593
89	ari-g	lk10	2H	Breviaristatum-g	47: 66	1655
90	ert-j	ert-j	2H	Erectoides-j	43: 70	484
91	ert-q	ert-q	6H	Erectoides-q	43: 71	1562
92	ert-u	br5, ari-o	7HL	Erectoides-u	45:100	496
93	ert-zd	br7, ari-o	7HL	Erectoides-zd	45:102	504
94	abo4	a4	2H	Albino seedling 4	26:133	167
95	abo13	alb,,p	2HL	Albino seedling 13	26:134	585
96	Rph15	Rph16	2HS	Reaction to <i>Puccinia hordei</i> 15	37:214	1586
97	acr1	acr, ril	2HL	Accordion rachis 1	47: 68	1617
98	Eam6	Ea6, Ea, HvCEN	2HS	Early maturity 6	46: 57	
99	lin1	s, rin	2HL	Lesser internode number 1	41: 92	2492
100	sld4	sld.d	2HS	Slender dwarf 4	43: 72	2479
101	als1	als	3HL	Absent lower laterals 1	43: 74	1065

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
102	uzu1	uz, ert-ii, HvBRI1	3HL	Uzu 1 or semi brachytic 1	47: 70	1300
104	yst1	yst, ys	3HS	Yellow streak 1	42:178	1140
105	xnt3	x _c , vir-1	3HS	Xantha seedling 3	26:139	66
106	abo6	a _c	3HS	Albino seedling 6	46: 60	30
107	wst1	wst, wst3	3HL	White streak 1	41: 97	159
108	alm1	al, ebu-a	3HS	Albino lemma 1	46: 61	270
109	yst2	yst2	3HS	Yellow streak 2	44: 81	570
111	dsp10	l _c	3HL	Dense spike 10	41: 99	71
112	abo9	a _n	3HS	Albino seedling 9	26:146	348
113	xnt6	x _s	3HS	Xantha seedling 6	26:147	117
114	cur2	cu2	3HL	Curly 2	44: 82	274
115	btr1	bt1	3HS	Non-brittle rachis 1	43: 78	1233
116	btr2	bt2	3HS	Non-brittle rachis 2	43: 80	842
117	fch2	f2, lg5	3HL	Chlorina seedling 2	46: 63	107
118	Int1	rnt, int-1	3HL	Low number of tillers 1	47: 74	833
119	des2	ds	3H	Desynapsis 2	43: 84	593
120	zeb1	zb	3HL	Zebra stripe 1	43: 86	1279
121	Rph3	Pa3	7HL	Reaction to <i>Puccinia hordei</i> 3	26:156	1316
122	Rph5	Pa5, Pa6	3HS	Reaction to <i>Puccinia hordei</i> 5	37:224	1597
123	Ryd2	Yd2	3HL	Reaction to BYDV 2	26:158	1315
124	vrs4	v4, int-e, HvRA2	3HL	Six-rowed spike 4	47: 76	775
125	lzd1	dw4	3H	Lazy dwarf 1	43: 87	1787
126	sld1	dw1	3HL	Slender dwarf 1	41:103	2488
127	Pub1	Pub	3HL	Pubescent leaf blade 1	44: 84	1576
128	sca1	sca	3HS	Short crooked awn 1	44: 85	2439
129	wst6	wst,j	3HL	White streak 6	41:105	2500
130	eam10	ea _{sp}	3HL	Early maturity 10	44: 86	2504
131	gra-a	gran-a	7H	Granum-a	47: 79	1757
132	ari-a	ari-a, lk7	3HS	Breviaristatum-a	41:106	1648
133	sdw2	sdw-b	3HL	Semidwarf 2	45:108	2466
134	ert-c	ert-c	3HL	Erectoides-c	41:108	471
135	ert-ii	uzu1	3HL	Erectoides-ii	45:104	483

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
136	Rph7	Pa7, Pa5	3HS	Reaction to <i>Puccinia hordei</i> 7	37:228	1318
137	Rph10	Rph10	3HL	Reaction to <i>Puccinia hordei</i> 10	26:174	1588
138	nec4	nec4	3H	Necrotic leaf spot 4	43: 88	
139	nec5	nec5	3H	Necrotic leaf spot 5	43: 89	
140	xnt8	xan,,h	3HS	Xantha seedling 8	26:177	582
141	rym5	Ym	3HL	Reaction to barley yellow mosaic virus 5	32: 90	
142	brh8	brh.ad	3HL	Brachytic 8	42:232	1671
143	sex8	sex.j	3HS	Shrunken endosperm xenia 8	43: 90	2471
144	sld5	sld5	3HS	Slender dwarf 5	44: 90	2483
146	cal-d	cal-d	3H	Calcaroides-d	40: 58	1698
147	mov2	mo	3HS	Multiovary 2	43: 91	3642
148	brh14	brh.q, ari-o	7HL	Brachytic 14	45:111	1682
149	Rpc1		3H	Reaction to <i>Puccinia coronata</i> var. <i>hordei</i> 1	37:232	1601
150	scl-b	scl-5	3H/6H	Scirpoides leaf-b	40: 60	
151	fch9	f9	4HS	Chlorina seedling 9	44: 92	571
152	Kap1	K	4HS	Hooded lemma 1	26:179	985
155	glf1	gl, cer-zh	4HL	Glossy leaf 1	47: 81	98
156	lbi2	lb2, ert-i	4HS	Long basal rachis internode 2	44: 93	572
157	brh2	br2, ari-l	4HL	Brachytic 2	47: 84	573
158	yhd1	yh	4HL	Yellow head 1	42:250	574
160	min2	en-min		Enhancer of minute 1	26:186	266
161	min1	min	4HL	Semi-minute dwarf 1	44: 97	987
163	sgh1	sh1	4HL	Spring growth habit 1	26:188	575
164	Hln1	Hn	4HL	Hairs on lemma nerves 1	44: 99	576
165	glf3	gl3, cer-j	4HL	Glossy leaf 3	47: 86	577
166	msg25	msg,,r	4HL	Male sterile genetic 25	45:113	744
167	rym1	Ym	4HL	Reaction to barley yellow mosaic virus 1	32: 96	
168	glo-a	glo-a	4H	Globosum-a	45:115	1328
169	lgn2	lg2	4HS	Light green 2	42:264	171
170	lgn3	lg3, lg7	1HL	Light green 3	44:103	171
171	lgn4	lg4, lg1	4HL	Light green 4	44:105	681

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
172	lks5	lk5, ari-c	4HL	Short awn 5	47: 89	1297
173	blx3	bl3	4HL	Non-blue aleurone xenia 3	26:198	2506
174	blx4	bl4	4HL	Non-blue (pink) aleurone xenia 4	26:199	2507
176	ovl1	ovl	4H	Ovaryless 1	35:191	
177	fch10		4H	Chlorina seedling 10	43: 95	1737
178	int-c	i, vrs5 HvTB1	4HS	Intermedium spike-c	47: 91	776
179	Hsh1	Hs	4HL	Hairy leaf sheath 1	44:107	986
180	sid1	nls	4HL	Single internode dwarf 1	43: 97	2477
181	eam9	ea,,c	4HL	Early maturity 9	26:204	1732
182	flo-a	flo-a	6HL	Extra floret-a	45:116	1741
183	Ynd1	Yn	4HS	Yellow node 1	44:109	1607
184	Zeo3	Zeo2, cly1.c	4HL	Zeocriton 3	47: 95	1611
185	brh5	brh.m	4HS	Brachytic 5	44:110	1678
186	sld3	ant17.567	4HS	Slender dwarf 3	40: 63	2480
187	brh9	brh.k	4HL	Brachytic 9	43: 99	1676
188	Alp1	Alp, Pht	4HL	Aluminum tolerance 1	47: 98	
189	Acr2	acr, lax	4HL	Accordion rachis 2	47:101	1071
190	tfm1		1HL	Thick filament 1	40: 67	
191	fch17	vy	1H/3H	Chlorina seedling 17	40: 68	1079
193	viv-b	viv-6	4HS	Viviparoides-b	43:100	
194	sld7	sld.f	4HL	Slender dwarf 7	43:101	2481
195	sex9	sex.l	4HL	Shrunken endosperm xenia 9	43:102	2473
196	sdw7	sdw.u	4HL	Semidwarf 7	43:103	2462
197	nec34	nec.k	4HS	Necroticans 34	43:104	
198	Rpt8	QRpts4	4HS	Reaction to <i>Pyrenophora teres</i> 8	43:105	
201	fch7	f7, clo-f7	1HL	Chlorina seedling 7	41:113	4
202	trd1	t, bra-c	1HL	Third outer glume 1	46: 65	227
203	Blp1	B	1HL	Black lemma and pericarp 1	46: 67	988
207	abo1	a _t	1HL	Albino seedling 1	26:210	51
208	fst2	fs2	1HL	Fragile stem 2	41:114	578
213	Sgh3	Sh3	1HL	Spring growth habit 3	26:212	764
214	eam8	ea _k , mat-a	1HL	Early maturity 8	46: 69	765

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
215	des6	des6	1HL/ 5HL	Desynapsis 6	43:106	597
218	Rph4	Pa4	1HS	Reaction to <i>Puccinia hordei</i> 4	42:302	1314
220	fch3	f3	1HS	Chlorina seedling 3	40: 71	851
221	wst5	wst5	1HL	White streak 5	46: 73	591
222	nec1	sp,,b	1HL	Necrotic leaf spot 1	43:108	989
223	zeb3	zb3, zb _c	1HL	Zebra stripe 3	40: 72	1451
224	ert-b	ert-b	1HL	Erectoides-b	40: 74	470
225	clh1	clh	7H/5H	Curled leaf dwarf 1	40: 76	1212
226	rvl1	rvl	1HL	Revoluted leaf 1	40: 77	608
227	sls1	sls	1HL	Small lateral spikelet 1	40: 78	2492
228	Sil1	Sil	1H	Subcrown internode length 1	40: 79	1604
229	cud2	cud2	1HL	Curly dwarf 2	47:103	1712
230	glo-e	glo-e	3HL	Globosum-e	45:117	1755
231	cur5	cu5	2HS	Curly 5	41:120	1710
232	Lys4	sex5	1HS	High lysine 4	40: 80	2475
233	xnt7	xan,,g	1HL	Xantha seedling 7	26:231	581
234	mov3	mo-a	1H	Multiovary 3	32:102	
235	lel1	lel	2H	Leafy lemma 1	46:74	1780
237	Rpt2	Rpt2c	1HS	Reaction to <i>Pyrenophora teres</i> 2	43:110	
238	ari-t	ari-25	1H	Breviaristatum-t	47:104	
239	sci-b	sci-4	1H/6H	Scirpoides-b	46: 76	
240	sdw6	sdw.f	1H/7H	Semidwarf 6	46:77	2449
241	Acr3	acr	1HL	Accordionrachis 3	47:105	1071
242	sld6	sld.g	1H	Slender dwarf 6	40: 87	2482
244	dsp11	dsp.am, dsp.ao	1HL	Dense spike 11	41:121	1722 1723
251	mul2	mul2	6HL	Multiflorus 2	26:232	1394
252	eam7	HvCO7	6HS	Early maturity 7	45:118	579
253	cul2	uc2	6HL	Uniculus 2	43:112	531
254	rob1	o, rob-o	6HS	Orange lemma 1	46: 78	707
255	xnt5	x _n	6HL	Xantha seedling 5	26:237	43
257	raw5	r,,e	6HL	Smooth awn 5	44:112	785
258	dsp9	l9, ert-e	6HL	Dense spike 9	47:107	1774

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
260	fch11	f11	6HL	Chlorina seedling 11	45:120	1738
261	nec2	nec2	6HS	Necrotic leaf spot 2	26:241	1224
262	cur1	cu1	3HL	Curly 1	46: 81	1705
263	cur3	cu3	6HL	Curly 3	41:125	1707
264	mtt5	mt,,f	6HL	Mottled leaf 5	41:126	2410
265	nec3	nec3	6HS	Necrotic leaf spot 3	43:116	1330
266	ert-e	l9, dsp9	6HL	Erectoides-e	47:109	477
267	Rph11	Rph11	6HL	Reaction to <i>Puccinia hordei</i> 11	26:247	1589
268	lax-b	lax-b	6HL	Laxatum-b	44:113	1776
269	lys6	lys6	6H	High lysine 6	44:114	1786
270	abo14	alb,,q	6HL	Albino seedling 14	26:250	586
271	abo15	alb,,t	6HS	Albino seedling 15	26:251	
272	Rpt5	Pt _a	6HL	Reaction to <i>Pyrenophora teres</i> 5	43:120	
274	ari-x	ari-22	6H	Breviaristatum-x	43:124	
275	lfl1	lfl, lfs	6HL	Leafless 1	47:111	609
301	fst1	fs	5HL	Fragile stem 1	26:252	629
302	mtt2	mt2	5HL	Mottled leaf 2	41:127	1398
303	var3	va3	5HL	Variegated 3	44:115	1277
304	wst2	wst2	5HL	White streak 2	46: 83	766
305	crm1	cm	5HL	Cream seedling 1	26:256	20
306	var1	va	5HL	Variegated 1	37:259	1278
308	lbi1	lb, rac-a	5HL	Long basal rachis internode 1	43:125	580
309	Sgh2	Sh2	5HL	Spring growth habit 2	26:259	770
311	dex1	sex2	5HS	Defective endosperm xenia 1	26:260	
312	raw1	r	5HL	Smooth awn 1	46: 84	27
313	fch6	f6, yv	5HL	Chlorina seedling 6	44:116	1390
314	vrs2	v2, int-b	5HL	Six-rowed spike 2	47:113	773
315	vrs3	v3, int-a	1HL	Six-rowed spike 3	47:115	774
317	ddt1	ddt	5HS	Reaction to DDT 1	26:266	331
319	rpg4	rpg4	5HL	Reaction to <i>Puccinia graminis</i> 4	26:267	2438
320	int-b	vrs2	5HL	Intermedium spike-b	47:118	1764
321	srh1	s, l	5HL	Short rachilla hair 1	46: 86	27
322	dsk1	dsk	5HL	Dusky 1	41:128	1714
323	nld1	nld	5HL	Narrow leafed dwarf 1	46: 88	769

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
324	cud1	cud	5HL	Curly dwarf 1	26:272	
325	crl1	crl, cl	6H	Curly lateral 1	41:129	
326	blf1	bb	5HL	Broad leaf 1	46: 90	
327	flo-b	flo-a	6HL	Extra floret-b	45:121	
328	ari-e	lk9, GPert	5HL	Breviaristatum-e	47:121	
329	ari-h	lk11	5HL	Breviaristatum-h	26:277	
330	ert-g	ert-g, br3	1HL	Erectoides-g	41:133	
331	ert-n	ert-n	5HL	Erectoides-n	44:120	488
332	Ert-r	Zeo1	2HL	Erectoides-r	47:123	492
333	Rph12	Pa12, Pa9	5HL	Reaction to <i>Puccinia hordei</i> 12	26:281	1590
334	raw6	r6	5HL	Smooth awn 6	26:282	2437
335	msg49	msg,jw	5HL	Male sterile genetic 49	45:122	2402
337	blf2	bb2, nlh	5HL	Broad leaf 2	41:137	1667
338	lys1	lys	5HL	High lysine 1	26:286	1784
339	lys3	sex3	5HL	High lysine 3	43:127	1785
340	raw2	r2	5HL	Smooth awn 2	46: 92	27
341	abo12	alb,,o	5HS	Albino seedling 12	26:290	583
342	glo-f	glo-e	5HL	Globosum-f	26:291	
343	Lfb1	Lfb	5HL	Leafy bract 1	41:140	1577
344	var2	va2	5HL	Variegated 2	32:104	2496
345	rym3	ym3	5HS	Reaction to barley yellow mosaic virus 3	32:105	
346	yst5	yst5	7HS	Yellow streak 5	43:130	2501
347	mnd4	m4	5HL	Many noded dwarf 4	44:122	1798
348	Eam5	HvPhyC-e	5HL	Early maturity 5	45:123	
349	brh4	brh.j	2HL	Brachytic 4	42:407	1675
350	brh6	brh.s	5HS	Brachytic 6	42:408	1683
351	gsh1	gs1, cer-q	2HS	Glossy sheath 1	46: 94	735
352	gsh2	gs2, cer-b	3HL	Glossy sheath 2	47:125	736
353	gsh3	gs3, cer-a	7HS	Glossy sheath 3	41:143	737
354	gsh4	gs4, cer-x	6HL	Glossy sheath 4	41:146	738
355	gsh5	gs5, cer-s	2HL	Glossy sheath 5	47:128	739
356	gsh6	gs6, cer-c	2HS	Glossy sheath 6	46:101	740
357	msg1	ms1	1HL	Male sterile genetic 1	45c:126	1810

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
358	msg2	ms2	2HL	Male sterile genetic 2	45:128	2371
359	msg3	ms3	2HS	Male sterile genetic 3	45:130	1130
360	msg4	ms4	1H	Male sterile genetic 4	45:132	2392
361	msg5	ms5	3HS	Male sterile genetic 5	45:133	2403
362	msg6	ms6	6HS	Male sterile genetic 6	45:135	2405
363	msg7	ms7	5HL	Male sterile genetic 7	45:137	2406
364	msg8	ms8	5HL	Male sterile genetic 8	45:139	2407
365	msg9	ms9	2HS	Male sterile genetic 9	45:141	2408
366	msg10	ms10	7HS	Male sterile genetic 10	45:142	1811
367	msg11	ms11	5HS	Male sterile genetic 11	45:144	1812
368	msg13	ms13	3HL	Male sterile genetic 13	45:146	1813
369	msg14	ms14	7HS	Male sterile genetic 14	45:147	1814
370	msg15	ms15		Male sterile genetic 15	45:149	1815
371	msg16	ms16	5HS	Male sterile genetic 16	45:150	1816
372	msg17	ms17	5HL	Male sterile genetic 17	45:152	1817
373	msg18	ms18	5HL	Male sterile genetic 18	45:153	1818
374	msg19	ms19	5HS	Male sterile genetic 19	45:155	1819
375	msg20	Msg,,ad	4H	Male sterile genetic 20	45:156	2372
376	msg21	ms21	1HL	Male sterile genetic 21	45:157	2373
377	seg1	se1	7HL	Shrunken endosperm genetic 1	45:158	750
378	seg2	se2	7HS	Shrunken endosperm genetic 2	26:326	751
379	seg3	se3, ant17	3H	Shrunken endosperm genetic 3	45:160	752
380	seg4	se4	7HL	Shrunken endosperm genetic 4	37:267	753
381	seg5	se5	7HS	Shrunken endosperm genetic 5	26:329	754
382	sex1	lys5	6HL	Shrunken endosperm xenia 1	26:330	755
383	msg22	ms22	7H	Male sterile genetic 22	45:162	741, 2374
384	msg23	ms23	7HL	Male sterile genetic 23	45:163	2375
385	msg24	ms24	4HL	Male sterile genetic 24	45:164	2376
386	des3	des3	2H/ 5HL	Desynapsis 3	43:140	594
387	des8	des8	3H	Desynapsis 8	41:151	599
388	des9	des9	7HL	Desynapsis 9	44:131	600
389	des10	des,,p	5HL	Desynapsis 10	41:152	601

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
390	des11	des11	3HL	Desynapsis 11	44:132	602
391	des12	des12	3H	Desynapsis 12	44:133	603
392	des13	des13	3H	Desynapsis 13	44:134	604
393	des14	des14	7H	Desynapsis 14	44:135	605
394	des15	des15	3HL	Desynapsis 15	44:136	606
395	msg26	msg,,u	7HS	Male sterile genetic 26	45:166	745
396	seg6	se6	3HL	Shrunkend endosperm genetic 6	44:138	2467
397	seg7	se7		Shrunkend endosperm genetic 7	37:269	2468
399	cer-d	cer-d	5HL	Eceriferum-d	41:153	425
400	cer-e	cer-e	1HL	Eceriferum-e	40:102	1518
401	cer-f	cer-f	1H	Eceriferum-f	40:104	427
402	cer-g	cer-g	2HL	Eceriferum-g	47:131	428
403	cer-h	cer-h	4HS	Eceriferum-h	41:157	429
404	cer-i	cer-i	5HL	Eceriferum-i	41:158	430
405	cer-k	cer-k	4HL	Eceriferum-k	41:160	432
406	cer-l	cer-l	3HL	Eceriferum-l	44:142	433
407	cer-m	cer-m	1HL/ 3HL	Eceriferum-m	41:161	434
408	cer-n	gs9	2HL	Eceriferum-n	44:143	435
409	cer-o	cer-o	1HL	Eceriferum-o	40:106	436
410	cer-p	cer-p	7HL	Eceriferum-p	41:162	437
411	cer-r	cer-r	3HL	Eceriferum-r	45:168	439
413	gsh8	cer-u, gs8	2HS	Glossy sheath 8	46:105	442
414	cer-v	cer-v	2HS	Eceriferum-v	44:147	443
415	cer-w	cer-w	5HL	Eceriferum-w	41:166	1519
417	cer-y	cer-y	1HS	Eceriferum-y	44:149	446
418	cer-z	cer-z	7HS	Eceriferum-z	44:150	447
419	cer-za	cer-za	5HL	Eceriferum-za	43:144	1521
420	cer-zb	cer-zb	5HS	Eceriferum-zb	42:508	1522
421	cer-zc	cer-zc	4HL/ 2HS	Eceriferum-zc	42:510	450
422	cer-zd	cer-zd	3HL	Eceriferum-zd	40:110	451
423	cer-ze	gl5	7HS	Eceriferum-ze	44:152	452

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
424	cer-zf	cer-zf	3H/7HS	Eceriferum-zf	42:516	453
425	cer-zg	cer-zg	4HL	Eceriferum-zg	26:377	454
427	cer-zi	cer-zi	1HL	Eceriferum-zi	41:168	456
428	cer-zj	cer-zj	5HL	Eceriferum-zj	42:520	457
429	cer-zk	cer-zk	2H	Eceriferum-zk	43:146	458
430	cer-zl	cer-zl		Eceriferum-zl	26:382	459
431	cer-zn	cer-zn	1H	Eceriferum-zn	40:112	1523
432	cer-zo	cer-zo	3HS	Eceriferum-zo	44:154	462
433	cer-zp	cer-zp	5HL	Eceriferum-zp	26:385	463
434	cer-zq	cer-zq		Eceriferum-zq	26:386	1524
435	cer-zr	cer-zr	5HL	Eceriferum-zr	44:155	1525
436	cer-zs	cer-zs		Eceriferum-zs	44:156	1526
437	cer-zt	cer-zt	2HS	Eceriferum-zt	44:157	1527
438	cer-zu	cer-zu	1HS	Eceriferum-zu	41:170	1528
439	cer-zv	cer-zv	4HL	Eceriferum-zv	47:133	1529
440	cer-zw	cer-zw		Eceriferum-zw	26:392	1530
441	cer-zx	cer-zx	3H	Eceriferum-zx	46:109	1531
442	cer-zy	cer-zy	1HS	Eceriferum-zy	40:116	1532
443	cer-zz	cer-zz	3HL	Eceriferum-zz	44:159	1533
444	cer-ya	cer-ya	3HS	Eceriferum-ya	26:396	1534
445	cer-yb	cer-yb	2HL	Eceriferum-yb	41:171	1535
446	cer-yc	cer-yc	6H/7HS	Eceriferum-yc	41:172	1536
447	cer-yd	cer-yd	3HS	Eceriferum-yd	26:399	1537
448	cer-ye	cer-ye	4H	Eceriferum-ye	43:149	1538
449	cer-yf	cer-yf	7H	Eceriferum-yf	44:160	1539
450	cer-yg	cer-yg	7HS	Eceriferum-yg	44:161	1540
451	cer-yh	cer-yh	3HS	Eceriferum-yh	26:403	1541
453	fer1			Few roots 1	46:112	2538
454	blx5	bl5	7HL	Non-blue aleurone xenia 5	26:404	2509
455	seg8	seg8	7H	Shrunken endosperm genetic 8	45:170	2469
460	cur4	cu4, glo-d	2HL	Curly 4	45:172	1708
461	zeb2	zb2, fch10	4HS	Zebra stripe 2	43:152	93
462	yst3	yst,,c	3HS	Yellow streak 3	44:163	48

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
463	gig1	gig, sf	2HL	Gigas 1	44:164	1650
464	msg27	msg,,ae	2HS	Male sterile genetic 27	45:174	2379
465	msg28	msg,,as	2HS	Male sterile genetic 28	45:175	2380
466	msg29	msg,,a	5HL	Male sterile genetic 29	45:176	2381
467	msg30	msg,,c	7HL	Male sterile genetic 30	45:177	2382
468	msg31	msg,,d	1HL	Male sterile genetic 31	45:178	2383
469	msg32	msg,,w	7H	Male sterile genetic 32	45:179	2384
470	msg33	msg,,x	2HS	Male sterile genetic 33	45:180	2385
471	msg34	msg,,av	6HS/ 7HS	Male sterile genetic 34	45:181	2386
472	abr1	abr	2HL	Accordion basal rachis internode 1	26:419	1563
473	com1	lax-270, bir1	5HL	Compositum 1	47:135	
474	lax-a	lax-a	5HL	Laxatum-a	46:113	
475	lax-c	lax-21	6HL	Laxatum-c	47:137	
476	lax-d	lax-83		Laxatum-d	47:139	
477	lax-f	lax-61		Laxatum-f	47:140	
478	lax-g	lax-25		Laxatum-g	47:141	
479	lax-h	lax-26	3H	Laxatum-h	47:142	
480	lax-i	lax-50		Laxatum-i	47:144	
481	lax-j	lax-49	2H	Laxatum-j	47:145	
482	lax-k	lax-84	2H/3H	Laxatum-k	47:147	
483	lax-l	lax-60	1H	Laxatum-l	47:148	
484	lax-m	lax-80		Laxatum-m	47:149	
485	lax-n	lax-67	2H/4HL /5H	Laxatum-n	47:150	
486	lax-o	lax-79		Laxatum-o	47:152	
487	lax-df	lax-204		Laxatum-df	47:153	
488	lax-ef	lax-225	2H	Laxatum-ef	47:154	
489	lax-ff	lax-216		Laxatum-ff	47:156	
490	lax-gf	lax-217		Laxatum-gf	47:157	
491	lax-hf	lax-244	2H/3HL /5H	Laxatum-hf	47:158	
492	lax-if	lax-246		Laxatum-if	47:160	
493	Lax-jf	lax-253	4HS	Laxatum-jf	47:161	

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
494	lax-kf	lax-295		Laxatum-kf	47:163	
495	lax-lf	lax-274		Laxatum-lf	47:164	
496	lax-mf	lax-302		Laxatum-mf	47:165	
497	lax-nf	lax-322	2HL/4H	Laxatum-nf	47:166	
498	msg35	msg.,dr	2HL	Male sterile genetic 35	45:183	2387
499	msg36	msg.,bk	6HS	Male sterile genetic 36	45:184	2388
500	msg37	msg.,hl	3HL	Male sterile genetic 37	45:186	2389
501	msg38	msg.,jl	3H	Male sterile genetic 38	45:187	2390
502	msg39	msg.,dm	3H	Male sterile genetic 39	45:188	2391
503	msg40	msg.,ac	6HL	Male sterile genetic 40	45:190	2393
504	msg41	msg.,aj	6HS	Male sterile genetic 41	45:191	
505	msg42	msg.,db	3H	Male sterile genetic 42	45:193	
506	msg43	msg.,br	2HL	Male sterile genetic 43	45:194	
507	msg44	msg.,cx	5HL	Male sterile genetic 44	45:195	
508	msg45	msg.,dp	5HL/ 7HS	Male sterile genetic 45	45:196	
509	msg46	msg.,ec	2H/6H	Male sterile genetic 46	45:197	
510	msg47	msg.,ep	3HS/ 7HS	Male sterile genetic 47	45:198	
511	Rpg1	T	7HS	Reaction to <i>Puccinia graminis</i> 1	26:437	
512	Rpg2	T2		Reaction to <i>Puccinia graminis</i> 2	26:439	
513	xnt2	x _b		Xantha seedling 2	26:440	
515	Rsp1	Sep		Reaction to <i>Septoria passerinii</i> 1	26:441	2510
516	Rsp2	Sep2		Reaction to <i>Septoria passerinii</i> 2	37:275	2511
517	Rsp3	Sep3		Reaction to <i>Septoria passerinii</i> 3	37:276	2512
518	sdw1	denso, Hv20ox ₂	3HL	Semidwarf 1	47:168	2513
519	mnd1	m	2H	Many-noded dwarf 1	43:154	253
520	msg48	msg.,jt	1H	Male sterile genetic 48	45:199	2401
521	mtt1	mt. mt3	1HL	Mottled leaf 1	41:179	622
522	cer-yi	cer-yi	2H	Eceriferum-yi	41:180	1542
523	cer-yj	cer-yj	1HS	Eceriferum-yj	40:124	1543
524	cer-yk	cer-yk	7HL	Eceriferum-yk	44:167	1544
525	cer-yl	cer-yl	4HL	Eceriferum-yl	47:171	1545
526	cer-ym	cer-ym	4HL	Eceriferum-ym	47:173	1546

Table 2. (continued)						
BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
527	cer-yn	cer-yn	1H	Eceriferum-yn	40:125	1547
528	cer-yo	cer-yo	4HS	Eceriferum-yo	44:168	1548
529	cer-yp	cer-yp	5HS	Eceriferum-yp	44:169	1549
530	cer-yq	cer-yq	5H	Eceriferum-yq	44:170	1550
531	cer-yr	cer-yr	5HL	Eceriferum-yr	44:171	1551
532	cer-ys	cer-ys	2HL	Eceriferum-ys	44:172	1552
533	cer-yt	cer-yt	1H/5H	Eceriferum-yt	40:126	1553
534	cer-yu	cer-yu	1H	Eceriferum-yu	40:127	1554
535	cer-yx	cer-yx	1H/3H /5H	Eceriferum-yx	40:128	1555
536	Cer-yy	Gle1	1HS	Eceriferum-yy	40:129	1556
537	cer-yz	cer-yz	1H/5H	Eceriferum-yz	44:173	1557
538	cer-xa	cer-xa	2HL/4H /5HL	Eceriferum-xa	44:174	1558
539	cer-xb	cer-xb	4H	Eceriferum-xb	44:175	1559
540	cer-xc	cer-xc	1H	Eceriferum-xc	44:176	1560
541	cer-xd	cer-xd	4H/5HL	Eceriferum-xd	44:177	1561
542	Dwf2	Dwf2		Dominant dwarf 2	24:170	
544	int-h	int-h	5H	Intermedium spike-h	44:179	1768
545	int-i	int-i	2HS	Intermedium spike-i	41:181	1769
546	int-k	int-k	7H	Intermedium spike-k	44:180	1770
547	int-m	int-m	5HL	Intermedium spike-m	44:181	1772
548	Fol-b	Ang	1HS	Angustifolium-b	40:131	17
549	Lga1	Log	7HS	Long glume awn 1	44:183	835
550	ari-b	ari-b		Breviaristatum-b	44:185	1649
551	ari-f	ari-21	7H	Breviaristatum-f	41:182	1654
552	ari-j	ari-32		Breviaristatum-j	44:186	1658
553	ari-k	ari-504	3H	Breviaristatum-k	44:187	1659
554	ari-m	brh1	7HS	Breviaristatum-m	47:175	1661
555	ari-n	ari-45	7H	Breviaristatum-n	41:185	1662
556	ari-o	ert-u, ert-zd	7HL	Breviaristatum-o	45:200	1663
557	ari-p	ari-27		Breviaristatum-p	46:120	1664
558	ari-q	ari-271	4H	Breviaristatum-q	44:188	1665
Table 2. (continued)						

BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
559	ari-r	ari-14	5H	Breviaristatum-r	41:187	1666
560	ert-f	ert-f	1H	Erectoides-f	40:133	478
561	ert-h	ert-h	5HL	Erectoides-h	44:189	481
562	ert-k	ert-k	6H	Erectoides-k	46:121	485
563	ert-l	ert-l		Erectoides-l	26:489	486
564	ert-p	ert-p		Erectoides-p	26:490	490
565	ert-s	ert-s		Erectoides-s	26:491	493
566	ert-t	brh3	2HS	Erectoides-t	45:203	494
567	ert-v	ert-v	6H	Erectoides-v	41:188	497
568	ert-x	ert-x	1H/7H	Erectoides-x	40:136	498
569	ert-y	ert-y		Erectoides-y	26:495	499
570	ert-z	ert-z		Erectoides-z	26:496	500
571	ert-za	ert-za	5H	Erectoides-za	44:190	501
572	ert-zb	ert-zb	7HL	Erectoides-zb	45:205	502
573	ert-zc	ert-zc	7HS	Erectoides-zc	45:206	503
574	ert-ze	ert-ze	5HS	Erectoides-ze	45:207	505
575	Rph6	Pa6		Reaction to <i>Puccinia hordei</i> 6	26:501	1598
576	Rph8	Pa8		Reaction to <i>Puccinia hordei</i> 8	26:502	1600
577	Rsg2	Rsg2		Reaction to <i>Schizaphis graminum</i> 2	37:283	2513
578	mat-b	mat-b	7HL	Praematurum-b	46:123	1788
579	mat-c	Eam6, HvCEN	2H	Praematurum-c	46:125	1789
580	mat-d	mat-d	4HL/ 6HL	Praematurum-d	45:208	1790
581	mat-e	mat-e		Praematurum-e	26:508	1791
582	mat-f	mat-f	1H	Praematurum-f	45:210	1792
583	mat-g	mat-g		Praematurum-g	26:510	1793
584	mat-h	mat-h	4HL	Praematurum-h	45:212	1794
585	mat-i	mat-i	7HL	Praematurum-i	45:214	1795
586	bra-d	bra-d	1HL	Bracteatum-d	40:139	1696
587	abo3	a2, alb-za		Albino seedling 3	26:514	165
588	abo10	a12		Albino seedling 10	26:515	57
589	abo11	a13, alb ^t		Albino seedling 11	26:516	233
590	Rph13	Rph13		Reaction to <i>Puccinia hordei</i> 13	28: 31	1591
Table 2. (continued)						

BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
591	Rph14	Rph14		Reaction to <i>Puccinia hordei</i> 14	28: 32	1592
592	yhd2	yh2		Yellow head 2	45:215	757
593	adp1	adp	3HL	Awned palea 1	43:158	1618
594	ant3	rub		Anthocyanin-deficient 3	29: 82	1641
595	ant4	ant4	4H	Anthocyanin-deficient 4	45:216	1642
596	ant5	rs2		Anthocyanin-deficient 5	29: 84	1643
597	ant6	ant6		Anthocyanin-deficient 6	29: 85	1644
598	ant13	ant13	6HL	Proanthocyanin-free 13	29: 86	1624
599	ant17	ant17	3HS	Proanthocyanin-free 17	46:128	1628
600	ant18	ant18		Proanthocyanin-free 18	45:221	1630
601	ant19	ant19		Proanthocyanin-free 19	29: 92	1631
602	ant20	ant20		Anthocyanin-rich 20	29: 93	1633
603	ant21	ant21	6H	Proanthocyanin-free 21	29: 94	1634
604	ant22	ant22	2HL	Proanthocyanin-free 22	41:191	1635
605	ant25	ant25		Proanthocyanin-free 25	29: 96	1638
606	ant26	ant26		Proanthocyanin-free 26	29: 97	1639
607	ant27	ant27		Proanthocyanin-free 27	29: 98	1640
608	ant28	ant28	3HL	Proanthocyanin-free 28	46:131	
609	ant29	ant29		Proanthocyanin-free 29	29:100	
610	ant30	ant30		Proanthocyanin-free 30	29:101	
611	Nec6	Sp	7HS	Necrotic leaf spot 6	43:159	977
612	gig2	gig2	4HL	Gigas 2	44:191	1750
613	brc1	brc-5, com2	2HS	Branched 1	45:224	
614	Zeo2	cly1.b, HvAP2	2HL	Zeocriton 2	47:177	637
615	wxs1	wxs1	7H/ 2HL	Waxy spike 1	43:160	3649
616	cul3	cul3	3HL	Uniculme 3	43:161	2494
617	cul4	uc-5, uc-3	3HL	Uniculme 4	46:132	2495
618	mnd3	mn3, m3	4HS	Many noded dwarf 3	44:194	1797
619	bra-a	bra-a	7H	Bracteatum-a	44:196	1693
620	cal-b	cal-b	5HL	Calcaroides-b	44:197	1697
621	Cal-c	Cal-c	5HL	Calcaroides-c	41:195	1567

Table 2. (continued)

BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
622	cal-e	cal-23	5HS	Calcaroides-e	32:123	
623	eli-a	lig-a		Eligulum-a	44:199	3647
624	ops1	op-3	7HS	Opposite spikelets 1	45:226	2427
625	sci-a	sci-3	5H	Scirpoides 1	44:200	
626	scl-a	scl-6	1HL	Scirpoides leaf-a	44:201	
627	viv-a	viv-5	2H	Viviparoides-a	45:227	2498
628	sex7	sex.i	5HL	Shrunken endosperm 7	32:129	2470
629	mtt6	mtt6	7HS	Mottled leaf 6	45:228	2411
630	Ari-s	ari-265	5H/7H	Breviaristatum-s	41:197	
631	brh3	brh.g, ert-t	2HS	Brachytic 3	45:229	1672
632	mnd5	mnd5	7HL	Many noded dwarf 5	44:202	
633	mnd6	den-6	5HL	Many noded dwarf 6	44:203	1713
634	pmr2	nec-50		Premature ripe 2	32:135	2421
635	nec7	nec-45	1H/6H /7H	Necroticans 7	43:166	2420
636	tst2		4HL	Tip sterile 2	43:167	1781
637	nar1	nar1	6HS	NADH nitrate reductase-deficient 1	35:194	2431
638	nar2	nar2	5HL	NADH nitrate reductase-deficient 2	35:195	2415
639	nar3	nar3	7HS	NADH nitrate reductase-deficient 3	35:196	2416
640	nar4	nar4	2Hl	NADH nitrate reductase-deficient 4	35:197	
641	nar5	nar5	5HL	NADH nitrate reductase-deficient 5	35:198	2417
642	nar6	nar6	2HL	NADH nitrate reductase-deficient 6	35:199	
643	nar7	nar7	6HL	NADH nitrate reductase-deficient 7	35:200	2418
644	nar8	nar8	6HS	NADH nitrate reductase-deficient 8	35:201	
645	bsp1			Bushy spike 1	43:168	3652
646	ovl2	ovl2		Ovaryless 2	43:169	3655
647	tst1	tst1	6HL	Tip sterile 1	43:170	3644
648	mov4	mo8		Multiovary 4	43:171	3643
649	asp1	asp1		Aborted spike 1	43:172	3654
650	sun1	sun1		Sensitivity to <i>Ustilago nuda</i> 1	43:173	3650
651	lam1	lam1		Late maturity 1	43:174	3653
652	ylf1	ylf1	7HS	Yellow leaf 1	43:175	
653	brh10	brh.l	2HS	Brachytic 10	45:231	1677
654	brh11	brh.n	5HS	Brachytic 11	45:232	1679

Table 2. (continued)

BGS no.	Locus symbol*		Chr. loc.†	Locus name or phenotype	Descr. vol. p.	GSHO no.‡
	Rec.	Prev.				
655	brh12	brh.o	5HS	Brachytic 12	45:233	1680
656	brh13	brh.p	5HS	Brachytic 13	45:234	1681
657	brh15	brh.u	2HL	Brachytic 15	44:205	1685
658	brh17	brh.ab	5HS	Brachytic 17	45:236	1669
659	brh18	brh13	5HS	Brachytic 18	45:237	1670
660	nld2		5H/6H /7H	Narrow leafed dwarf 2	43:176	3645
661	dub1		5HL	Double seed 1	37:301	
663	sdw3	gai	2HS	Semidwarf 3	46:134	
667	Rpt1	Pt	3HL	Reaction to <i>Pyrenophora teres</i> 1	43:177	
671	nec8	nec.w	5HL	Necrotic leaf spot 8	43:179	3600
672	nec9		3HL	Necrotic leaf spot 9	43:181	3599
673	est1	cs	5HL	Corn stalk	41:199	
674	mtt8	Mut 1661		Mottled leaf 8	43:182	3597
675	mtt9	Mut 2721		Mottled leaf 9	44:207	3598
676	fch16	clo.117	2HS	Chlorina seedling 16	40:144	
677	mtt7	mtt.h	2HS	Mottled leaf 7	42:753	
678	ari-u	ari-245	2HS	Breviaristatum-u	45:239	
679	acr4	acr-3	2H/6HL	Accordion rachis 4	47:181	
680	ari-v	ari-137	5HS	Breviaristatum-v	41:202	
681	nec10	necS 1-1	3H	Necroticans 10	43:184	3607
682	nec11		1H	Necroticans 11	43:185	3610
683	nec12			Necroticans 12	43:186	3613
684	nec13			Necroticans 13	43:187	3616
685	nec14			Necroticans 14	43:188	3619
686	nec15			Necroticans 15	43:189	3620
687	nec16			Necroticans 16	43:190	3621
688	nec17			Necroticans 17	43:191	3622
689	nec18			Necroticans 18	43:192	3623
690	nec19			Necroticans 19	43:193	3624
691	nec20			Necroticans 20	43:194	3625
692	nec21			Necroticans 21	43:195	3626
693	Nec22			Necroticans 22	43:196	3627
694	nec23			Necroticans 23	43:197	3628
Table 2. (continued)						

BGS no.	Locus symbol		Chr. loc. [†]	Locus name or phenotype	Descr. vol. p.	GSHO no. [‡]
	Rec.	Prev.				
695	Nec24			Necroticans 24	43:198	3629
696	nec25			Necroticans 25	43:199	3630
697	Nec26			Necroticans 26	43:200	3631
698	nec27			Necroticans 27	43:201	3633
699	nec28			Necroticans 28	43:202	3635
700	nec29			Necroticans 29	43:203	3636
701	nec30			Necroticans 30	43:204	3637
702	nec31			Necroticans 31	43:205	3638
703	nec32			Necroticans 32	43:206	3639
704	nec33			Necroticans 33	43:207	3640
707	Rpr1		4H	Required for <i>Puccinia graminis</i> resistance 1	42:757	
711	Rpt3	QRpts2	2HS	Reaction to <i>Pyrenophora teres</i> 3	43:208	
713	Rpt6		5HL	Reaction to <i>Pyrenophora teres</i> 6	43:210	
714	Rpt7	Qrpts4	4HL	Reaction to <i>Pyrenophora teres</i> 7	43:211	
716	ibl1	ibl1	5HL	Intense blue aleurone 1	46:136	
718	ops2	op-2	5HL	Opposite spikelets 2	43:213	2426
719	ops3	op-1	5HS	Opposite spikelets 3	43:214	2425
720	viv-c	viv-1	5H	Viviparoides-c	43:215	2497
721	ari-w	ari-153	7H	Breviaristatum-w	43:216	
722	ari-y	ari-9	5H	Breviaristatum-y	43:217	
723	mov5	mov.o		Multiovary 5	43:218	3671
724	lks6	lks.q	1H/5H /6H	Short awn 6	43:219	3674
725	ovl3			Ovaryless 3	43:220	3687
726	mnd7			Many noded dwarf 7	43:221	3686
727	ubs5			Unbranched style 5	43:222	3675
728	fxp1			Fenoxaprop-p-ethyl reaction 1	43:223	3684
729	dsk2	msg.,df	7HL	Dusky 2	44:208	
730	lab1		5HL	Labile1	46:137	
731	rpr2	γ08-118; R43-22#1	6H	Required for <i>Puccinia graminis</i> resistance 2	46:139	
732	rpr3	γ08-112; R12-31#3		Required for <i>Puccinia graminis</i> resistance 3	46:141	3696

Table 2: continued

BGS no.	Locus symbol		Chr. loc. [†]	Locus name or phenotype	Descr. vol. p.	GSHO no. [‡]
733	rpr4	γ08-114; R36-37#1		Required for <i>Puccinia graminis</i> resistance 4	46:142	3697
734	rpr5	γ08-117; R42-33#5		Required for <i>Puccinia graminis</i> resistance 5	46:143	3699
735	rpr6	γ08-119; R47-23#1		Required for <i>Puccinia graminis</i> resistance 6	46:144	3700
736	rpr7	γ08-115; R3-18#3		Required for <i>Puccinia graminis</i> resistance 7	46:145	3701
737	rcr1	γ08-122; (R4-29)		Required for resistance to <i>Cochliobolus sativus</i> 1	45:249	3703
738	rcr2	γ08-123; (R4-40)		Required for resistance to <i>Cochliobolus sativus</i> 2	45:250	3704
739	rcr3	γ08-124		Required for resistance to <i>Cochliobolus sativus</i> 3	45:251	3705
740	Pbg1	Pbg	7H	Pubescence on glume 1	47:182	
741	stb1	stb		Stubble1	47:183	
743	twk1	tw	7H	Tweaky spike 1	47:185	
744	acr5	acr-2	7H	Accordion rachis 5	47:187	

* Recommended locus symbols are based on utilization of a three-letter code for barley genes as approved at the business meeting of the Seventh International Barley Genetics Symposium at Saskatoon, Saskatchewan, Canada, on 05 August 5 1996.

† Chromosome numbers and arm designations are based on a resolution passed at the business meeting of the Seventh International Barley Genetics Symposium at Saskatoon, Saskatchewan, Canada, on August 05 1996. The Burnham and Hagberg (1956) designations of barley chromosomes were 1 2 3 4 5 6 and 7 while new designations based on the Triticeae system are 7H 2H 3H 4H 1H 6H and 5H, respectively.

‡ The seed stock associated with each BGS number is held as a GSHO stock number in the Barley Genetics Stock Collection at the USDA-ARS National Small Grains Germplasm Research Facility, Aberdeen, ID 83210, USA.

Table 3. An alphabetic listing of recently published Barley Genetic Stock (BGS) descriptions for loci in barley (*Hordeum vulgare*), including information on recommended locus names, recommended locus symbols, chromosomal locations, description citation, and original cultivars.

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr. vol. p.	Parental cultivar
Rec.	Prev.					
abo1	a _t	207	1HL	Albino seedling 1	26:210	Trebi
abo2	a2	53	2HS	Albino seedling 2	26: 89	Nilsson-Ehle No 2
abo3	alb-za	587		Albino seedling 3	26:514	Unknown cultivar
abo4	a4	94	2H	Albino seedling 4	26:133	Unknown cultivar
abo6	a _c	106	3HS	Albino seedling 6	46: 60	Colsess
abo8	a _{c2}	4	7HS	Albino seedling 8	26: 47	Coast
abo9	a _n	112	3HS	Albino seedling 9	26:146	Nigrinudum
abo10	a _{t2}	588		Albino seedling 10	26:515	Canadian Thorpe
abo11	a _{t3}	589		Albino seedling 11	26:516	Trebi
abo12	alb,,o	341	5HS	Albino seedling 12	26:290	Titan
abo13	alb,,p	95	2HL	Albino seedling 13	26:134	Titan
abo14	alb,,q	270	6HL	Albino seedling 14	26:250	Shabet
abo15	alb,,t	271	6HS	Albino seedling 15	26:251	Betzes
abr1	abr	472	2HL	Accordion basal rachis internode 1	26:419	Bonus
acr1	acr, ril	97	2HL	Accordion rachis 1	47: 68	ACBV89B229
Acr2	acr, lax	189	4HL	Accordion rachis 2	47:101	Clho 6164
Acr3	acr	241	1HL	Accordion rachis 3	47:105	Burma Girl
acr4	acr-3	679	2H/ 6HL	Accordion rachis 4	47:181	Bonus
acr5	acr-2	744	7H	Accordion rachis 5	47:187	
adp1	adp	593	3HL	Awed palea 1	43:158	Unknown line
alm1	al	108	3HS	Albino lemma 1	46: 61	Russia 82
Alp1	Alp, Pht	188	4HL	Aluminum tolerance 1	47: 98	
als1	als	101	3HL	Absent lower laterals 1	43: 74	Montcalm
ant1	rs	33	7HS	Anthocyanin-less 1	46: 49	Bonus
ant2	pr	80	2HL	Anthocyanin-less 2	46: 55	Foma
ant3		594		Anthocyanin-deficient 3	29: 82	Bonus
ant4		595	4H	Anthocyanin-deficient 4	45:216	Foma
ant5		596		Anthocyanin-deficient 5	29: 84	Bonus
ant6		597		Anthocyanin-deficient 6	29: 85	Foma
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr. vol. p.	Parental cultivar
Rec.	Prev.					
ant13		598	6HL	Proanthocyanidin-free 13	29: 86	Foma
ant17		599	3HS	Proanthocyanidin-free 17	46:128	Nordal
ant18		600	3H	Proanthocyanidin-free 18	45:221	Nordal
ant19		601		Proanthocyanidin-free 19	29: 92	Alf
ant20		602		Anthocyanidin-rich 20	29: 93	Foma
ant21		603	6H	Proanthocyanidin-free 21	29: 94	Georgie
ant22		604	2HL	Proanthocyanidin-free 22	41:191	Hege 802
ant25		605		Proanthocyanidin-free 25	29: 96	Secobra 18193
ant26		606		Proanthocyanidin-free 26	29: 97	Grit
ant27		607		Proanthocyanidin-free 27	29: 98	Zebit
ant28		608	3HL	Proanthocyanidin-free 28	46:131	Grit
ant29		609		Proanthocyanidin-free 29	29:100	Ca 708912
ant30		610		Proanthocyanidin-free 30	29:101	Gunhild
ari-a	ari-1	132	3HS	Breviaristatum-a	41:106	Bonus
ari-b	ari-11	550		Breviaristatum-b	44:185	Bonus
ari-e	lk9, GPert	328	5HL	Breviaristatum-e	47:121	Bonus
ari-f	ari-21	551	7H	Breviaristatum-f	41:182	Bonus
ari-g	lk10	89	2H	Breviaristatum-g	47: 66	Bonus
ari-h	lk11	329	5HL	Breviaristatum-h	26:277	Foma
ari-j	ari-32	552		Breviaristatum-j	44:186	Bonus
ari-k	ari-504	553	3H	Breviaristatum-k	44:187	Bonus
ari-m	brh1	554	7HS	Breviaristatum-m	47:175	Bonus
ari-n	ari-45	555	7H	Breviaristatum-n	41:185	Bonus
ari-o	ert-u, ert-zd	556	7HL	Breviaristatum-o	45:200	Bonus
ari-p	ari-27	557		Breviaristatum-p	46:120	Foma
ari-q	ari-271	558	4H	Breviaristatum-q	44:188	Kristina
ari-r	ari-14	559	5H	Breviaristatum-r	41:187	Bonus
Ari-s	ari-265	630	5H/7H	Breviaristatum-s	41:197	Kristina
ari-t	ari-25	238	1H	Breviaristatum-t	47:104	Bonus
ari-u	ert-t	678	2HS	Breviaristatum-u	45:100	Foma
ari-v	ari-137	680	5HS	Breviaristatum-v	41:202	Foma
ari-w	ari-153	721	7H	Breviaristatum-w	43:216	Foma
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. Loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
ari-x	ari-22	274	6H	Breviaristatum-x	43:124	Bonus
ari-y	ari-9	722	5H	Breviaristatum-y	43:217	Bonus
asp1		649		Aborted spike 1	43:172	Steptoe
blf1	bb	326	5HL	Broad leaf 1	46: 90	Bonus
blf2	bb2, nlh	337	5HL	Broad leaf 2	41:137	Hannchen
Blp1	B	203	1HL	Black lemma and pericarp 1	46: 67	Nigrinudum
blx1	bl	15	4HL	Non-blue aleurone xenia 1	26: 60	Goldfoil
blx2	bl2	19	7HS	Non-blue aleurone xenia 2	26: 65	Nepal
blx3	bl3	173	4HL	Non-blue aleurone xenia 3	26:198	Blx
blx4	bl4	174	4HL	Non-blue (pink) aleurone xenia 4	26:199	Ab 6
blx5	bl5	454	7HL	Non-blue aleurone xenia 5	26:404	BGM 122
bra-a		619	7H	Bracteatum-a	44:196	Bonus
bra-d		586	1HL	Bracteatum-d	40:139	Foma
brc1	brc-5, com2	613	2HS	Branched 1	45:224	BGRC 13145
brh1	br	1	7HS	Brachytic 1	47: 50	Himalaya
brh2	br2	157	4HL	Brachytic 2	47: 84	Svanhals
brh3	brh.g, ert- t	631	2HS	Brachytic 3	45:229	Birgitta
brh4	brh.j	349	2HL	Brachytic 4	42:407	Birgitta
brh5	brh.m	185	4HS	Brachytic 5	44:110	Birgitta
brh6	brh.s	350	5HL	Brachytic 6	42:408	Akashinriki
brh7	brh.w	41	7H	Brachytic 7	42: 98	Volla
brh8	brh.ad	142	3HL	Brachytic 8	42:232	Birgitta
brh9	brh.k	187	4HL	Brachytic 9	43: 99	Birgitta
brh10	brh.l	653	2HS	Brachytic 10	45:231	Birgitta
brh11	brh.n	654	5HS	Brachytic 11	45:232	Birgitta
brh12	brh.o	655	5HS	Brachytic 12	45:233	Birgitta
brh13	brh.p	656	5HS	Brachytic 13	45:234	Birgitta
brh14	ari-o	148	7HL	Brachytic 14	45:111	Akashinriki
brh15	brh.u	657	2HL	Brachytic 15	44:205	Julia
brh16	brh.v	44	7HL	Brachytic 16	45: 89	Korál
brh17	brh.ab	658	5HS	Brachytic 17	45:236	Morex
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. Loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
brh18	brh13	659	5HS	Brachytic 18	45:237	Triumph
bsp1		645		Bushy spike 1	43:168	Morex
btr1	bt1	115	3HS	Non-brittle rachis 1	43: 78	A 222
btr2	bt2	116	3HS	Non-brittle rachis 2	43: 80	Sakigoke
cal-b		620	5HL	Calcaroides-b	44:197	Bonus
Cal-c		621	5HL	Calcaroides-c	41:195	Bonus
cal-d		146	3H	Calcaroides-d	40: 58	Foma
cal-e		622	5HS	Calcaroides-e	32:123	Semira
cer-d	cer-5	399	5HL	Eceriferum-d + + + +	41:153	Bonus
cer-e	cer-8	400	1HL	Eceriferum-e -/+ + + +	40:102	Bonus
cer-f	cer-9	401	1H	Eceriferum-f + + + +	40:104	Bonus
cer-g	cer-10	402	2HL	Eceriferum-g + + + +	47:131	Bonus
cer-h	cer-13	403	4HS	Eceriferum-h - + + +	41:157	Bonus
cer-i	cer-16	404	5HL	Eceriferum-i - + + +	41:158	Bonus
cer-k	cer-39	405	4HL	Eceriferum-k + + + +	41:160	Bonus
cer-l	cer-14	406	3HL	Eceriferum-l + + + +	44:142	Bonus
cer-m	cer-15	407	1H/3H	Eceriferum-m +/++ + + +	41:161	Bonus
cer-n	gs9,cer-20	408	2HL	Eceriferum-n - - + + & - +/- + +	44:143	Bonus
cer-o	cer-28	409	1HL	Eceriferum-o -/+ + + +	40:106	Bonus
cer-p	cer-37	410	7HL	Eceriferum-p + + + +	41:162	Bonus
cer-r	cer-19	411	3HL	Eceriferum-r +/- + + +	45:168	Bonus
cer-t	cer-22	412	5HL	Eceriferum-t +/- + + +	41:164	Bonus
cer-v	cer-49	414	2HS	Eceriferum-v +/- + + +	44:147	Bonus
cer-w	cer-48	415	5HL	Eceriferum-w +/- + + +	41:166	Bonus
cer-xa	cer-585	538	2HL/4 H/5HL	Eceriferum-xa + + + -	44:174	Foma
cer-xb	cer-943	539	4H	Eceriferum-xb - + + +	44:175	Bonus
cer-xc	cer-1371	540	1H	Eceriferum-xc + + + +	44:176	Bonus
cer-xd	cer-1455	541	4H/ 5HL	Eceriferum-xd + + + +	44:177	Bonus
cer-y	cer-72	417	1HS	Eceriferum-y + +/++ + +	44:149	Bonus
cer-ya	cer-180	444	3HS	Eceriferum-ya + + + -	26:396	Bonus
cer-yb	cer-200	445	2HL	Eceriferum-yb + + + -	41:171	Bonus
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
cer-yc	cer-135	446	6H/ 7HS	Eceriferum-yc - ++ ++	41:172	Bonus
cer-yd	cer-139	447	3HS	Eceriferum-yd - ++ ++	26:399	Bonus
cer-ye	cer-267	448	4H	Eceriferum-ye ++ ++ -	43:149	Foma
cer-yf	cer-652	449	7H	Eceriferum-yf ++ ++ +	44:160	Bonus
cer-yg	cer-1014	450	7HS	Eceriferum-yg - - -	44:161	Carlsberg II
cer-yh	cer-116	451	3HS	Eceriferum-yh - ++ ++	26:403	Bonus
cer-yi	cer-254	522	2H	Eceriferum-yi ++ ++ -	41:180	Foma
cer-yj	cer-667	523	1HS	Eceriferum-yj ++ ++ -	40:124	Bonus
cer-yk	cer-627	524	7HL	Eceriferum-yk + + ++	44:167	Bonus
cer-yl	cer-187	525	4HL	Eceriferum-yl - - ++	47:171	Bonus
cer-ym	cer-753	526	4HL	Eceriferum-ym - - -	47:173	Bonus
cer-yn	cer-1112	527	1H	Eceriferum-yn + + ++	40:125	Kristina
cer-yo	cer-647	528	4HS	Eceriferum-yo ++ ++ +	44:168	Bonus
cer-yp	cer-949	529	5HS	Eceriferum-yp ++ ++ +	44:169	Bonus
cer-yq	cer-1246	530	5H	Eceriferum-yq ++ ++ -	44:170	Kristina
cer-yr	cer-492	531	5HL	Eceriferum-yr -/+ + ++	44:171	Foma
cer-ys	cer-680	532	2HL	Eceriferum-ys ++ ++ -	44:172	Bonus
cer-yt	cer-758	533	1H/5H	Eceriferum-yt - ++ ++	40:126	Bonus
cer-yu	cer-158	534	1H	Eceriferum-yu ++ ++ -	40:127	Bonus
cer-yx	cer-421	535	1H/3H /5H	Eceriferum-yx + + ++	40:128	Foma
Cer-yy	Gle1	536	1HS	Eceriferum-yy - ++ ++	40:129	Bonus
cer-yz	cer-1452	537	1H/5H	Eceriferum-yz + + ++	44:173	Bonus
cer-z	cer-52	418	7HS	Eceriferum-z - - ++	44:150	Bonus
cer-za	cer-126	419	5HL	Eceriferum-za ++ ++ -	43:144	Foma
cer-zb	cer-38	420	5HS	Eceriferum-zb - ++ ++	42:508	Bonus
cer-zc	ccer-65	421	4HL/ 2HS	Eceriferum-zc +/- ++ ++	42:510	Bonus
cer-zd	cer-67	422	3HL	Eceriferum-zd ++ ++ -	40:110	Bonus
cer-ze	gl5	423	7HS	Eceriferum-ze ++ ++ -	44:152	Bonus
cer-zf	cer-70	424	3H/ 7HS	Eceriferum-zf ++ ++ +	42:516	Bonus
cer-zg	cer-214	425	4HL	Eceriferum-zg ++ ++ +	26:377	Foma
Table 3. (continued)						
Locus symbol*		BGS		Locus name or phenotype		Parental cultivar

Rec.	Prev.	no.	Chr. loc.†		Descr.v ol. p.	
cer-zi	cer-68	427	1HL	Eceriferum-zi + + + +	41:168	Bonus
cer-zj	cer-78	428	5HL	Eceriferum-zj + + + + -	42:520	Bonus
cer-zk	cer-85	429	2H	Eceriferum-zk + + + /-	26:381	Bonus
cer-zl	cer-17	430		Eceriferum-zl - - + +	26:382	Bonus
cer-zn	cer-244	431	1H	Eceriferum-zn +/- + + + +	40:112	Foma
cer-zo	cer-229	432	3HS	Eceriferum-zo - + + + +	44:154	Foma
cer-zp	cer-313	433	5HL	Eceriferum-zp + + + + -	26:385	Bonus
cer-zq	cer-248	434		Eceriferum-zq + + + + -	26:386	Foma
cer-zr	cer-260	435	5HL	Eceriferum-zr +/- + + + +	44:155	Foma
cer-zs	cer-467	436		Eceriferum-zs + + + + +	44:156	Foma
cer-zt	cer-389	437	2HS	Eceriferum-zt + + + + +	44:157	Foma
cer-zu	cer-122	438	1HS	Eceriferum-zu - + + + +	41:170	Foma
cer-zv	cer-268	439	4HL	Eceriferum-zv - - -	47:133	Foma
cer-zw	cer-286	440		Eceriferum-zw + + + + +	26:392	Foma
cer-zx	cer-100	441	3H	Eceriferum-zx + + + + +	46:111	Bonus
cer-zy	cer-118	442	1HS	Eceriferum-zy + + + + +	40:116	Bonus
cer-zz	cer-615	443		Eceriferum-zz + + + + -	44:159	Bonus
clh1	clh	225	7H/5H	Curled leaf dwarf 1	40: 76	Hannchen
com1	lax-270, bir1	473	5HL	Compositum 1	47:135	Foma
com2	bir2	71	2HS	Compositum 2	45: 95	CIMMYT freak
cr11	cl	325	6H	Curly lateral 1	41:129	Montcalm
crm1	cm	305	5HL	Cream seedling 1	26:256	Black Hulless
cst1	cs	673	5HL	Corn stalk	41:199	Husky
cud1	cud	324	5HL	Curly dwarf 1	26:272	Akashinriki
cud2		229	1HL	Curly dwarf 2	47:103	Akashinriki
cul2	uc2	253	6HL	Uniculm 2	43:112	Kindred
cul3	cul3	616	3HL	Uniculme 3	43:161	Donaria
cul4	uc-5	617	3HL	Uniculme 4	46:132	Bonus
cur1	cu1	262	3HL	Curly 1	46: 81	48-cr cr-17
cur2	cu2	114	3HL	Curly 2	44: 82	Choshiro
cur3	cu3	263	6HL	Curly 3	41:125	Akashinriki
cur4	glo-d	460	2HL	Curly 4	45:172	Asahi 5
Table 3. (continued)						
Locus symbol*		BGS		Locus name or phenotype		Parental cultivar

Rec.	Prev.	no.	Chr. loc.†		Descr.v ol. p.	
cur5	cu5	231	2HS	Curly 5	41:120	Glenn
ddt1	ddt	317	5HS	Reaction to DDT 1	26:266	Spartan
des1	lc	12	7H	Desynapsis 1	42: 58	Mars
des2	ds	119	3H	Desynapsis 2	43: 84	Husky
des3		386	2H/ 5HL	Desynapsis 3	43:140	Betzes
des4		13	7H	Desynapsis 4	44: 54	Betzes
des5		14	7HL	Desynapsis 5	44: 56	Betzes
des6		215	1HL	Desynapsis 6	43:106	Betzes
des7		64	3H	Desynapsis 7	43: 67	Betzes
des8		387	3H	Desynapsis 8	41:151	Betzes
des9		388	7HL	Desynapsis 9	44:131	Betzes
des10		389	3HL	Desynapsis 10	41:152	Betzes
des11		390	3HL	Desynapsis 11	44:132	Betzes
des12		391	3H	Desynapsis 12	44:133	Betzes
des13		392	3H	Desynapsis 13	44:134	Betzes
des14		393	7H	Desynapsis 14	44:135	Betzes
des15		394	3HL	Desynapsis 15	44:136	Ingrid
dex1	sex2	311	5HS	Defective endosperm xenia 1	26:260	BTT 63-j-18-17
dsk1	dsk	322	5HL	Dusky 1	41:128	Chikurin-Ibaraki 1
dsk2		729	7HL	Dusky 2	44:208	Betzes
dsp1	l	9	7HS	Dense spike 1	43: 50	Honen 6
dsp9	l9, ert-e	258	6HL	Dense spike 9	47:107	Akashinriki
dsp10	lc	111	3HS	Dense spike 10	41: 99	Club Mariout
dsp11	dsp	244	1HL	Dense spike 11	41:121	Akashinriki
dub1		661	6HL	Double seed 1	37:301	Bonus
Dwf2		542		Dominant dwarf 2	24:170	Klages / Mata
Eam1	Ea, Ppd-H1	65	2HS	Early maturity 1	47: 54	Estate
Eam5	HvPhyC- e	348	5HL	Early maturity 5	45:123	Higuerilla*2/ Gobernadora
Eam6	Ea6, Ea, HvCEN	98	2HS	Early maturity 6	46: 57	Morex
eam7	HvCO7	252	6HS	Early maturity 7	45:118	California Mariout
Table 3. (continued)						
Locus symbol*		BGS		Locus name or phenotype		Parental cultivar

Rec.	Prev.	no.	Chr. loc.†		Descr.v ol. p.	
eam8	ea _k , mat-a	214	1HL	Early maturity 8	46: 69	Kinai 5
eam9	ea _{,,c}	181	4HL	Early maturity 9	26:204	Tayeh 8
eam10	ea _{sp}	130	3HL	Early maturity 10	44: 86	Super Precoz
eli-a	lig-a	623		Eligulum-a	44:199	Foma
eog 1	e	57	2HL	Elongated outer glume 1	43: 64	Triple Bearded Club Mariout
ert-a	ert-6	28	7HS	Erectoides-a	41: 76	Gull
ert-b	ert-2	224	1HL	Erectoides-b	40: 74	Gull
ert-c	ert-1	134	3HL	Erectoides-c	41:108	Gull
ert-d	ert-7	29	7HS	Erectoides-d	42: 82	Gull
ert-e	l9, dsp9	266	6HL	Erectoides-e	47:109	Bonus
ert-f	ert-18	560	1H	Erectoides-f	40:133	Bonus
ert-g	ert-g	330	1HL	Erectoides-g	41:133	Bonus
ert-h	ert-25	561	5HL	Erectoides-h	44:189	Bonus
ert-ii	uzul	135	3HL	Erectoides-ii	45:109	Bonus
ert-j	ert-31	90	2H	Erectoides-j	43: 70	Bonus
ert-k	ert-32	562	6H	Erectoides-k	46:121	Bonus
ert-l	ert-12	563		Erectoides-l	26:489	Maja
ert-m	ert-34	30	7HS	Erectoides-m	46: 47	Bonus
ert-n	ert-51	331	5HL	Erectoides-n	44:120	Bonus
ert-p	ert-44	564		Erectoides-p	26:490	Bonus
ert-q	ert-101	91	6H	Erectoides-q	43: 71	Bonus
Ert-r	Ert-52	332	2HL	Erectoides-r	47:123	Bonus
ert-s	ert-50	565		Erectoides-s	26:491	Bonus
ert-t	brh3	566	2HS	Erectoides-t	45:203	Bonus
ert-u	ari-o	92	7HL	Erectoides-u	45:100	Bonus
ert-v	ert-57	567	6H	Erectoides-v	41:188	Bonus
ert-x	ert-58	568	1H/7H	Erectoides-x	40:136	Bonus
ert-y	ert-69	569		Erectoides-y	26:495	Bonus
ert-z	ert-71	570		Erectoides-z	26:496	Bonus
ert-za	ert-102	571	5H	Erectoides-za	44:189	Bonus
ert-zb	ert-132	572	7HL	Erectoides-zb	45:205	Bonus
ert-zc	ert-149	573	7HS	Erectoides-zc	45:206	Bonus
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
ert-zd	ari-o	93	7HL	Erectoides-zd	45:102	Bonus
ert-ze	ert-105	574	5HS	Erectoides-ze	45:207	Bonus
fch1	f, lg	55	2HS	Chlorina seedling 1	40: 49	Minn 84-7
fch2	f2, lg5	117	3HL	Chlorina seedling 2	46: 63	28-3398
fch3	f3	220	1HS	Chlorina seedling 3	40: 71	Minn 89-4
fch4	f4	17	7HL	Chlorina seedling 4	43: 54	Montcalm
fch5	f5	18	7HS	Chlorina seedling 5	43: 56	Gateway
fch6	f6	313	5HL	Chlorina seedling 6	44:116	Himalaya
fch7	f7	201	1HL	Chlorina seedling 7	41:113	Smyrna
fch8	f8	5	7HS	Chlorina seedling 8	41: 62	Comfort
fch9	f9	151	4HS	Chlorina seedling 9	44: 92	Ko A
fch10		177	4H	Chlorina seedling 10	43: 95	Unknown cultivar
fch11	f11	260	6HL	Chlorina seedling 11	45:207	Himalaya
fch12	f _c	2	7HS	Chlorina seedling 12	41:60	Colsess
fch13	f13	86	5HL	Chlorina seedling 13	44: 77	Nigrinudum
fch14	f14	87	2HL	Chlorina seedling 14	44: 78	Shyri
fch15	or	52	2HS	Chlorina seedling 15	40: 48	Trebi IV
fch16	clo.117	676	2HS	Chlorina seedling 16	40:144	Bonus
fch17	vy	191	1H/3H	Chlorina seedling 17	40: 68	Himalaya / Ingrescens
fer1		453		Few roots 1	46:112	Bomi Abed
flo-a		182	6HL	Extra floret-a	45:116	Foma
flo-b	flo-a	327	6HL	Extra floret-b	45:121	Foma
flo-c	flo-a	74	6HL	Extra floret-c	45: 97	Foma
fol-a		73	2HL	Angustifolium-a	43: 69	Proctor
Fol-b	Ang	548	1HS	Angustifolium-b	40:131	Unknown
fst1	fs	301	5HL	Fragile stem 1	26:252	Kamairazu
fst2	fs2	208	1HL	Fragile stem 2	41:114	Oshichi
fst3	fs3	24	7HS	Fragile stem 3	41: 74	Kobinkatagi 4
fxp1		728		Fenoxaprop-p-ethyl reaction 1	43:223	Morex
gig1	gig	463	2H	Gigas 1	44:164	Tochigi Golden Melon
gig2		612	4HL	Gigas 2	44:191	ND12463

Table 3. (continued)

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p	Parental cultivar
Rec.	Prev.					
glf1	gl, cer-zh	155	4HL	Glossy leaf 1 ++ ++ -	47: 81	Himalaya
glf3	gl3, cer-j	165	4HL	Glossy leaf 3 ++ ++ -	47: 86	Goseshikoku
glo-a		168	4H	Globosum-a	45:115	Proctor
glo-b		336	5HL	Globosum-b	26:284	Villa
glo-c	glo-c	72	2H	Globosum-c	43: 68	Villa
glo-e		230	1HL	Globosum-e	45:117	Foma
glo-f		342	5HL	Globosum-f	26:291	Damazay
gpa1	gp	59	2HL	Grandpa 1	45: 91	Lyallpur
gra-a	gran-a	131	7H	Granum-a	47: 79	Donaria
gsh1	gs1, cer-q	351	2HS	Glossy sheath 1 - - ++	46: 94	PI 195285
gsh2	gs2, cer-b	352	3HL	Glossy sheath 2 - - ++	47:125	Atlas
gsh3	gs3, cer-a	353	7HS	Glossy sheath 3 - - ++	41:143	Mars
gsh4	gs4, cer-x	354	6HL	Glossy sheath 4 - - ++	41:146	Gateway
gsh5	gs5, cer-s	355	2HL	Glossy sheath 5 + - ++	47:128	Jotun
gsh6	gs6, cer-c	356	2HS	Glossy sheath 6 - - ++	46:101	Betzes
gsh7	gs7	81	1H/2H /5H	Glossy sheath 7 - - ++	40: 55	Akashinriki
gsh8	gs8, cer-u	413	2HS	Glossy sheath 8 + + ++	46:105	Akashinriki
Gth1	G	69	2HL	Toothed lemma 1	47: 59	Machine (Wexelsen)
hcm1	h	77	2HL	Short culm 1	26:115	Morex
Hln1	Hn	164	4HL	Hairs on lemma nerves 1	44: 99	Kogane-mugi
Hsh1	Hs	179	4HL	Hairy leaf sheath 1	44:107	Kimugi
ibl1	ibl1	716	5HL	Intense blue aleurone 1	46:136	Ethiopian 637
int-b	vrs2	320	5HL	Intermedium spike-b	47:118	Bonus
int-c	i, vrs5, HvTB1	178	4HS	Intermedium spike-c	47: 92	Gamma 4
int-f	int-20	543	2HS/ 3HL	Intermedium spike-f	44:178	Foma
int-h	int-42	544	5H	Intermedium spike-h	44:179	Kristina
int-i	int-39	545	2HS	Intermedium spike-i	41:181	Kristina
int-k	int-47	546	7H	Intermedium spike-k	44:180	Kristina
int-m	int-85	547	5HL	Intermedium spike-m	44:181	Bonus
Kap1	K	152	4HS	Hooded lemma 1	26:179	Colsess

Table 3. (continued)

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
lab1		730	5HL	Labile 1	46:137	
lam1		651		Late maturity 1	43:174	Steptoe
lax-a	lax-8	474	5HL	Laxatum-a	46:113	Bonus
lax-b	lax-1	268	6HL	Laxatum-b	44:113	Bonus
lax-c	lax-21	475	6HL	Laxatum-c	47:137	Bonus
lax-d	lax-83	476		Laxatum-d	47:139	Bonus
lax-f	lax-61	477		Laxatum-f	47:140	Bonus
lax-g	lax-25	478		Laxatum-g	47:141	Bonus
lax-h	lax-26	479	3H	Laxatum-h	47:142	Bonus
lax-i	lax-50	480		Laxatum-i	47:144	Bonus
lax-j	lax-49	481	2H	Laxatum-j	47:145	Bonus
lax-k	lax-84	482	2H/3H	Laxatum-k	47:147	Bonus
lax-l	lax-60	483	1H	Laxatum-l	47:148	Bonus
lax-m	lax-67	484		Laxatum-m	47:149	Bonus
lax-n	lax-80	485	2H/4H L/5H	Laxatum-n	47:150	Bonus
lax-o	lax-79	486		Laxatum-o	47:152	Bonus
lax-df	lax-204	487		Laxatum-df	47:153	Foma
lax-ef	lax-225	488	2H	Laxatum-ef	47:154	Foma
lax-ff	lax-216	489		Laxatum-ff	47:156	Foma
lax-gf	lax-217	490		Laxatum-gf	47:157	Foma
lax-hf	lax-244	491	2H/3H L/5H	Laxatum-hf	47:158	Foma
lax-if	lax-246	492		Laxatum-if	47:160	Foma
Lax-jf	lax-253	493	4HS	Laxatum-jf	47:161	Foma
lax-kf	lax-295	494		Laxatum-kf	47:163	Foma
lax-lf	lax-274	495		Laxatum-lf	47:164	Foma
lax-mf	lax-302	496		Laxatum-mf	47:165	Foma
lax-nf	lax-322	497	2HL/ 4H	Laxatum-nf	47:166	Foma
lbi1	lb	308	5HL	Long basal rachis internode 1	43:125	Wisconsin Pedigree 38
lbi2	lb2	156	4HS	Long basal rachis internode 2	44: 92	Montcalm
lbi3	lb3	27	7HL	Long basal rachis internode 3	42: 79	Montcalm
Table 3. (continued)						
Locus symbol*				Locus name or phenotype		Parental cultivar

Rec.	Prev.	BGS no.	Chr. loc.†		Descr.v ol. p.	
lel1	lel	235	2H	Leafy lemma 1	46: 74	G7118
Lfb1	Lfb	343	5HL	Leafy bract 1	41:140	Montcalm
lfl1	lfl	275	6HL	Leafless 1	47:111	Golden Melon
Lga1	Log	549	7HS	Long glume awn 1	44:183	Guy Mayle
lgn2	lg2	169	4HS	Light green 2	42:264	Minn 75
lgn3	lg3, lg7	170	1HL	Light green 3	44:103	No 154
lgn4	lg4, lg1	171	4HL	Light green 4	44:105	Himalaya / Ingrescens
lig1	li, aur-a	60	2HL	Liguleless 1	45: 93	Muyoji
lit1	s, rin	99	2HS	Lesser internode number 1	41: 92	Natural occurrence
Lks1	Lk	75	2HL	Awnless 1	41: 84	<i>Hordeum inerme</i>
lks2	lk2	10	7HL	Short awn 2	45: 80	Honen 6
lks5	lk5	172	4HL	Short awn 5	47: 89	CIho 5641
lks6	lks.q	724	1H/5H /6H	Short awn 6	43:219	Morex
lnt1	rnt	118	3HL	Low number of tillers 1	47: 74	Mitake
lys1	lys	338	5HL	High lysine 1	26:286	Hiproly
lys3	sex3	339	5HL	High lysine 3	43:127	Bomi Abed
Lys4	sex5	232	1HS	High lysine 4	26:230	Bomi Abed
lys6		269	6H	High lysine 6	44:114	Bomi Abed
lzd1	dw4	125	3H	Lazy dwarf 1	43: 87	Akashinriki
mat-b	mat-7	578	7HL	Praematurum-b	46:123	Bonus
mat-c	Eam6, HvCEN	579	2H	Praematurum-c	46:125	Bonus
mat-d	mat-14	580	4HL/ 6HL	Praematurum-d	45:208	Bonus
mat-e	mat-18	581		Praematurum-e	26:508	Bonus
mat-f	mat-23	582	1H	Praematurum-f	45:210	Bonus
mat-g	mat-30	583		Praematurum-g	26:510	Bonus
mat-h	mat-36	584	4HL	Praematurum-h	45:212	Bonus
mat-i	mat-37	585	7HL	Praematurum-i	45:214	Bonus
min1	min	161	4HL	Semi-minute dwarf 1	44: 97	Taisho-mugi
min2	en-min	160		Enhancer of minute 1	26:186	Kaiyo Bozu
mnd1	m	519	2H	Many noded dwarf 1	43:154	Mesa
Table 3. (continued)						
Locus symbol*				Locus name or phenotype		Parental cultivar

Rec.	Prev.	BGS no.	Chr. loc. [†]		Descr.v ol. p.	
mnd3	m3	618	4HS	Many noded dwarf 3	44:194	Montcalm
mnd4	m4	347	5HL	Many noded dwarf 4	44:122	Akashinriki
mnd5		632	7HL	Many noded dwarf 5	44:202	C2-95-199
mnd6	den-6	633	5HL	Many noded dwarf 6	44:203	Bonus
mnd7		726		Many noded dwarf 7	43:221	Steptoe
mov1	mo5	43	7HL	Multiovary 1	43: 59	Steptoe
mov2	mo	147	3HS	Multiovary 2	43: 91	Steptoe
mov3	mo-a	234	1H	Multiovary 3	32:102	Akashinriki
mov4	mo8	648		Multiovary 4	43:171	Steptoe
mov5	mov.o	723		Multiovary 5	43:218	Morex
msg1	ms, ms1	357	1HL	Male sterile genetic 1	45:126	CIho 5368
msg2	ms2	358	2HL	Male sterile genetic 2	45:128	Manchuria
msg3	ms3	359	1HL	Male sterile genetic 3	45:130	Gateway
msg4	ms4	360	1H	Male sterile genetic 4	45:132	Freja
msg5	ms5	361	3HS	Male sterile genetic 5	45:133	Carlsberg II
msg6	ms6	362	6HS	Male sterile genetic 6	45:135	Hanna
msg7	ms7	363	5HL	Male sterile genetic 7	45:137	Dekap
msg8	ms8	364	5HL	Male sterile genetic 8	45:139	Betzes
msg9	ms9	365	2HS	Male sterile genetic 9	45:141	Vantage
msg10	ms10	366	7HS	Male sterile genetic 10	45:142	Compana
msg11	ms11	367	5HS	Male sterile genetic 11	45:144	Gateway
msg13	ms13	368	3HL	Male sterile genetic 13	45:146	Haisa II
msg14	ms14	369	7HS	Male sterile genetic 14	45:147	Unitan
msg15	ms15	370		Male sterile genetic 15	45:149	Atlas/2*Kindred
msg16	ms16	371	5HS	Male sterile genetic 16	45:150	Betzes
msg17	ms17	372	5HL	Male sterile genetic 17	45:152	Compana
msg18	ms18	373	5HL	Male sterile genetic 18	45:153	Compana
msg19	ms19	374	5HS	Male sterile genetic 19	45:155	CIho 14393
msg20	msg,,ad	375	4H	Male sterile genetic 20	45:156	Hannchen
msg21	ms21	376	1HL	Male sterile genetic 21	45:157	Midwest Bulk
msg22	ms22	383	7H	Male sterile genetic 22	45:162	Glacier / Compana
msg23	ms23	384	5H	Male sterile genetic 23	45:163	Betzes
msg24	ms24	385	4HL	Male sterile genetic 24	45:164	Betzes

Table 3. (continued)

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
msg25	msg,,r	166	2HS	Male sterile genetic 25	45:113	Betzes
msg26	msg,,u	395	7HS	Male sterile genetic 26	45:166	Unitan
msg27	msg,,ae	464	2HS	Male sterile genetic 27	45:174	Firlbecks III
msg28	msg,,as	465	2HS	Male sterile genetic 28	45:175	York
msg29	msg,,a	466	5HL	Male sterile genetic 29	45:176	Ackermans MGZ
msg30	msg,,c	467	7HL	Male sterile genetic 30	45:177	Compana
msg31	msg,,d	468	1HL	Male sterile genetic 31	45:178	51Ab4834
msg32	msg,,w	469	7H	Male sterile genetic 32	45:179	Betzes
msg33	msg,,x	470	2HS	Male sterile genetic 33	45:180	Betzes
msg34	msg,,av	471	6HS/ 7HS	Male sterile genetic 34	45:181	Paragon
msg35	msg,,dr	498	2HL	Male sterile genetic 35	45:183	Karl
msg36	msg,,bk	499	6HS	Male sterile genetic 36	45:184	Betzes
msg37	msg,,hl	500	3HL	Male sterile genetic 37	45:186	Clermont
msg38	msg,,jl	501	3H	Male sterile genetic 38	45:187	Ingrid
msg39	msg,,dm	502	3H	Male sterile genetic 39	45:188	P11
msg40	msg,,ac	503	6HL	Male sterile genetic 40	45:190	Conquest
msg41	msg,,aj	504	6HS	Male sterile genetic 41	45:191	Betzes
msg42	msg,,db	505	3H	Male sterile genetic 42	45:193	Betzes
msg43	msg,,br	506	2HL	Male sterile genetic 43	45:194	Betzes
msg44	msg,,cx	507	5HL	Male sterile genetic 44	45:195	HA6-33-02
msg45	msg,,dp	508	5HL/ 7HS	Male sterile genetic 45	45:196	RPB439-71
msg46	msg,,ec	509	2H/6H	Male sterile genetic 46	45:197	Hector
msg47	msg,,ep	510	3HS/ 7HS	Male sterile genetic 47	45:198	Sel12384CO
msg48	msg,,jt	520	1H	Male sterile genetic 48	45:199	Simba
msg49	msg,,jw	335	5HL	Male sterile genetic 49	45:122	ND7369
msg50	msg,,hm	34	7HL	Male sterile genetic 50	45: 88	Berac
mss1	mss	84	5H	Midseason stripe 1	44: 75	Montcalm
mss2		39	7HS	Midseason stripe 2	44: 59	ND11258
mtt1	mt	521	1HS	Mottled leaf 1	41:179	Montcalm
mtt2	mt2	302	5HL	Mottled leaf 2	41:127	Montcalm
mtt4	mt,,e	78	2HL	Mottled leaf 4	41: 86	Victorie
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
mtt5	mt,,f	264	6HL	Mottled leaf 5	41:126	Akashinriki
mtt6		629	7HS	Mottled leaf 6	45:228	ND6809
mtt7	mtt.h	677	2HS	Mottled leaf 7	42:753	Morex
mtt8	Mut 1661	674		Mottled leaf 8	43:182	Bowman Rph3.c
mtt9	Mut 2721	675		Mottled leaf 9	44:207	Bowman Rph3.c
mul2		251	6HL	Multiflorus 2	26:232	Montcalm
nar1		637	6HS	NADH nitrate reductase-deficient 1	35:194	Steptoe
nar2		638	5HL	NADH nitrate reductase-deficient 2	35:196	Steptoe
nar3		639	7HS	NADH nitrate reductase-deficient 3	35:197	Winer
nar4		640	2HL	NADH nitrate reductase-deficient 4	35:198	Steptoe
nar5		641	5HL	NADH nitrate reductase-deficient 5	35:199	Steptoe
nar6		642	2HL	NADH nitrate reductase-deficient 6	35:200	Steptoe
nar7		643	6HL	NADH nitrate reductase-deficient 7	35:201	Steptoe
nar8		644	6HS	NADH nitrate reductase-deficient 8	35:202	Steptoe
nec1	sp,,b	222	1HL	Necrotic leaf spot 1	43:108	Carlsberg II
nec2	nec2	261	6HS	Necrotic leaf spot 2	26:241	Carlsberg II
nec3	nec3	265	6HS	Necrotic leaf spot 3	43:116	Proctor
nec4	nec4	138	3H	Necrotic leaf spot 4	43: 88	Proctor
nec5	nec5	139	3H	Necrotic leaf spot 5	43: 89	Diamant
Nec6	Sp	611	7HS	Necrotic leaf spot 6	43:159	Awnless Atlas
nec7	nec-45	635	1H/6H /7H	Necroticans 7	43:166	Kristina
nec8	nec.w	671	5HL	Necrotic leaf spot 8	43:179	Bowman Rph3.c
nec9	Mut 3091	672	3HL	Necrotic leaf spot 9	43:181	Bowman Rph3.c
nec10	necS 1-1	681	3H	Necroticans 10	43:184	Steptoe
nec11		682	1H	Necroticans 11	43:185	Steptoe
nec12		683		Necroticans 12	43:186	Steptoe
nec13		684		Necroticans 13	43:187	Steptoe
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
nec14		685		Necroticans 14	43:188	Steptoe
nec15		686		Necroticans 15	43:189	Steptoe
nec16t		687		Necroticans 16	43:190	Steptoe
nec17		688		Necroticans 17	43:191	Steptoe
nec18		689		Necroticans 18	43:192	Steptoe
nec19		690		Necroticans 19	43:193	Steptoe
nec20		691		Necroticans 20	43:194	Steptoe
nec21		692		Necroticans 21	43:195	Steptoe
Nec22		693		Necroticans 22	43:196	Steptoe
nec23		694		Necroticans 23	43:197	Steptoe
Nec24		695		Necroticans 24	43:198	Steptoe
nec25		696		Necroticans 25	43:199	Steptoe
Nec26		697		Necroticans 26	43:200	Steptoe
nec27		698		Necroticans 27	43:201	Steptoe
nec28		699		Necroticans 28	43:202	Morex
nec29		700		Necroticans 29	43:203	Morex
nec30		701		Necroticans 30	43:204	Morex
nec31		702		Necroticans 31	43:205	Morex
nec32		703		Necroticans 32	43:206	Morex
nec33		704		Necroticans 33	43:207	CIho 4196
nec34	nec.k	197	4HS	Necroticans 34	43:104	ND13917
nld1	nld	323	5HL	Narrow leafed dwarf 1	46: 88	Nagaoka
nld2		660	5H/6H /7H	Narrow leafed dwarf 2	43:176	Steptoe
nud1	n, nud	7	7HL	Naked caryopsis 1	44: 51	Himalaya
ops1	op-3	624	7HS	Opposite spikelets 1	45:226	Bonus
ops2	op-2	718	5HL	Opposite spikelets 2	43:213	Foma
ops3	op-1	719	5HS	Opposite spikelets 3	43:214	Bonus
ovl1		176	4H	Ovaryless 1	35:191	Kanto Bansei Gold
ovl2	ovl2	646		Ovaryless 2	43:169	Harrington
ovl3		725		Ovaryless 3	43:220	Harrington
Pbg1	Pbg	740	7H	Pubescens on glume 1	47:182	
pmr1	pmr	40	7HS	Premature ripe 1	44: 60	Glenn
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
pmr2	nec-50	634		Premature ripe 2	32:135	Bonus
Pre2	Re2	76	2HL	Red lemma and pericarp 2	46: 53	Buckley 3277
Pub1	Pub	127	3HL	Pubescent leaf blade 1	44: 84	Multiple Dominant
Pvc1	P _c	68	2HL	Purple veined lemma 1	44: 67	Buckley 2223-6
Pyr1	Pyr.g	42	3HL	Pyramidatum 1	41: 78	Pokko/Hja80001
raw1	r	312	5HL	Smooth awn 1	46: 84	Lion
raw2	r2	340	5HL	Smooth awn 2	46: 92	Lion
raw5	r ₅ ,e	257	6HL	Smooth awn 5	44:112	Akashinriki
raw6	r6	334	5HL	Smooth awn 6	26:282	Glenn
rcr1	γ08-122	737		Required for resistance to <i>Cochliobolus sativus</i> 1	45:249	Morex
rcr2	γ08-123	738		Required for resistance to <i>Cochliobolus sativus</i> 2	45:250	Morex
rcr3	γ08-124	739		Required for resistance to <i>Cochliobolus sativus</i> 3	45:251	Morex
rob1	o	254	6HS	Orange lemma 1	46: 78	CIho 5649
Rpc1		149	3H	Reaction to <i>Puccinia coronata</i> var. <i>hordei</i> 1	37:232	Hor 2596
Rpg1	T	511	7HS	Reaction to <i>Puccinia graminis</i> 1	26:437	Chevron
Rpg2	T2	512		Reaction to <i>Puccinia graminis</i> 2	26:439	Hietpas 5
rpg4		319	5HL	Reaction to <i>Puccinia graminis</i> 4	26:267	Q21861
Rph1	Pa	70	2H	Reaction to <i>Puccinia hordei</i> 1	26:107	Oderbrucker
Rph2	Pa2	88	5HS	Reaction to <i>Puccinia hordei</i> 2	37:212	Peruvian
Rph3	Pa3	121	7HL	Reaction to <i>Puccinia hordei</i> 3	26:156	Estate
Rph4	Pa4	218	1HS	Reaction to <i>Puccinia hordei</i> 4	42:302	Gull
Rph5	Pa5	122	3HS	Reaction to <i>Puccinia hordei</i> 5	37:224	Magnif 102
Rph6	Pa6	575	3HS	Reaction to <i>Puccinia hordei</i> 6	26:501	Bolivia
Rph7	Pa7	136	3HS	Reaction to <i>Puccinia hordei</i> 7	37:228	Cebada Capa
Rph8	Pa8	576		Reaction to <i>Puccinia hordei</i> 8	26:502	Egypt 4
Rph9	Pa9, Pa12	32	5HL	Reaction to <i>Puccinia hordei</i> 9	37:201	HOR 2596
Rph10		137	3HL	Reaction to <i>Puccinia hordei</i> 10	26:174	Clipper C8
Table 3. (continued)						

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
Rph11		267	6HL	Reaction to <i>Puccinia hordei</i> 11	26:247	Clipper C67
Rph12	Pa12, Pa9	333	5HL	Reaction to <i>Puccinia hordei</i> 12	26:281	Triumph
Rph13		590		Reaction to <i>Puccinia hordei</i> 13	28: 31	PI 531849
Rph14		591		Reaction to <i>Puccinia hordei</i> 14	28: 32	PI 584760
Rph15	Rph16	96	2HL	Reaction to <i>Puccinia hordei</i> 15	37:214	PI 355447
rpr1		707	4H	Required for <i>Puccinia graminis</i> resistance 1	42:757	Morex
rpr2	γ08-118	731	6H	Required for <i>Puccinia graminis</i> resistance 2	46:139	Morex
rpr3	γ08-112	732		Required for <i>Puccinia graminis</i> resistance 3	46:141	Morex
rpr4	γ08-114	733		Required for <i>Puccinia graminis</i> resistance 4	46:142	Morex
rpr5	γ08-117	734		Required for <i>Puccinia graminis</i> resistance 5	46:143	Morex
rpr6	γ08-119	735		Required for <i>Puccinia graminis</i> resistance 6	46:144	Morex
rpr7	γ08-115	736		Required for <i>Puccinia graminis</i> resistance 7	46:145	Morex
Rpt1	Pt	667	3HL	Reaction to <i>Pyrenophora teres</i> 1	43:177	Tifang
Rpt2	Rpt2c	237	1HS	Reaction to <i>Pyrenophora teres</i> 2	43:110	CIho 9819
Rpt3	QRpts2	711	2HS	Reaction to <i>Pyrenophora teres</i> 3	43:208	Tennessee Awnless D22-5
Rpt4	QRpt7	48	7HL	Reaction to <i>Pyrenophora teres</i> 4	43: 61	Galleon
Rpt5	Rpta	272	6HL	Reaction to <i>Pyrenophora teres</i> 5	43:120	CIho 5791, CIho 9819
Rpt6		713	5HL	Reaction to <i>Pyrenophora teres</i> 6	43:210	CIho 9819
Rpt7	Qrpts4	714	4HL	Reaction to <i>Pyrenophora teres</i> 7	43:211	Halcyon
Table 3. (continued)						
Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p	Parental cultivar
Rec.	Prev.					

Rpt8	QRpts4	198	4HS	Reaction to <i>Pyrenophora teres</i> 8	43:105	Q21861
Rsg1	Grb	22	7H	Reaction to <i>Schizaphis graminum</i> 1	37:199	Omugi
Rsg2		577		Reaction to <i>Schizaphis graminum</i> 2	37:283	PI 426756
rsm1	sm	35	7HS	Reaction to BSMV 1	26: 84	Modjo 1
Rsp1	Sep	515		Reaction to <i>Septoria passerinii</i> 1	26:441	CIho 14300
Rsp2	Sep2	516		Reaction to <i>Septoria passerinii</i> 2	37:275	PI 70837
Rsp3	Sep3	517		Reaction to <i>Septoria passerinii</i> 3	37:276	CIho 10644
rtt1	rt	51	2HS	Rattail spike 1	47: 53	Goldfoil
Run1	Un	21	7HS	Reaction to <i>Ustilago nuda</i> 1	26: 67	Trebi
rv11	rvl	226	1HL	Revoluted leaf 1	40: 77	Hakata 2
Ryd2	Yd2	123	3HL	Reaction to BYDV 2	26:158	CIho 2376
Rym1	Ym	167	4HL	Reaction to BaYMV 1	32: 96	Mokusekko 3
Rym2	Ym2	20	7HL	Reaction to BaYMV 2	26: 66	Mihori Hadaka 3
rym3	ym3	345	5HS	Reaction to BaYMV 3	32:105	Chikurin Ibaraki
rym5	Ym	141	3HL	Reaction to BaYMV 5	32: 90	Mokusekko 3
sbk1	sk, cal-a	62	2HS	Subjacent hood 1	40: 51	Tayeh 13
sca1	sca	128	3HS	Short crooked awn 1	44: 85	Akashinriki
sci-a	sci-3	625	5H	Scirpoides-a	44:200	Bonus
sci-b	sci-4	239	1H/6 H	Scirpoides-b	46: 76	Bonus
scl-a	scl-6	626	1HL	Scirpoides leaf-a	44:201	Foma
scl-b	scl-5	150	3H/6 H	Scirpoides leaf-b	40: 60	Bonus
sdw1	denso, Hv20ox ₂	518	3HL	Semidwarf 1	47:168	M21
sdw2	sdw-b	133	3HL	Semidwarf 2	45:108	Mg2170
sdw3	gai	663	2HS	Semidwarf 3	46:134	Hv287
sdw4		45	7HL	Semidwarf 4	46: 51	
sdw6	sdw.f	240	1H/7 H	Semidwarf 6	46: 77	Vada

Table 3. (continued)

Locus symbol*		BGS no.	Chr. loc.†	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					

sdw7	sdw.u	196	4HL	Semidwarf 7	43:103	Glenn
seg1	se1	377	7HL	Shrunken endosperm genetic 1	45:158	Betzes
seg2	se2	378	7HS	Shrunken endosperm genetic 2	26:326	Betzes
seg3	se3	379	3H	Shrunken endosperm genetic 3	45:160	Compana
seg4	se4	380	7HL	Shrunken endosperm genetic 4	37:267	Compana
seg5	se5	381	7HS	Shrunken endosperm genetic 5	26:329	Sermo / 7*Glacier
seg6	se6	396	3HL	Shrunken endosperm genetic 6	44:138	Ingrid
seg7	se7	397		Shrunken endosperm genetic 7	37:269	Ingrid
sex7	sex.i	628	5HL	Shrunken endosperm xenia 7	32:129	I90-374
sex8	sex.j	143	3HS	Shrunken endosperm xenia 8	43: 90	I89-633-1
sex9	sex.l	195	4HL	Shrunken endosperm xenia 9	43:102	Alf
sgh1	sh1	163	4HL	Spring growth habit 1	26:188	Iwate Mensury C
Sgh2	Sh2	309	5HL	Spring growth habit 2	26:259	Indian Barley
Sgh3	Sh3	213	1HL	Spring growth habit 3	26:212	Tammi / Hayakiso 2
sid1	nls	180	4HL	Single internode dwarf 1	43: 97	Akashinriki
Sil1	Sil	228	1H	Subcrown internode length 1	40: 79	NE 62203
sld1	dw-1	126	3HL	Slender dwarf 1	41:103	Akashinriki
sld2		83	2HS	Slender dwarf 2	44: 74	Akashinriki
sld3	ant-567	186	4HS	Slender dwarf 3	40: 63	Manker
sld4	sld.d	100	2HS	Slender dwarf 4	43: 72	Glacier
sld5		144	3HS	Slender dwarf 5	44: 90	Indian Dwarf
sld6	sld.gs	242	1H	Slender dwarf 6	40: 87	Glenn
sld7	sld.f	194	4HL	Slender dwarf 7	43:101	Glenn
sld8	sld.i	49	7HS/ 4HL	Slender dwarf 8	43: 63	Wisconsin Pedigree 38
sls1	sls	227	1HS	Small lateral spikelet 1	40: 78	Morex
smn1	smn	38	3H/5 H	Seminudoides 1	43: 58	Haisa
snb1	sb	26	7HS	Subnodal bract 1	26: 72	L50-200
srh1	S, l	321	5HL	Short rachilla hair 1	46: 86	Lion
stb1	stb	741		Stubble 1	47:183	
sun1	sun1	650		Sensitivity to <i>Ustilago nuda</i> 1	43:173	Steptoe
tfm1		190	1HL	Thick filament 1	40: 67	Volla

Table 3. (continued)

Locus symbol*		BGS no.	Chr. loc. [†]	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					

trd1	t, bra-c	202	1HL	Third outer glume 1	46: 65	Valki
trp1	tr	61	4HL	Triple awned lemma 1	41: 82	CIho 6630
tst1	tst1	647	6HL	Tip sterile 1	43:170	Steptoe
tst2		636	4HL	Tip sterile 2	43:167	Donaria
twk1	tw	743	7H	Tweaky spike 1	47:185	
ubs4	lks2, ari-d	11	7HL	Unbranched style 4	45: 84	Ao-Hadaka
ubs5		727		Unbranched style 5	43:222	Harrington
uzu1	uz, HvBR11	102	3HL	Uzu 1 or semi brachytic 1	47: 70	Baitori
var1	va	306	5HL	Variegated 1	37:259	Montcalm
var2	va2	344	5HL	Variegated 2	32:104	Montcalm
var3	va3	303	5HL	Variegated 3	44:115	Montcalm
viv-a	viv-5	627	2H	Viviparoides-a	45:227	Foma
viv-b	viv-6	193	4HS	Viviparoides-b	43:100	Foma
viv-c	viv-1	720	5H	Viviparoides-c	43:215	Foma
vrs1	v	6	2HL	Six-rowed spike 1	37:192	Trebi
vrs1	lr	58	2HL	Six-rowed spike 1	26: 94	Nudihaxtoni
vrs1	V ^d	66	2HL	Two-rowed spike 1	26:103	Svanhals
vrs1	V ^t	67	2HL	Deficiens 1	47: 57	White Deficiens
vrs2	v2, int-b	314	5HL	Six-rowed spike 2	47:113	Svanhals
vrs3	v3, int-a	315	1HL	Six-rowed spike 3	47:115	Hadata 2
vrs4	v4, int-e, HvRA2	124	3HL	Six-rowed spike 4	47: 76	MFB 104
wax1	wx	16	7HS	Waxy endosperm 1	42: 65	Oderbrucker
wnd1	wnd	23	4Hl	Winding dwarf 1	42: 74	Kogen-mugi
wst1	wst	107	3HL	White streak 1	41: 97	CIho 11767
wst2	wst2	304	5HL	White streak 2	46: 83	Manabe
wst4		56	2HL	White streak 4	44: 51	Kanyo 7
wst5	wst5	221	1HL	White streak 5	46: 73	Carlsberg II
wst6	wst,,j	129	3HL	White streak 6	41:105	Akashinriki
wst7	rb	79	2HL	White streak 7	47: 61	GS397
wxs1	wxs	615	7H/ 2HL	Waxy spike 1	43:160	Steptoe

Table 3. (continued).

Locus symbol*		BGS no	Chr. loc. [†]	Locus name or phenotype	Descr.v ol. p.	Parental cultivar
Rec.	Prev.					
Xnt1	X _a	25	7HL	Xantha seedling 1	26: 71	Akashinriki
xnt2	x _b	513		Xantha seedling 2	26:440	Black Hulless

xnt3	x _c	105	3HS	Xantha seedling 3	26:139	Colsess
xnt4	x _{c2}	36	7HL	Xantha seedling 4	26: 85	Coast
xnt5	x _n	255	6HL	Xantha seedling 5	26:237	Nepal
xnt6	x _s	113	3HS	Xantha seedling 6	26:147	Smyrna
xnt7	xan,,g	233	1HL	Xantha seedling 7	26:231	Erbet
xnt8	xan,,h	140	3HS	Xantha seedling 8	26:177	Carlsberg II
xnt9	xan,,i	37	7HL	Xantha seedling 9	26: 86	Erbet
yhd1	yh	158	4HL	Yellow head 1	42:250	Kimugi
yhd2	yh2	592		Yellow head 2	45:215	Compana
ylf1	ylf1	652	7HS	Yellow leaf 1	43:175	Villa
Ynd1	Yn	183	4HS	Yellow node 1	44:109	Morex
yst1	yst	104	3HS	Yellow streak 1	42:178	Gateway
yst2		109	3HS	Yellow streak 2	44: 81	Kuromugi 148 / Mensury C
yst3	yst,,c	462	3HS	Yellow streak 3	44:163	Lion
yst4		85	2HL	Yellow streak 4	44: 76	Glenn
yst5	yst5	346	7HS	Yellow streak 5	43:130	Bowman / ant10.30
yvs1	y _x	63	2HS	Virescent seedling 1	26: 99	Minn 71-8
yvs2	y _c	3	7HS	Virescent seedling 2	26: 46	Coast
yhd2	yh2	592		Yellow head 2	45:215	Compana
ylf1	ylf1	652	7HS	Yellow leaf 1	43:175	Villa
zeb1	zb	120	3HL	Zebra stripe 1	43: 86	Mars
zeb2	zb2, fch10	461	4HL	Zebra stripe 2	43:152	Unknown cultivar
zeb3	zb3, zb	223	1HL	Zebra stripe 3	40: 72	Utah 41
Zeo1	Knd, Ert-r	82	2HL	Zeocriton 1	47: 63	Donaria
Zeo2	cly1.b, HvAP2	614	2HL	Zeocriton 2	47:177	36Ab51
Zeo3	Zeo2, cly1.c	184	4HL	Zeocriton 3	47: 95	Morex

* Recommended locus symbols are based on utilization of a three-letter code for barley genes as approved at the business meeting of the Seventh International Barley Genetics Symposium at Saskatoon, Saskatchewan, Canada, on 05 August 1996.

† Chromosome numbers and arm designations are based on the Triticeae system. Utilization of this system for naming of barley chromosomes was at the business meeting of the Seventh International Barley Genetics Symposium at Saskatoon, Saskatchewan, Canada, on 05 August 1996. The Burnham

and Hagberg (1956) designations of barley chromosomes were 1 2 3 4 5 6 and 7 while new designations based on the Triticeae system are 7H 2H 3H 4H 1H 6H and 5H, respectively.

Stock number: BGS 1
Locus name: Brachytic 1
Locus symbol: *brh1*

Previous nomenclature and gene symbolization:

Brachytic = *br* (15, 17).
Breviaristatum-i = *ari-i* (7, 12).
Breviaristatum-28 = *ari-28* (14).
Dwarf x = *dx1* (8).

Inheritance:

Monofactorial recessive (15, 17).
Located in chromosome 7HS (5); *brh1.a* is about 9.3 cM distal from the *fch12* (chlorina seedling 12) locus (17); *brh1.ae* mapped 0.8 cM distal from RFLP marker BCD129 and cosegregated with marker MWG2074B (13); *brh1.a* is about 5.0 cM from AFLP marker E4134-8 in subgroup 1 of the Proctor/Nudinka map (16); *brh1.a* is about 13.6 cM proximal from SSR marker HVM04 in 7H bin 02 (3); *brh1.a* is associated with SNP markers 2_1419 to 2_0245 (positions 0.00 to 13.19 cM) in 7H bin 01 of the Bowman backcross-derived line BW074 (4); *brh1.x* is associated with SNP markers 1_0949 to 1_1495 (positions 0.00 to 8.77 cM) in 7H bin 01 of the Bowman backcross-derived line BW079 (4); *brh1.t* is associated with SNP markers 1_1495 to 1_0025 (positions 8.77 to 26.00 cM) in 7H bins 01 to 02 of the Bowman backcross-derived line BW078 (4); an unnamed *brh1* mutant is associated with SNP markers 1_0851 to 2_0485 (positions 17.32 to 121.90 cM) in 7H bins 02 to 07 of the Bowman backcross-derived line BW872 (4); the genomic position of the *Brh1* locus at 12.74 cM in 7HS (2), in 1H bin 01 near the border with 1H bin 02.

Description:

Plants have short leaves, culms, spikes, awns, and kernels. The seedling leaf is about 2/3 normal length. A similar reduction in the size of other organs is observed, but the awns were less than 1/2 normal length (8). The mutant phenotype was easy to classify at all stages of growth. The approximately 20% reduction in kernel weight was caused primarily by a reduction in kernel length, 7.8 vs. 9.6 mm. Grain yields of the Bowman backcross-derived lines for *brh1* mutants were about 2/3 normal and lodging was greatly reduced (2, 3). Börner (1) reported that *ari-i.38* seedlings are sensitive to gibberellic acid. Powers (15) stated that the assigned gene symbol for this mutant is *br* and that L.J. Stadler selected this symbol. The *ari-m* (breviaristatum-m) mutants occur in the *Brh1* locus (2). The *brh1* mutants are deficient in the G α subunit of a heterotrimeric G protein, which is an important regulator of culm length in barley (2, 9). The *brh1.a* mutant did not affect malt quality negatively (2).

Origin of mutant:

A spontaneous mutant in Himalaya (Clho 1312) (15, 17).

Mutational events:

brh1.a (GSHO 25) in Himalaya (Clho 1312) (17); *brh1.c* (GSHO 229) in Moravian (PI 539135) (18), but SNP marker patterns this mutant originated from Himalaya and is identical to the *brh1.a* mutant (2); *ari-i.38* (NGB 115888, GSHO 1657) in Bonus (NGB 14657, PI 189763) (12, 19); *brh1.e* (GSHO 1690) in Aramir (PI 467786) (19); *brh1.f* (*dx1*, GSHO 1422) in Domen (NGB 2104, Clho 9562) (8); *brh1.t* (OUM136, GSHO 1691) in

Akashinriki (PI 467400, OUJ659); *brh1.x* (7125, GSHO 1692) in Volla (PI 280423); *brh1.z* (Hja80001) in Aapo (PI 467771); *brh1.aa* (Hja80051) in a Hja80001 cross (6, 10); *brh1.ae* (FN53, GSHO 3646) in Steptoe (CIho 15229) (6, 11); an unnamed variant of *brh1* in L50-200 (Alb Acc 67A, GSHO 1217) associated with expression of the subnodal bract 1 (*snb1.a*) gene (20). The *ari-m* (breviaristatum-m) mutants were demonstrated to be alleles at the *brh1* locus (2); *ari-m.12* (NGB 115858, GSHO 1661), *-m.28* (NGB 115876) in Bonus (NGB 14657, PI 189763), *-m.141* (NGB 115951), *-m.177* (NGB 115987) in Foma (NGB 14659, CIho 11333), *-m.251* (NGB 116059) in Kristina (NGB 14661, NGB 1500) (12); *ari-m.269* (NGB 116081) in Kristina (12, 14).

Mutant used for description and seed stocks:

brh1.a in Himalaya (GSHO 25); *ari-i.38* (GSHO 1657, NGB 115888) in Bonus, *brh1.a* in Bowman (PI 483237)*7 (GSHO 1820, BW074, NGB 20481); *ari-i.38* in Bowman*6 (GSHO 1821, BW047, NGB 20455); *brh1.e* in Bowman*7 (GSHO 1822, BW077, NGB 20484); *brh1.t* in Bowman*7 (GSHO 1823, BW078, NGB 20485); *brh1.x* in Bowman*7 (GSHO 1824, BW079, NGB 20486); *brh1.z* in Bowman*7 (GSHO 2179, BW080, NGB 20487); *brh1.aa* in Bowman*6 (GSHO 1668, BW075, NGB 20482); *brh1.ae* in Bowman*4 (BW076, NGB 20483); an unnamed *brh1* allele in Bowman*2 (BW872, NGB 22306); *ari-m.28* (GSHO 1661, NGB 115876) in Bonus; *ari-m.28* in Bowman (PI 483237)*6 (GSHO 2161); *ari-m.28* in Bowman*8 (BW051, NGB 20459).

References:

1. Börner, A. 1996. GA response in semidwarf barley. *Barley Genet. Newsl.* 25:24-26.
2. Braumann, I., C. Dockter, S. Beier, A. Himmelbach, F. Lok, U. Lundqvist, B. Skadhauge, N. Stein, S. Zakhrabekova, R. Zhou, and M. Hansson. 2018. Mutations in the gene of the G α subunit of the heterotrimeric G protein are the cause for the *brachytic1* semi-dwarf phenotype in barley and applicable for practical breeding. *Hereditas* 155:10.
3. Dahleen, L.S., L.J. Vander Wal, and J.D. Franckowiak. 2005. Characterization and molecular mapping of genes determining semidwarfism in barley. *J. Hered.* 96:654-662.
4. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
5. Fedak, G., T. Tsuchiya, and S.B. Helgason. 1972. Use of monotelotrisomics for linkage mapping in barley. *Can. J. Genet. Cytol.* 14:949-957.
6. Franckowiak, J.D. 1995. The brachytic class of semidwarf mutants in barley. *Barley Genet. Newsl.* 24:56-59.
7. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
8. Holm, E., and K. Aastveit. 1966. Induction and effects of the brachytic allele in barley. *Adv. Front Plant Sci.* 17:81-94.
9. Ito, A., A. Yasuda, K. Yamaoka, M. Ueda, A. Nakayama, S. Takatsuto, and I. Honda. 2017. Brachytic 1 of barley (*Hordeum vulgare* L.) encodes the α subunit of heterotrimeric G protein. *J. Plant Physiol.* 213:209-215.
10. Kivi, E. 1986. (Personal communications).
11. Kleinhofs, A. (Unpublished).
12. Kucera, J., U. Lundqvist, and Å. Gustafsson. 1975. Inheritance of breviaristatum mutants in barley. *Hereditas* 80:263-278.
13. Li, M., D. Kudrna, and A. Kleinhofs. 2001. Fine mapping of a semi-dwarf gene *brachytic1* in barley. *Barley Genet. Newsl.* 31:14-17.

14. Lundqvist, U. (Unpublished).
15. Powers, L. 1936. The nature of the interactions of genes affecting four quantitative characters in a cross between *Hordeum deficiens* and *vulgare*. *Genetics* 21:398-420.
16. Pozzi, C., D. di Pietro, G. Halas, C. Roig, and F. Salamini. 2003. Integration of a barley (*Hordeum vulgare*) molecular linkage map with the position of genetic loci hosting 29 developmental mutants. *Heredity* 90:390-396.
17. Swenson, S.P. 1940. Genetic and cytological studies on a brachytic mutant in barley. *J. Agric. Res.* 60:687-713.
18. Szarejko, I., and M. Maluszynski. 1984. New brachytic mutant of spring barley variety Aramir. *Barley Genet. Newsl.* 14:33-35.
19. Tsuchiya, T. 1974. Allelic relationships of genes for short-awned mutants in barley. *Barley Genet. Newsl.* 4:80-81.
20. Walker, G.W.R., J. Dietrich, R. Miller, and K.J. Kasha. 1963. Recent barley mutants and their linkages II. Genetic data for further mutants. *Can. J. Genet. Cytol.* 5:200-219.

Prepared:

T. Tsuchiya and T.E. Haus. 1971. *Barley Genet. Newsl.* 1:104.

Revised:

T. Tsuchiya. 1980. *Barley Genet. Newsl.* 10:100.
J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:44.
J.D. Franckowiak and L.S. Dahleen. 2007. *Barley Genet. Newsl.* 37:188-189.
J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:58-59.
J.D. Franckowiak. 2013. *Barley Genet. Newsl.* 43:48-49.
J.D. Franckowiak and U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:50-52.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 51, Rattail spike 1, *rtt1*

Stock number: BGS 51
Locus name: Rattail spike 1
Locus symbol: *rtt1*

Previous nomenclature and gene symbolization:

Rattail spike = *rt* (2).

Inheritance:

Monofactorial recessive (1).

Located in chromosome 2HS (1), *rtt1.a* is about 6.0 cM distal from the *fch15* (chlorina seedling 15) locus (1), in 2H bin 05 or 06.

Description:

Spikes are highly modified with numerous immature spikelets and complete sterility. The arrangement of bracts in the spikelets suggests that they contain numerous spikelets. Because the homozygous recessive is completely sterile, the stock must be maintained in the heterozygous condition (1). A photograph of the rattail spikes is available (4, 5). A semidominant interaction between *rtt1.a* and one version of the normal allele (*Rt'* or *Rtt1.b*) in Okaiku 3 was reported by Takahashi et al. (3).

Origin of mutant:

A spontaneous mutant in Goldfoil (PI 5975) (1).

Mutational events:

rtt1.a (GSHO 216) in Goldfoil (PI 5975) (1), *Rtt1.b* (*Rt'*) in Okaiku 3 (OUL011) (3).

Mutant used for description and seed stocks:

rtt1.a (GSHO 216) in Goldfoil.

References:

1. Robertson, D.W. 1967. Linkage studies of various barley mutations (*Hordeum* species). Crop Sci. 7:41-42.
2. Robertson, D.W., G.A. Wiebe, and R.G. Shands. 1947. A summary of linkage studies in barley: Supplement I, 1940-1946. J. Am. Soc. Agron. 39:464-473.
3. Takahashi, R., J. Hayashi, and I. Moriya. 1976. New find of an allele *Rt'* semi-dominant over *rt* for rattail spike. Barley Genet. Newsl. 6:74.
4. Reid, D.A., and G.A. Wiebe. 1979. Taxonomy, botany, classification, and world collection. p. 79-104. In Barley: Origin, botany, culture winterhardiness, genetics, utilization, pests. USDA Agric. Handb. No. 338.
5. www.nordgen.org/bgs.

Prepared:

T. Tsuchiya and T.E. Haus. 1971. Barley Genet. Newsl. 1:111.

Revised:

J.D. Franckowiak. 2017. Barley Genet. Newsl. 47:53.

BGS 65, Early maturity 1, *Eam1*

Stock number: BGS 65
Locus name: Early maturity 1
Locus symbol: *Eam1*

Previous nomenclature and gene symbolization:

Early maturity = *Ea* (8).
Early maturity 1 = *Ea1* (7).
Photoperiod response gene H1 = *Ppd-H1* (11).

Inheritance:

Monofactorial dominant (9, 13).
Located in chromosome 2HS (1, 8, 13); weakly linked to the *vrs1* (six-rowed spike 1) locus (1, 15, 19); the *Ppd-H1* (*Eam1*) locus is in the 6 cM interval between the RFLP loci XMWG858 and XpsrB9, 1 cM proximal to XM WG858 (11, 12); specific sequence tagged site (STS) markers were developed for the *Eam1* gene of Igri (3); in bin 2H-04 near RFLP marker MWG858 (13); in Azumamugi 6 cM proximal from marker ABG602 (15), *Eam1.c* is associated with SNP markers 1_0216 to 2_1338 (positions 47.48 to 66.78 cM) in 2H bins 04 to 06 of the Bowman backcross-derived line BW280 (4); *Eam1.d* is associated with SNP markers 12_1386 to 2_1261 (position 50.56 cM) in 2H bin 05 of the Bowman backcross-derived line BW281 (4); *Eam1.f* is associated with SNP markers 1_0216 to 2_1015 (positions 47.48 to 48.68 cM) in 2H bin 04 of the Bowman backcross-derived line BW282 (4), likely in 2H bin 05.

Description:

This gene controls the strong response of some cultivars to long photoperiods. The *Eam1* gene is described as a simply inherited dominant and the main factor for early maturity (1, 7, 8, 14). Under long-day condition, a near-isogenic line with *Eam1* is reported to be 7 to 8 days earlier than the *eam1* line 'ea1' (16). The early maturity gene is weakly linked to the *vrs1* (six-rowed spike 1) locus (1, 13, 19). In North Dakota, USA, progenies from crosses of Bowman to *Hordeum vulgare* subsp. *spontaneum*, California Coast and North African accessions, facultative winter barleys, far northern spring barleys (from Finland and Alaska), and winter barleys, extremely early segregates were found (6). The *Eam1* locus in barley is homoeologous to the *Ppd1*, *Ppd2*, and *Ppd3* loci in wheat, which control strong photoperiod response (11, 12). The reduced photoperiod responsiveness of the *ppd-H1* (*eam1.a*) mutant is explained by altered circadian expression of the photoperiod pathway gene CONSTANS (CO) (10, 19) and reduced expression of its downstream target, FLOWERING LOCUS T (*HvFT1*), a key regulator of flowering (2, 18). The CCT (CO, CO-like, and TOC1) domain mutation was a G-to-T change and produced a Gly-to-Trp change in the CCT domain of the barley pseudo-response regulator (PRR) gene (18). The DNA code sequence of the dominant alleles was variable among accessions while the recessive allele was not (18). Alleles at the *Eam1* locus interact with flowering time control mutants at the *eam7* (17) and *eam10* (2), but not *eam8* (5), loci to induce earlier heading under both short and long days.

Origin of mutant:

The strong photoperiod response to short nights occurs naturally in many barley cultivars and its wild relative, *Hordeum vulgare* subsp. *spontaneum*.

Mutational events:

A single dominant gene controls early maturity based on a cross to Lion (*Eam1.b*) (PI

32767, GSHO 27) (8); *Eam1.e* (GSHO 1569) was isolated from the Australian cultivar Prior (PI 221325) (9, 16); *eam1.a* is the normal allele in Manchurian six-rowed cultivars (7, 16); *eam1.a* is the allele present likely in many two-rowed cultivars from Europe origin (16).

Mutant used for description and seed stocks:

Eam1.c in Estate (PI 57700, GSHO 1316); *Eam1.c* from Estate in Bowman (PI 483237)*7 (GSHO 1871); *Eam1.c* from Estate in Bowman*8 (BW280, NGB 20564); *Eam1.d* from KT1031 (a winter barley from Bulgaria, GSHO 1568) in Bowman*9 (GSHO 1872, BW281, NGB 20565); *Eam1.e* from Prior in Betzes*7 (Erbet, Clho 13826) (4); *Eam1.f* from *Hordeum vulgare* subsp. *spontaneum* in Bowman*8 (GSHO 1873, BW282, NGB 20566).

References:

1. Barbacki, S. 1930. [Studies in barley. II. Variability and inheritance of some physiological characters. The importance of breeding characters]. Men. Inst. Nat. Polon. Ec. Rur. Pulawy 11:579-610. [In Polish with English summary.]
2. Campoli, C., A. Pankin, B. Drosse, C.M. Casao, S.J. Davis, and M. von Korff. 2013. *HvLUX1* is a candidate gene underlying the *early maturity 10* locus in barley: phylogeny, diversity, and interactions with the circadian clock and photoperiodic pathways. New Phytologist 199:1045-1059.
3. Decousset, L., S. Griffiths, R.P. Dunford, N. Pratchett, and D.A. Laurie. 2000. Development of STS markers closely linked to the *Ppd-H1* photoperiod response gene of barley (*Hordeum vulgare* L.). Theor. Appl. Genet. 101:1202-1206.
4. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. Plant Physiol. 155:617-627.
5. Faure, S., A.S. Turner, D. Gruszka, V. Christodoulou, S.J. Davis, M. von Korff, and D.A. Laurie. 2012. Mutation at the circadian clock gene *EARLY MATURITY 8* adapts domesticated barley (*Hordeum vulgare*) to short growing seasons. Proc. Natl. Acad. Sci., USA 109: 8328-8333.
6. Franckowiak, J.D. (Unpublished).
7. Franckowiak, J.D. 1992. Mapping a gene for photoperiod sensitivity in barley. Agron. Abstr. 1992:96.
8. Griffée, F. 1925. Correlated inheritance of botanical characters in barley, and the manner of reaction to *Helminthosporium sativum*. J. Agric. Res. 30:915-935.
9. Hockett, E.A., and R.F. Eslick. 1972. Registration of Erbet barley (reg. no. 127). Crop Sci. 12:712-713.
10. Laurie, D.A. 1997. Comparative genetics of flowering time. Plant Mol. Biol. 35:167-177.
11. Laurie, D.A., N. Pratchett, J.H. Bezzant, and J.W. Snape. 1994. Genetic analysis of a photoperiod response gene on the short arm of chromosome 2 (2H) of *Hordeum vulgare* (barley). Heredity 72:619-627.
12. Laurie, D.A., N. Pratchett, J.H. Bezzant, and J.W. Snape. 1995. RFLP mapping of five major genes and eight quantitative trait loci controlling flowering time in a winter X spring barley (*Hordeum vulgare* L.) cross. Genome 38:575-585.
13. Mackie, W.W. 1926. Inheritance of resistance to *Helminthosporium californicum* in a cross between Chevalier barley, a resistant variety, and Abyssinian, a susceptible variety. Phytopathology 16:764. [Abstr.].
14. Neatby, K.W. 1929. An analysis of the inheritance of quantitative characters and

linkage in barley. *Sci. Agric.* 9:701-718.

15. Sameri, M., K. Takeda, and T. Komatsuda. 2006. Quantitative trait loci controlling agronomic traits in recombinant inbred lines from a cross of oriental- and occidental-type barley cultivars. *Breed. Science* 56:243-252.

16. Smail, V.W., R.F. Eslick, and E.A. Hockett. 1986. Effect of genetically and environmentally induced heading date differences on yield and adaptation of an isogenic barley pair. *Crop Sci.* 26:889-893.

17. Stracke, S., and A. Börner. 1998. Molecular mapping of the photoperiod response gene *ea7* in barley. *Theor. Appl. Genet.* 97:797-800.

18. Turner, A., J. Beales, S. Faure, R.P. Dunford, and D.A. Laurie. 2005. The pseudo-response regulator *Ppd-H1* provides adaptation to photoperiod in barley. *Science* 310:1031-1034.

19. Wexelsen, H. 1934. Quantitative inheritance and linkage in barley. *Hereditas* 17:307-348.

Prepared:

J.D. Franckowiak and L.W. Gallagher. 1997. *Barley Genet. Newsl.* 26:101-102.

Revised:

J.D. Franckowiak. 2014. *Barley Genet. Newsl.* 44:64-66.

J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:54-56.

Stock number: BGS 67
Locus name: Six-rowed spike 1 (deficiens spike)
Locus symbol: *Vrs1.t*

Revised locus symbol:

Historically, alleles at the six-rowed spike 1 (*vrs1*) locus were included in several Barley Genetic Stock (BGS) descriptions even though they were known to be alleles at the same locus. The most common spike type variant is the six-rowed allele (*vrs1.a*) (3, 6, 7). Other alleles at the *vrs1* locus are described in BGS 6, BGS 58, and BGS 66.

Previous nomenclature and gene symbolization:

Hordeum deficiens = *V* (5).

Hordeum deficiens = *V^t* (11).

Renamed *Vrs1.t1* based on allelism tests (7).

Mutation in homeodomain-leucine zipper I-class homeobox = *HvHox1* (3, 6).

Inheritance:

Monofactorial incomplete dominant (6); the *Vrs1.t1* allele is the most dominant allele in the multiple allelic series at the *vrs1* (six-rowed spike 1) locus (11), but interacts as an incomplete dominant with some alleles (2, 7).

Located in chromosome 2HL (8, 10); the *vrs1* locus is about 15.4 cM distal from the Toothed lemma 1 (*Gth1*) locus (10); *vrs1.a* is about 30.5 cM distal from the *eog1* (elongated outer glume 1) locus (8); *Vrs1.t1* is associated with SNP markers 2_0781 to 2_1351 (positions 135.19 to 145.80 cM) in 2H bins 09 to 10 of the Bowman backcross-derived line BW900 (1).

Description:

Plants homozygous for *Vrs1.t1* (*V^t*) allele always have a deficiens phenotype regardless of the allele present at the *int-c* (intermedium spike-c) locus in chromosome 4H.

Deficiens is characterized by the extreme reduction in size of lateral spikelets (11). The *Vrs1.t1* allele in Engleawnless does not recombine with the *Lks1.a* allele at the *Lks1* (Awnless 1) locus because a short paracentric inversion is present in this stock (9).

Plants in Bowman backcross line for *Vrs1.t1* (BW900) were similar to Bowman except spikes had 1 to 2 more kernels per spike and kernels were about 10% lighter (2).

Expression of the *Vrs1* gene is strictly localized in the lateral-spikelet primordia of immature spikes suggesting that the VRS1 protein suppresses development of lateral spikelets (3). Re-sequencing a 2.1 kb fragment region including the *HvHox1* region of the *vrs1* locus found that the six-rowed variants (*vrs1.a1*, *vrs1.a2* and *vrs1.a3*) and the deficiens variant (*Vrs1.t1*) had isoforms different from those of wild barleys and two-rowed barleys (6). Plants with *Vrs1.t1* gene had smaller lateral spikelet meristems compared with the typical two-rowed type (7). Transcriptome analysis revealed that a number of *histone* superfamily genes were upregulated in the *deficiens* mutant suggesting that enhanced cell proliferation in the central spikelet may contribute to larger kernel size (7).

Origin of mutant:

Natural occurrence in two-rowed barleys from Ethiopia previously classified as *Hordeum deficiens* (12). Haplotype analysis revealed that barley carrying the *deficiens* allele (*Vrs1.t1*) originated from two-rowed types carrying the *Vrs1.b2* allele, which is predominantly found in germplasm from Northern Africa (7).

Mutational events:

Vrs1.t1 (*V*¹) in White Deficiens (CIho 7316, GSHO 684) (7, 12); *Vrs1.t2* (*Deficiens* 2) (NGB 115174), *Vrs1.t3* (*Semi-deficiens* 12) (NGB 115188) in Foma (NGB 14659, CIho 11333) (4, 7).

Mutant used for description and seed stocks:

Vrs1.t1 (*V*¹) in White Deficiens (GSHO 684); *Vrs1.t1* in Bowman (PI 483237)*8 (GSHO 1909, BW900, NGB 22332).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Komatsuda, T., M. Pourkheirandish, C. He, P. Azhaguvel, H. Kanamori, D. Perovic, N. Stein, A. Graner, T. Wicker, A. Tagiri, U. Lundqvist, T. Fujimura, M. Matsuoka, T. Matsumoto, and M. Yano. 2007. Six-rowed barley originated from a mutation in a homeodomain-leucine zipper I-class homeobox gene. *Proc. Natl. Acad. Sci. USA* 104:1424-1429.
4. Lundqvist, U. (Unpublished).
5. Powers, L. 1936. The nature of the interaction of genes affecting four quantitative characters in a cross between *Hordeum deficiens* and *Hordeum vulgare*. *Genetics* 21:398-420.
6. Saisho, D., M. Pourkheirandish, H. Kanamori, T. Matsumoto, and T. Komatsuda. 2009. Allelic variation of row type gene *Vrs1* in barley and implication of the functional divergence. *Breed. Sci.* 59: 621-628.
7. Sakuma, S., U. Lundqvist, Y. Kakei, V. Thirulogachandar, T. Suzuki, K. Hori, J. Wu, A. Tagiri, T. Rutten, R. Koppolu, Y. Shimada, K. Houston, W.T.B. Thomas, R. Waugh, T. Schnurbusch, and T. Komatsuda. 2017. Extreme suppression of lateral floret development by a single amino acid change in the VRS1 transcription factor. *Plant Physiol.* 175:1720-1731.
8. Swenson, S.P., and D.G. Wells. 1944. The linkage relation of four genes in chromosome 1 of barley. *J. Am. Soc. Agron.* 36:429-435.
9. Tsuchiya, T., and R.J. Singh. 1972. Another case of paracentric inversion in a genetic stock, Engleawnless. *Barley Genet. Newsl.* 2:110-111.
10. Wexelsen, H. 1934. Quantitative inheritance and linkage in barley. *Hereditas* 18:307-308.
11. Woodward, R.W. 1947. The *l*^h, *l*, *i* alleles in *Hordeum deficiens* genotypes of barley. *J. Am. Soc. Agron.* 39:474-482.
12. Woodward, R.W. 1949. The inheritance of fertility in the lateral florets of the four barley groups. *Agron. J.* 41:317-322.

Prepared:

T.E. Haus. 1975. *Barley Genet. Newsl.* 5:105.

Revised:

J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:104.

J.D. Franckowiak and U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:57-58.

Stock number: BGS 69
Locus name: Toothed lemma 1
Locus symbol: *Gth1*

Previous nomenclature and gene symbolization:

Toothed lemma = G (5, 9).

Inheritance:

Monofactorial dominant (7, 8), but modifiers were reported (5, 7).

Located in chromosome 2HL (5, 9); *Gth1.a* is about 15.4 cM proximal from the *vrs1* (six-rowed spike 1) locus (2, 5, 9); *gth1.b* is associated with SNP markers 2_0528 to 1_0823 (positions 118.78 to 133.59 cM) in 2H bins 08 to 09 of the Bowman backcross-derived line BW413 (1); the association of palea trichome develop with spike type (4) suggests that alleles at *Gth1* locus also control this trait.

Description:

Large teeth or barbs are present on the upper part of lateral lemma nerves. The barbs are easiest to see on green spikelets. This trait may be difficult to study because Ubisch (8) reported three sizes of teeth, including one that could be seen only with magnification. According to Ubisch (8), one gene pair controls the presence and absence of large teeth and one or two other genes are responsible for less developed teeth. Segregation for only one pair of factors is reported by Wexelsen (9) and in subsequent studies. Other reports on inheritance of lemma teeth were summarized by Smith (6). Besides barbs on the lateral lemma veins and slight heavier kernels, no other agronomic or morphological differences were observed between Bowman and the Bowman backcross-derived line for *gth1.b*, BW413 (3). Association of trichomes with vascular bundle traces may not be limited to the lemma because similar variations in shape of trichomes were observed on the palea (4). Trichomes with a round base and a domed peak appeared predominantly on two-rowed cultivars. But most of the six-rowed cultivars featured trichomes with a similar base, but a prickly-like point that aligned with the direction of elongation of the epidermal cells (4). The palea trichomes may play a role in infection by the scab fungus, *Fusarium graminearum*, because the prickly-type trichomes functioned to trap conidia and were sites of fungal penetration (4).

Origin of mutant:

Gth1.a appears to be the normal allele; while the recessive allele *gth1.b* is present in many two-rowed barley cultivars of Occidental origin (4).

Mutational events:

Gth1.a (GSHO 309) in Machine (Clho 4982) (9); *gth1.b* is present in many two-rowed cultivars of European origin (4).

Mutant used for description and seed stocks:

Gth1.a (GSHO 309) in Machine; *Gth1.a* is present in Bowman (PI 483237); *gth1.b* from ND11894 (Gitane//ND4642/Robust/3/Bearpaw/4/TR218/ND7015/3/ Norbert//ND4856/M38) in Bowman*4 (GSHO 1897); *gth1.b* in Bowman*6 (BW413, NGB 20646).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Finch, R. A., and E. Simpson. 1978. New colours and complementary colour genes in

barley. Z. Pflanzenzücht. 81:40-53.

3. Franckowiak, J.D. (Unpublished).

4. Imboden, L., D. Afton, and F. Trail. 2017. Surface interactions of *Fusarium graminearum* on barley. Mol. Plant Path. 19:1332-1342.

5. Karpechenko, G.D., and K.V. Ivanova. 1932. Linkage of genes I and G in barley. Bull. Lab. Genet. Leningrad 9:97-108. [In Russian with English summary.]

6. Smith, L. 1951. Cytology and genetics of barley. Bot. Rev. 17:1-51, 133-202, 285-355.

7. Tschermak, E. von. 1901. Über Züchtung neuer Getreiderassen mittelst künstlicher Kreuzung. Kritisch-historische Betrachtungen. Zeitschrift für das landwirtschaftliche Versuchswesen Oesterreich 4:1029-1060.

8. Ubisch, G. von. 1919. Beitrag zu einer Faktorenanalyse von Gerste. II. Z. Indukt. Abstammungs. Vererbungs. 20:65-117.

9. Wexelsen, H. 1934. Quantitative inheritance and linkage in barley. Hereditas 18:307-348.

Prepared:

T.E. Haus. 1975. Barley Genet. Newsl. 5:109.

Revised:

T.E. Haus. 1978. Barley Genet. Newsl. 8:161.

J.D. Franckowiak. 1997. Barley Genet. Newsl. 26:106.

J.D. Franckowiak. 2014. Barley Genet. Newsl. 44:68-69.

J.D. Franckowiak. 2017. Barley Genet. Newsl. 47:59-60.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 79, White streak 7, *wst7*

Stock number: BGS 79
Locus name: White streak 7
Locus symbol: *wst7*

Previous nomenclature and gene symbolization:

Ribbon grass = *rb* (7).

White streak-k = *wst*,*k* (11).

White streak-B = *wst*,*B* (9).

Inheritance:

Monofactorial recessive (2, 12).

Located in chromosome 2HL (6, 8, 9, 10); *wst7.k* is about 22.0 cM distal from the *gpa1* (grandpa 1) locus (2, 10); *wst7.k* is over 29.4 cM distal from the *lig1* (liguleless 1) locus (9); *wst7.k* is about 6.1 cM from RFLP marker MWG949A (1); *wst7.k* is associated with SNP markers 2_1346 and 1_283 (positions 233.94 and 234.63 cM) in 2H bin 14 of the Bowman backcross-derived line BW916 (3), in 2H bin 14.

Description:

Vertical white streaks of variable width and number develop in the leaf blades of young secondary tillers. Fewer white streaks and fewer tillers with white streaks occur as environmental conditions become warm. White streaks can be found until near maturity, but they are difficult to observe after heading under field conditions. Often the lower or first leaves on early tillers have more and wider streaks. There were no apparent effects of the *wst7.k* gene on agronomic traits in the Bowman backcross-derived line BW916 (5).

Origin of mutant:

A spontaneous mutant isolated by Robertson (7, 12).

Mutational events:

wst7.k (GSHO 247) in an unknown cultivar (2, 12).

Mutant used for description and seed stocks:

wst7.k (GSHO 247) in an unknown cultivar; *wst7.k* from R.I. Wolfe's Multiple Recessive Marker Stock (GSHO 3451) in Bowman (PI 483237)*7 (GSHO 1935, BW916, NGB 22347).

References:

1. Costa, J.M., A. Corey, M. Hayes, C. Jobet, A. Kleinhofs, A. Kopisch-Obusch, S.F. Kramer, D. Kudrna, M. Li, O. Piera-Lizaragu, K. Sato, P. Szues, T. Toojinda, M.I. Vales, and R.I. Wolfe. 2001. Molecular mapping of the Oregon Wolfe Barleys: a phenotypically polymorphic doubled-haploid population. *Theor. Appl. Genet.* 103:415-424.
2. Doney, D.L. 1961. An inheritance and linkage study of barley with special emphasis on purple pigmentation or the auricle. M.S. Thesis. Utah State Univ., Logan.
3. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
4. Franckowiak, J.D. 1996. Coordinator's report: Chromosome 2. *Barley Genet. Newsl.* 25:88-90.
5. Franckowiak, J.D. (Unpublished).
6. Kasha, K.J. 1982. Coordinator's report: Chromosome 6. *Barley Genet. Newsl.* 12:90-92.

7. Robertson, D.W., G.A. Wiebe, and R.G. Shands. 1947. A summary of linkage studies in barley: Supplement I, 1940-1946. *J. Am. Soc. Agron.* 39:464-473.
8. Schondelmaier, J., G. Fischbeck, and A. Jahoor. 1993. Linkage studies between morphological and RFLP markers in the barley genome. *Barley Genet. Newsl.* 22:57-62.
9. Shin, J.S., S. Chao, L. Corpuz, and T.K. Blake. 1990. A partial map of the barley genome incorporating restriction fragment length polymorphism, polymerase chain reaction, isozyme, and morphological marker loci. *Genome* 23:803-810.
10. Walker, G.W.R. 1974. Linkage data for *rb* and *mt3*. *Barley Genet. Newsl.* 4:90-91.
11. Wolfe, R.I., and J.D. Franckowiak. 1991. Multiple dominant and recessive genetic marker stocks in spring barley. *Barley Genet. Newsl.* 20:117-121.
12. Woodward, R.W. 1957. Linkages in barley. *Agron. J.* 49:28-32.

Prepared:

J.D. Franckowiak and R.I. Wolfe. 1997. *Barley Genet. Newsl.* 117.

Revised:

J.D. Franckowiak. 2007. *Barley Genet. Newsl.* 37:207-208.

J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:87-88.

J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:61-62.

Stock number: BGS 82
Locus name: Zeocriton 1
Locus symbol: Zeo1

Revised locus symbols:

Since the zeocriton spike phenotype was first described by Hayes and Harlan (8), *Zeo1* became the recommended locus symbol for various mutants at this locus. The semi-dominant dense spike variants at the complex *Zeo1* locus exhibit a range of phenotypes associated with restricted elongation of the rachis internodes. Based on DNA sequencing, three distinct phenotypic groups of dense spike variants are associated with the *Hordeum vulgare* *APELATA2* (*AP2*)-like transcription factor (*HvAP2*) locus (10, 13). The phenotypic expressions were historically assigned different locus symbols, but each group is associated with specific molecular changes in the *HvAP2* transcript (10). Alleles at this locus are assigned locus symbols: The *Zeo1* mutants exhibit the most extreme phenotypes, the *Zeo2* variants have intermediate phenotypes, and the *Zeo3* variants show a lesser degree in shortening of rachis internodes (10). Molecular markers and/or pedigree information are necessary to separate variants assigned to the *Zeo2* (BGS614) and *Zeo3* (BGS184) groups (10).

Previous nomenclature and gene symbolization:

Erectoides-52 = *ert-52* (6, 7).

Erectoides-r = *Ert-r* (12).

"Kurz und dicht" = *Knd* (15, 18).

Cleistogamy = *Cly* (9, 10, 13, 17).

Hordeum vulgare *APELATA2* (*AP2*)-like transcription factor = *HvAP2* (10).

Inheritance:

Monofactorial incomplete dominant (6, 14).

Located in chromosome 2HL (9, 11, 17); *Zeo1.a* is about 9.2 cM distal from the *lig1* (liguleless 1) locus (11); *Zeo1.a* is about 7.3 cM distal from RFLP marker *cnx1* in 2H bin 13 (1); the *Ert-r.52* allele in Bowman backcross-derived line BW322 is associated with SNP markers 2_0715 to 1_0551 (positions 213.08 to 221.70 cM) in 2H bin 13 (2); the *Zeo1.a* allele in BW937 is associated with SNP markers 2_0715 to 2_1453 (positions 213.08 to 245.71 cM) in 2H bin 13/14 (2); the *Zeo1.b* allele in BW938 is associated with SNP markers 1_1486 to 2_0590 (positions 202.70 to 218.47) in 2H bin 13 (2); the *HvAP2* locus is near marker BOPA2_12_10579 (10), in 2H bin 13.

Description:

The name zeocriton (little barley) has been historically applied to cultivars with very short rachis internodes and a pyramid shape. This name and the locus symbol *Zeo1* were applied to induced mutants having a similar phenotype (18). Homozygous *Zeo1* plants have shorter culms (short peduncle), very compact spikes, large outer glumes with long awns, and reduced fertility. Plants heterozygous for the *Zeo1* mutant have short culms, compact spikes, and wide kernels. (18). Generally, the spike emerges from the side of the sheath in homozygotes (5, 18). Although the name *zeocriton* is used for this gene, this gene is not from Spratt, the dense ear type described by Engledow (3) or those described by Hayes and Harlan (8). Spikes of plants with the *Ert-r.52* mutant are compact in heterozygotes and very compact in homozygotes, with rachis internode length values from 1.4 to 1.8 mm. Homozygotes are about 2/3 normal height with

excellent vigor. The glumes associated with lateral spikelets are 3 to 4 times larger than normal. Lodicule size is reduced (14). Heterozygotes are intermediate in plant height, have slightly more lax spikes, and have normal glumes in lateral spikelets (14). GA₃ treatment of plants as the flag leaf emerges decreases spike density (16). Plants in Bowman backcross-derived lines for *Ert-r.52* (BW322), *Zeo1.a* (BW937), and *Zeo1.b* (BW938) are about 2/3 the height of Bowman. Rachis internodes are about 1/2 normal length (2.2 vs. 4.4 mm). Mutant plants head about two days later than Bowman; they have about 2 more kernels per spike and their grain yields are about 3/4 of those of Bowman (5). The BW938 (*Zeo1.b*) averaged one more fertile rachis node than Bowman (10). DNA sequencing showed that the *Zeo1* mutants occur in a *Hordeum vulgare* APELATA2 (*AP2*)-like transcription factor, *HvAP2* (10, 13). The dense spike and cleistogamous (small lodicules) phenotypes are a consequence of a perturbed interaction between microRNA 172 (*Hv-miR172*) and its corresponding binding site on the mRNA from the *HvAP2* gene, which acts early in spike development to regulate turnover of *HvAP2* mRNA (10, 13). The *Zeo1* mutants occur in the last intron of *HvAP2*, the binding site of *Hv-miR172*, and prevent cleavage of the *HvAP2* mRNA (10, 13).

Origin of mutant:

An X-ray induced mutant in Donaria (PI 161974) (15); a neutron induced mutant in Bonus (NGB 14657, PI 189763) (6).

Mutational events:

Zeo1.a (Mut 2657, GSHO 1613) in Donaria (PI 161974) (15) has haplotype 4 in the *HVAP2* gene (10); *Zeo1.b*, received as "Kurz und dicht" and placed in R.I. Wolfe's Multiple Dominant Marker Stock (GSHO 1614) (4, 18) has haplotype 1 (10); *Ert-r.52* (NGB 112651, GSHO 492), *-r.67* (NGB 112666) in Bonus (NGB 14657, PI 189763) (12) have haplotype 1 (10); *Ert-r.329* (NGB 112844) in Foma (NGB 14659, Clho 11333) (14) has haplotype 1 (10); *Ert-r.453* (NGB 112968) in Foma (12) has haplotype 2 (10). Other variants in the *HvAP2* gene are assigned to the *Zeo2* (BGS614) or *Zeo3* (BGS184) groups (10).

Mutant used for description and seed stocks:

Zeo1.a in Donaria (GSHO 1613); *Zeo1.a* in Bowman (PI 483237)*5 (GSHO 1931); *Zeo1.a* in Bowman*7 (BW937, NGB 22366); *Zeo1.b* from R.I. Wolfe's Multiple Dominant Marker Stock (GSHO 1614) in Bowman*9 (GSHO 1932, BW938, NGB 22367); *Ert-r.52* (*Zeo1.a*) in Bonus (GSHO 492, NGB 112651); *Ert-r.52* in Bowman (PI 483237)*8 (GSHO 2123, BW322, NGB 22117).

References:

1. Costa, J.M., A. Corey, M. Hayes, C. Jobet, A. Kleinhofs, A. Kopisch-Obusch, S.F. Kramer, D. Kudrna, M. Li, O. Piera-Lizaragu, K. Sato, P. Szücs, T. Toojinda, M.I. Vales, and R.I. Wolfe. 2001. Molecular mapping of the Oregon Wolfe Barleys: a phenotypically polymorphic doubled-haploid population. *Theor. Appl. Genet.* 103:415-424.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
3. Engledow, F.L. 1924. Inheritance in barley. III. The awn and the lateral floret (cont'd): fluctuation: a linkage: multiple allelomorphs. *J. Genet.* 14:49-87.
4. Franckowiak, J.D. 1992. Allelism tests among selected semidwarf barleys. *Barley Genet. Newsl.* 21:17-23.
5. Franckowiak, J.D. (Unpublished).
6. Hagberg, A., Å. Gustafsson, and L. Ehrenberg. 1958. Sparsely contra densely ionizing

radiations and the origin of erectoid mutants in barley. *Hereditas* 44:523-530.

7. Hagberg, A., G. Persson, and A. Wiberg. 1963. Induced mutations in the improvement of self-pollinated crops. p. 105-124. *In* E. Åkerberg and A. Hagberg (eds.) *Recent Plant Breeding Research. Svalöf 1946-1961*. Almqvist & Wiksell, Stockholm.

8. Hayes, H.K., and H.V. Harlan. 1920. The inheritance of the length of internode in the rachis of the barley spike. U.S. Dept. Agr., Bull. 869. 26 pp.

9. Honda, I., Y. Turuspekov, T. Komatsuda, and Y. Watanabe. 2005. Morphological and physiological analysis of cleistogamy in barley (*Hordeum vulgare*). *Physiol. Plant* 124:524-531.

10. Houston, K., S.M. McKim, J. Comadran, N. Bonar, I. Druka, N. Uzrek, E. Cirillo, J. Guzy-Wrobel, N.C. Collins, C. Halpin, M. Hansson, C. Dockter, A. Druka, and R. Waugh. 2013. Variation in the interaction between alleles of *HvAPETALA2* and microRNA172 determines the density of grains on the barley inflorescence. *Proc. Natl. Acad. Sci. USA* 110:16675-16680.

11. Luna Villafañá, A., and J.D. Franckowiak. 1995. (Unpublished).

12. Lundqvist, U., and J.D. Franckowiak. 1997. BGS 332, Erectoides-r, Ert-r. *Barley Genet. Newsl.* 26:280.

13. Nair, S.K., N. Wang, Y. Turuspekov, M. Pourkheirandish, S. Sinsuwongwat, G. Chen, M. Sameri, A. Tagiri, I. Honda, Y. Watanabe, H. Kanamori, T. Wicker, N. Stein, Y. Nagamura, T. Matsumoto, and T. Komatsuda. 2010. Cleistogamous flowering in barley arises from the suppression of microRNA-guided *HvAP2* mRNA cleavage. *Proc. Natl. Acad. Sci. USA* 107:490-495.

14. Persson, G., and A. Hagberg. 1969. Induced variation in a quantitative character in barley. Morphology and cytogenetics of *erectoides* mutants. *Hereditas* 61:115-178.

15. Scholz, F., and O. Lehmann. 1959. Die Gaterslebener Mutanten der Saatgerste in Beziehung zur Formenmannigfaltigkeit der Art *Hordeum vulgare* L.s.l. I. Kulturpflanze 6:123-166.

16. Stoy, V., and A. Hagberg. 1967. Effects of growth regulators on ear density mutants in barley. *Hereditas* 58:359-384.

17. Turuspekov, Y., Y. Mano, I. Honda, N. Kawada, T. Watanabe, and T. Komatsuda. 2004. Identification and mapping of cleistogamy genes in barley. *Theor. Appl. Genet.* 109:480-487.

18. Wolfe, R.I. (Unpublished).

Prepared:

J.D. Franckowiak and R.I. Wolfe. 1997. *Barley Genet. Newsl.* 26:120.

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:280. as BGS 332, Erectoides-r, Ert-r.

Revised:

J.D. Franckowiak. 2007. *Barley Genet. Newsl.* 37:209.

J.D. Franckowiak and U. Lundqvist. 2011. *Barley Genet. Newsl.* 41:89-90.

J.D. Franckowiak and U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:63-65.

BGS 89, Breviaristatum-g, *ari-g*

Stock number: BGS 89
Locus name: Breviaristatum-g
Locus symbol: *ari-g*

Previous nomenclature and gene symbolization:

Breviaristatum-18 = *ari-18* (4, 5).

Short awn 10 = *lk10* (7).

Inheritance:

Monofactorial recessive (5).

Located in chromosome 2H (1, 3); *ari-g.24* is associated based on linkage drag with the *Gth1* (Toothed lemma 1) locus (1), but the association may be caused by poor expression of the *Gth1.a* allele in the Bowman backcross-derived line, BW045 (2, 3); *ari-g.24* is associated with SNP markers 1-0796 to 2_0528 (positions 95.53 to 118.78 cM) in 2H bins 07 to 08 of a heterozygous plant from the Bowman backcross-derived line BW045 (1).

Description:

Plants with the *ari-g* mutation are semidwarf and very weak. Under greenhouse conditions, plants are 1/4 to 1/3 normal height and stiff, spikes are short and small, and seed set is low. Leaf blades are very narrow, often involuted, and are white and green striped (5, 6). When grown in the field at Fargo, North Dakota, USA, plant development is very slow, the upper portion of the sheath is white as new leaf blades emerge, and spikes are seldom produced. Thus, the Bowman backcross-derived line for *ari-g.24*, BW045, is maintained as a segregating stock (2).

Origin of mutant:

An X-ray induced mutant in Bonus (NGB 14657, PI 189763) (5).

Mutational events:

ari-g.18 (NGB 115865) in Bonus (NGB 14657, PI 189763) (5); *ari-g.24* (NGB 115872, GSHO 1655) in Bonus (6); *ari-g.134* (NGB 115944), *-g.138* (NGB 115948), *-g.142* (NGB 115952), *-g.158* (NGB 115968), *-g.221* (NGB 116030), *-g.234* (NGB 116044), *-g.246* (NGB 116056) in Foma (NGB 14659, CIho 11333), *-g.267* (NGB 116078) in Kristina (NGB14661, NGB 1500) (5); *ari-g.303* (NGB 116127) in Kristina (6).

Mutant used for description and seed stocks:

ari-g.24 in Bonus (NGB 115872, GSHO 1655); *ari-g.24* in Bowman*8 (BW045, NGB 20453).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Franckowiak, J.D. 1995. Notes on linkage drag in Bowman backcross derived lines of spring barley. *Barley Genet. Newsl.* 24:63-70.
4. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
5. Kucera, J., U. Lundqvist, and Å. Gustafsson. 1975. Inheritance of breviaristatum mutants in barley. *Hereditas* 80:263-278.
6. Lundqvist, U. (Unpublished).

Barley Genetics Newsletter (2017): 46:50-187.

7. Tsuchiya, T. 1974. Allelic relationships of genes for short-awned mutants in barley. *Barley Genet. Newsl* 4:80-81.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:128.

Revised:

U. Lundqvist and J.D. Franckowiak. 2014. *Barley Genet. Newsl.* 44:79-80.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:66-67.

BGS 97, Accordion rachis 1, *acr1*

Stock number: BGS 97
Locus name: Accordion rachis 1
Locus symbol: *acr1*

Phenotype explanation:

The accordion rachis phenotype was developed by Robert I. Wolfe at the Beaverlodge, Alberta Station of Agriculture and Agri-Food Canada. The F2 progenies from crosses and backcrosses of Bowman to this stock, ACBV89B229, contained very few plants with the accordion rachis phenotype. Examination of the SNP molecular data for the Bowman backcross-derived line, BW009 (Bowman*7/ACBV89B229), identified three chromosomal regions that were retained from the donor parent (1). Only a region of 2HL near the centromere was not present in other BW lines with elongated rachis internodes. This region was assumed to be where the *acr1.a* allele is located. The other regions were present in BW lines showing a dominant inheritance pattern for slightly elongated rachis internodes. The locus in the retained 4HL region was named Accordion rachis 2, *Acr2*, and the locus in the 1HL region was named Accordion rachis 3, *Acr3*.

Previous nomenclature and gene symbolization:

Lax rachis internode = *ril* (2).

Accordion rachis = *acr* (5, 6).

Inheritance:

Monofactorial recessive (2, 5, 6), part of a multiple genic series involving *acr1*, *Acr2*, and *Acr3* required for expression (3).

Located in chromosome 2HL (3); *acr1.a* is near the *vrs1* (six-rowed spike 1) locus because no recombination with the *Vrs1.t* (deficiens) allele was observed (3); *acr1.a* is associated with SNP markers 1_0525 to 2_0667 (positions 65.03 to 117.73 cM) in 2H of the Bowman backcross-derived lines BW009 and BW439 (1), likely in 2H bins 06 to 08.

Description:

Rachis internodes are greatly elongated and often bent or pleaded as the spike emerges from the boot or sheath of the flag leaf (5). The line ACBV89B229, developed by R.I. Wolfe to maximize rachis internode length, exhibits extreme elongation of rachis internodes, rachis internode length values up to 7.7 mm, and occasionally trapping of the spike tip in the boot (3, 7). Elongation of the rachis internodes is associated with slightly elongated outer glumes and the deficiens (*Vrs1.t*) spike phenotype. Two dominant modifiers, *Acr2* (see BGS 189) and *Acr3* (see BGS 241), caused variable expression of the accordion trait in different genetic backgrounds. In crosses to Bowman, segregation for *acr1* fit a three gene model based on DNA segments retained in the Bowman backcross-derived lines BW009 and BW439 (1, 3). Although the *acr1* gene is apparently associated with the deficiens spike type, the large centromeric segment of 2H retained does not overlap the six-rowed spike 1 (*vrs1*) locus (1). A pericentric inversion cannot be eliminated as a possibility. Plants in the BW009 and BW439 lines were about 3/4 of normal height and peduncles were about 1/2 of normal length. The number of fertile rachis nodes was reduced by about 3 and heading was delayed by up to 4 days. Kernels appeared thinner and weighted about 10% less. Test weights were low and grain yield was about 3/4 of normal (3).

Origin of mutant:

A naturally occurring deviant (4) incorporated into several genetic stocks, *acr1.a* in

Beaverlodge stock ACBV89B229 was used to study this locus (3, 7).

Mutational events:

acr1.a in ACBV89B229 (GSHO 1617) and ACBV89232 (GSHO 1573) (7).

Mutant used for description and seed stocks:

acr1.a in ACBV89B229 (GSHO 1617); *acr1.a* in Bowman (PI 483237)*4 (GSHO 1899);

acr1.a in Bowman*7 (BW009, NGB 20417 and BW439, NGB 20671).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Enns, H. 1961. Inheritance and linkage studies in barley using chromosomal interchanges and marker stocks. Ph.D. Thesis. Univ. Saskatchewan. pp. 70.
3. Franckowiak, J.D. (Unpublished).
4. Franckowiak, J.D. 1995. Notes on linkage drag in Bowman backcross derived lines of spring barley. *Barley Genet. Newsl.* 24:63-70.
5. Haus, T.E. 1957. Inheritance of rachis internode length and an abnormal rachis type in barley. *Dissertation Abstr.* 49:179-180.
6. Nilan, R.A. 1964. The cytology and genetics of barley, 1951-1962. *Monogr. Suppl.* 3, *Res. Stud.* Vol. 32, No. 1. Washington State Univ. Press, Pullman.
7. Wolfe, R.I. 1990. (Personal communications).

Prepared:

J.D. Franckowiak. 2002. *Barley Genet. Newsl.* 32:85.

Revised:

J.D. Franckowiak. 2010. *Barley Genet. Newsl.* 40:56-57.

J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:68-69.

Stock number: BGS 102
Locus name: Uzu 1 (semi-brachytic)
Locus symbol: *uzu1*

Previous nomenclature and gene symbolization:

Normal vs *uzu* = *h* (28).
Uzu = *u* (15).
Uzu (semi-brachytic) = *uz* (27).
Uzu 2 = *uz2* (13, 30, 32).
Uzu 3 = *uz3* (13, 30, 32).
Hordeum vulgare BR-insensitive 1 = *HvBRI1* (3).
Erectoides-79 = *ert-79* (11).
Breviaristatum-256 = *ari-256* (14).
Hordeum vulgare DWARF = *HvDWARF* (7, 8).
Brassinosteroid deficient 1 = *brd1* (7).

Inheritance:

Monofactorial recessive (15, 23, 25, 27).
Located in chromosome 3HL (21, 22, 27); *uzu1.a* is about 17.6 cM proximal from the *alm1* (albino lemma 1) locus (26); *uzu1.a* is in bin 3H 06 near cDNA marker C1271 (3); *uzu1.a* is about 10.1 cM from AFLP marker E3733-6 in subgroup 27 of the Proctor/Nudinka map (19); *uzu1.a* is associated with SNP markers 1_0373 to 1_1314 (positions 92.55 to 107.40 cM) in 3H bins 06 to 07 of the Bowman backcross-derived line BW885 (5); *uzu1.a* with *sld1.a* (slender dwarf 1) is associated with SNP markers 1_0653 to 2_0115 (positions 92.55 to 126.83 cM) in 3H bins 06 to 08 of the Bowman line BW912 (5); *uzu1.a* with *wst1.c* (white streak 1) is associated with SNP markers 1_1258 to 2_0155 (positions 79.88 to 229.92 cM) in 3H bins 05 to 15 of the Bowman line BW860 (5); *ert-ii.79* is associated with SNP markers 2_0686 to 2_0931 (positions 67.01 to 104.39 cM) in 3H bins 05 to 06 of the Bowman line BW312 (5); *uzu1.256* (formerly *ari.256*) is associated with SNP markers 1_0728 to 2_1405 (positions 96.75 to 187.28 cM) in 3H bins 06 to 12 of Bowman line BW033 (5); the *ert-ii.79* allele at the *uzu1* (*HvBRI1*) locus is positioned at 57.1 cM (4) on the barley genome map (29), in 3H bin 06.

Description:

The *uzu1.a* gene has pleiotropic effects on the elongation of the coleoptile, leaf, culm, rachis internode, awn, glume, and kernel (24, 25, 27). These organs are often reduced in length and increased in width. Changes in organ length are temperature sensitive, but heading date and maturity are unaltered. The coleoptile of *uzu* plants shows a prominent projection or hook near the apex. Sometimes the coleoptile of the mutant shows a V-shaped notch on the side opposite from the projection. Thus, the apex of the coleoptile has two notches, one on each side (25, 30, 31). The temperature sensitive reduction in culm length of *uzu1.a* plants ranged from less than 15% in cool environments to over 75% in warm ones (6). The Bowman backcross-derived line for *uzu1.a*, BW885, produced plants that were 20 to 40% shorter than Bowman, awns were about 1/3 of normal length, rachis internodes were shorter, 3.0 vs. 4.7 mm, and leaf blades were shorter and wider. Kernels of BW885 were shorter, 7.9 vs. 9.5 mm, and lighter, averaged 4.7 vs. 5.7 mg. Spikes of BW885 often had 2 more kernels than those of Bowman. Grain

yields of BW885 ranged from 1/3 to 3/4 those of Bowman (6). Chono et al. (3) reported that the *uzu1.a* variant is caused by a mutation that changed a highly conserved residue of the kinase domain of the *HvBRI1* protein [*BRI1* (brassinosteroid insensitive 1) of *Arabidopsis*] from His-857 to Arg-857. The *uzu1* (*HvDWARF*) gene encodes for the enzyme brassinosteroid C6-oxidase, which is involved in brassinosteroid biosynthesis (7). When grown at low temperatures, the *uzu1.a* mutant was a semidwarf with 80% of wild-type culm length. The overall plant architecture is more erect, with acute leaf blade angles. Short-awned spikes are compact with dense basal spikelets, and frequently with opposite spikelets in the tip caused by irregular elongation of rachis internodes. Leaf blade margins and auricles of *uzu1.a* plants have a slightly undulating appearance (4). When grown at 26°C, *uzu1.a* plants (BW885) showed extreme dwarfing, less than 1/3 the height of Bowman plants. This extreme dwarfing caused by temperature was not observed with other mutants at the *uzu1* locus (4). In progeny from crosses to the BW885 line, tillering was reduced (1). The *uzu1.a* variant was associated with decreased incidence of crown rot, *Fusarium pseudograminearum* (2). Differential phenotypic expressions and responses to heat and drought stress were observed among the *uzu1* mutants (4, 7, 12).

Origin of mutant:

Natural occurrence in some cultivars of Japanese origin (24, 25).

Mutational events:

uzu1.a (OUJ371, PI 182624, GSHO 1300) in East Asian cultivars with a winter growth habit (20, 25, 32); *uzu1.b* (092AR) in Aramir (PI 467781) (9, 10); *uzu1.c* 36 (Katovice 32-1-1) in the doubled-haploid line 36-H930 (4); *ert-ii.79* (NGB 112678, GSHO 483) in Bonus (NGB 14657, PI 189763) (4, 10); *uzu1.256* (formerly *ari.256*) (NGB 116065) in Kristina (NGB 14661, NGB 1500) (4, 14); 522DK (*brd1-a*) and 527DK (*brd1-b*) from Delisa (PI 315937) (7, 8); *brd1-c* and *brd1-d* from Sebastian (7).

Mutant used for description and seed stocks:

uzu1.a (OUJ371, PI 182624, GSHO 1300) in Baitori 11 (OUJ 043); *uzu1.a* in Bowman (PI 483237)*7 (GSHO 1963, BW885, NGB 20787); *uzu1.a* with *wst1.c* (OUL074, GSHO 569) from Akashinriki (PI 467400, OUJ659) in Bowman*8 (GSHO 1967, BW912, NGB 22343); *uzu1.a* with *sld1.a* (OUM148, GSHO 2489) from Akashinriki in Bowman*8 (GSHO 1971, BW860, NGB 22297); *ert-ii.79* in Bowman (PI 483237)*7 (GSHO 1982, BW312, NGB 22108); *uzu1.256* (formerly *ari.256*) from Kristina in Bowman*6 (BW033, NGB20441).

References:

1. Babb, S., and G.J. Muehlbauer. 2003. Genetic and morphological characterization of the barley unculm2 (*cul2*) mutant. *Theor. Appl. Genet.* 106:846-857.
2. Chen, G., W. Yan, Y. Liu, Y. Wei, M. Zhou, Y.L. Zheng, J.M. Manners, and C. Liu. 2014. The non-gibberellic acid-responsive semi-dwarfing gene *uzu* affects *Fusarium* crown rot resistance in barley. *BMC Plant Biol.* 2014 14:22.
3. Chono, M., I. Honda, H. Zeniya, K. Yoneyama, D. Saisho, K. Takeda, S. Takatsuto, T. Hoshino and Y. Watanabe. 2003. A semidwarf phenotype of barley *uzu* results from a nucleotide substitution in the gene encoding a putative brassinosteroid receptor. *Plant Physiol.* 133:1209-1219.
4. Dockter, C., D. Gruszka, I. Braumann, A. Druka, I. Druka, J. Franckowiak, S.P. Gough, A. Janeczko, M. Kurowska, J. Lundqvist, U. Lundqvist, M. Marzec, I. Matyszczyk, A.H. Müller, J. Oklestkova, B. Schulz, S. Zakhrebekova, and M. Hansson. 2014. Induced variations in brassinosteroid genes define barley height and sturdiness,

- and expand the green revolution genetic toolkit. *Plant Physiol.* 166:1912-1927.
5. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
6. Franckowiak, J.D. (Unpublished).
7. Gruszka, D. M. Gorniak, E. Glodowska, E. Wierus, J. Oklestkova, A. Janeczko, M. Maluszynski, and I. Szarejko. 2016. A reverse-genetics mutational analysis of the barley *HvDWARF* gene results in identification of a series of alleles and mutants with short stature of various degree and disturbance in BR biosynthesis allowing a new insight into the process. *Int. J. Mol. Sci.* 17:600.
8. Gruszka, D.; I. Szarejko.; and M. Maluszynski. 2011. Identification of barley *DWARF* gene involved in brassinosteroid synthesis. *Plant Growth Regul.* 65:343-358.
9. Gruszka, D., I. Szarejko, and M. Maluszynski. 2011. New allele of *HvBR11* gene encoding brassinosteroid receptor in barley. *J. Appl. Genet.* 52:257-268.
10. Gruszka, D., J. Zbieszczek, M. Kwasniewski, I. Szarejko, and M. Maluszynski. 2006. A new allele in a *uzu* gene encoding brassinosteroid receptor. *Barley Genet. Newsl.* 36:1-2.
11. Hagberg, A., G. Persson, and A. Wiberg. 1963. Induced mutations in the improvement of self-pollinated crops. p. 105-124. *In* E. Åkerberg and A. Hagberg (eds.) *Recent Plant Breeding Research. Svalöf 1946-1961.* Almqvist & Wiksell, Stockholm.
12. Janeczko, A., D. Gruszka, E. Pociecha, M. Dziurka, M. Filek, B. Jurczyk, H.M. Kalaji, M. Kocurek, and P. Waligorski. 2016: Physiological and biochemical characterisation of watered and drought-stressed barley mutants in the *HvDWARF* gene encoding C6-oxidase involved in brassinosteroid biosynthesis. *Plant Physiol. Biochem.* 99:126-141.
13. Leonard, W.H., H.O. Mann, and L. Powers. 1957. Partitioning method of genetic analysis applied to plant height inheritance in barley. *Colorado Agric. Expt. St. Tech. Bull.* 60:1-24.
14. Lundqvist, U. (Unpublished).
15. Miyake, K., and Y. Imai. 1922. [Genetic studies in barley. 1.] *Bot. Mag., Tokyo* 36:25-38. [In Japanese.]
16. Persson, G. 1969. An attempt to find suitable genetic markers for the dense ear loci in barley I. *Hereditas* 62:25-96.
17. Persson, G. 1969. An attempt to find suitable genetic markers for the dense ear loci in barley II. *Hereditas* 63:1-28.
18. Persson, G., and A. Hagberg. 1969. Induced variation in a quantitative character in barley. Morphology and cytogenetics of *erectoides* mutants. *Hereditas* 61:115-178.
19. Pozzi, C., D. di Pietro, G. Halas, C. Roig, and F. Salamini. 2003. Integration of a barley (*Hordeum vulgare*) molecular linkage map with the position of genetic loci hosting 29 developmental mutants. *Heredity* 90:390-396.
20. Saisho, D., K. Tanno, M. Chono, I. Honda, H. Kitano, and K. Takeda. 2004. Spontaneous brassinolide-insensitive barley mutants “uzu” adapted to East Asia. *Breed. Sci.* 54:409-416.
21. Singh, R.J., A. Shahla, and T. Tsuchiya. 1982. Telotrisomic analysis of three genes with newly obtained telotrisomic, Triplo 3S, in barley. *Barley Genet. Newsl.* 12:42-44.
22. Singh, R.J., and T. Tsuchiya. 1974. Further information on telotrisomic analysis in barley. *Barley Genet. Newsl.* 4:66-69.
23. So, M., S. Ogura, and Y. Imai. 1919. [A linkage group in barley.] *J. Sci. Agric. Soc. Jpn.* 208:1093-1117. [In Japanese.]

24. Takahashi, R. 1942. Studies on the classification and the geographical distribution of the Japanese barley varieties. I. Significance of the bimodal curve of the coleoptile length. Ber. Ohara Inst. landw. Forsch. 9:71-90.
25. Takahashi, R. 1951. Studies on the classification and geographical distribution of the Japanese barley varieties. II. Correlative inheritance of some quantitative characters with the ear types. Ber. Ohara Inst. landw. Forsch. 9:383-398.
26. Takahashi, R., and J. Hayashi. 1959. Linkage study of albino lemma character in barley. Ber. Ohara Inst. landw. Biol., Okayama Univ. 11:132-140.
27. Takahashi, R., and J. Yamamoto. 1951. Studies on the classification and geographical distribution of the Japanese barley varieties. III. On the linkage relation and the origin of the "uzu" or semi-brachytic character in barley. Ber. Ohara Inst. landw. Forsch. 9:399-410.
28. Takezaki, Y. 1927. On the genetical formulae of the length of spikes and awns in barley, with reference to the computation of the valency of the heredity factors. Rep. Agric. Exp. Sta., Tokyo 46:1-42.
29. The International Barley Genome Sequencing Consortium. 2012. A physical, genetic and functional sequence assembly of the barley genome. Nature 491:711-716.
30. Tsuchiya, T. 1972. Genetics of *uz*, *uz2* and *uz3* for semi-brachytic mutations in barley. Barley Genet. Newsl. 2:87-90.
31. Tsuchiya, T. 1976. Allelism testing in barley. II. Allelic relationships of three uzu genes. Crop Sci. 16:496-499.
32. Tsuchiya, T. 1981. Further results on the allelic relationships of three uzu genes in barley. J. Hered. 72:455-458.

Prepared:

T. Tsuchiya and T.E. Haus. 1971. Barley Genet. Newsl. 1:124.

Revised:

- T. Tsuchiya. 1984. Barley Genet. Newsl. 14:92.
J.D. Franckowiak and T. Konishi. 1997. Barley Genet. Newsl. 26:136-137.
J.D. Franckowiak. 2007. Barley Genet. Newsl. 37:220-221.
J.D. Franckowiak. 2011. Barley Genet. Newsl. 41:94-96.
J.D. Franckowiak and U. Lundqvist. 2015. Barley Genet. Newsl. 45:109-112.
J.D. Franckowiak and U. Lundqvist. 2017. Barley Genet. Newsl. 47:70-73.

Stock number: BGS 118
Locus name: Low number of tillers 1
Locus symbol: *Int1*

Previous nomenclature and gene symbolization:

Reduced number of tillers = *rnt* (7).

Intermedium spike-l = *int-l* (6).

Inheritance:

Monofactorial recessive (7).

Located in chromosome 3HL (7); *Int1.a* is about 30.5 cM distal from the *uzu1* (*uzu* 1) locus (7, 8); *Int1.a* mapped about 7.8 cM from SSR marker GBM1043 in 3HL (4); *Int1.a* was not associated with any SNP markers in the Bowman backcross-derived line BW494 (2); *int-l.81* is associated with SNP markers 2_1517 to 2_1405 (positions 158.21 to 187.28 cM) in 3H bins 11 to 12 of the Bowman backcross-derived line BW428 (2), in 3H bin11 (1).

Description:

The tiller number is reduced to 2 to 4 per plant. These tillers are formed soon after seedling emergence; hence, no late-emerging tillers are observed. Culms are thick and stiff, and leaves are dark green (7). Plant mutants at the *Int1* locus fail to produce secondary tillers (1). Occasional spike malformations occur in most environments. The spike may have irregular rachis internode lengths and is relatively short. The lower portion of the spike appears more compact than the upper portion (1, 6). Lateral spikelets in two-rowed cultivars are enlarged and have a pointed apex. Plants homozygous for a recessive allele at the *Int1* locus headed slightly earlier than normal sibs (6). Compared to Bowman, the grain yields of the backcross-derived lines for *Int1.a* (BW494) and *int-l.81* (BW428) were about 10% of those of Bowman. Kernels were longer and wider and weighed up to 20% more (3). Double mutant plants with the *Int1.a* and *int-b.3* (intermedium spike-b) genes produced unicum plants (1). The *Int1.a* gene showed an epistatic interaction with high tillering mutants *gra-a.1* (*granum-a*) *int-m.85* (intermedium spike-m), *mnd1.a* (many noded dwarf 1) and *mnd6.6* (many noded dwarf 6), producing double mutant plants with 2 to 3 tillers (1). Reduced transcript levels for Contig12274, Bell-like homeodomain protein (*JuBel2*) in *Int1.a* plants and co-segregation with *JuBel2* were observed (1).

Origin of mutant:

A spontaneous mutant in the hybrid Chikurin Ibaragi 2/Miho Hadaka (7).

Mutational events:

Int1.a (GSHO 833) in Mitake (OUJ408) (7); *int-l.81* (NGB 115499, GSHO 1771) in Bonus (NGB 14657, PI 189763) (3, 6); *Int1.b* (FN468, GSHO 3678) in Steptoe (CIho 15229) may be an allele based on phenotypic similarity (5).

Mutant used for description and seed stocks:

Int1.a (GSHO 833) in Mitake; *int-l.81* (NGB 115499, GSHO 1771) in Bonus; *Int1.a* in Bowman (PI 483237)*8 (GSHO 1984, BW494, NGB 22157); *int-l.81* in Bowman*6 (GSHO 1961, BW428, NGB 22152).

References:

1. Dabbert, T., R.J. Okagaki, S. Cho, S. Heinen, J. Boddu, and G.J. Muehlbauer. 2010. The genetics of barley low-tillering mutants: *low number of tillers-1* (*Int1*). Theor. Appl.

Genet. 121:705-715.

2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
3. Franckowiak, J. (Unpublished).
4. Liu, Y.X., X-H. Sun, B.-L. Xu, and G.-Y. Chen. 2012. Genetic analysis and molecular mapping of low-tillering mutants (*cul2.b* and *Int1.a*) in barley (*Hordeum vulgare* L.). *J. Plant Sci.* 7:105-112.
5. Kleinhofs, A. (Unpublished).
6. Lundqvist, U., and A. Lundqvist. 1988. Induced intermedium mutants in barley: origin, morphology and inheritance. *Hereditas* 108:13-26.
7. Nonaka, S. 1973. A new type of cultivar, Mitake, with very few in number, but thick and stiff culms. *Barley Genet. Newsl.* 3:45-47.
8. Nonaka, S. 1974. Further information of the *rnt* locus on chromosome 3. *Barley Genet. Newsl.* 4:56-58.

Prepared:

S. Nonaka. 1973. *Barley Genet. Newsl.* 3:120.

Revised:

- T. Tsuchiya. 1980. *Barley Genet. Newsl.* 10:113.
- J.D. Franckowiak and U. Lundqvist. 1997. *Barley Genet. Newsl.* 26:153.
- J.D. Franckowiak. 2013. *Barley Genet. Newsl.* 43:82-83.
- J.D. Franckowiak and U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:74-75.

Stock number: BGS 124
Locus name: Six-rowed spike 4
Locus symbol: *vrs4*

Previous nomenclature and gene symbolization:

Multiflorus = *mul* (8, 19).
Intermediate spike-e = *int-e* (6, 13).
Six-rowed spike 4 = *v4* (3).
Hordeum vulgare RASMOSA2 = *HvRA2* (10).

Inheritance:

Monofactorial recessive (3, 4, 5, 7).
Located in chromosome 3HL (5, 16, 17, 18); *vrs4.k* is about 27.5 cM from the *uzu1* (*uzu* 1) locus (5); *int-e.58* is associated with SNP markers 1_0672 to 2_1083 (positions 38.56 to 156.06 cM) in 3H bins 04 to 10 of the Bowman backcross-derived line BW423 (1); *mul1.a* is associated with SNP markers 1_0762 and 2_0115 (positions 38.56 and 126.83 cM) in 3H bins 04 and 08 of line BW606 (1); *vrs4.k* is associated with SNP markers 1_0863 to 1_0926 (positions 64.85 to 85.26 cM) in 3H bin 05 and with SNP markers 2_1493 to 1_1330 (positions 161.43 to 178.12 cM) in 3H bin 10 of the Bowman backcross-derived line BW903 (1); the lack of SNP marker heterogeneities in the centromeric region of the BW lines and more markers retained in 3HS indicate that the *vrs4* locus is more likely in 3H bin 05 (1, 2).

Description:

Alleles at this locus enhance the development of lateral spikelets and formation of additional spikelets to various degrees. The *int-e* mutants, which were isolated in two-rowed cultivars, have enlarged lateral spikelets that may set seed in the upper two-thirds of the spike. Kernels in lateral spikelets are smaller than those from central spikelets in *int-e* mutants. The rachilla may be deformed by partial formation of an extra spikelet. The awn size on lateral spikelets ranges from a pointed apex to 3/4 normal length (14). The *mul1.a* and *vrs4.k* alleles, which were isolated in six-rowed cultivars, may produce numerous extra spikelets at the base of the lateral spikelets and on the rachilla (4, 8). Plants of the backcross derived line BW903 with the *vrs4.k* allele and BW606 with *mul1.a* are slightly taller than Bowman, plants lodge easily, and rachis internodes are slightly longer. Kernels of BW423 with *int-e.58* had weights that range similar to those of Bowman to 10% less. Kernels of BW606 and BW903, which include small lateral ones, had average weights 20 to 40% less than those of Bowman. The kernel weights for BW606 were slightly less than those of BW093. Kernels of all BW lines with *vrs4* alleles are about 10% shorter than those of Bowman (2). Compared to Bowman, BW903 had more kernels per spike, but tiller numbers and seed weights were reduced (11). Expression analyses through mRNA in situ hybridization and microarray showed that *Vrs4* (*HvRA2*) controls the row-type pathway through *Vrs1* (Six-rowed spike 1, *Hordeum vulgare* Homeobox 1 = *HvHox1*), which is a negative regulator of lateral spikelet fertility (10). Scanning electron microscopy imaging of wild type and *vrs4* mutants at double ridge stage revealed no morphological differences. However, the appearance of two additional mounds on either side of the lateral spikelet meristems was the first visible deviation observed in *vrs4* mutant inflorescences. The additional spikelets emerging at rachis internodes were frequently fertile and developed into kernels (10).

Origin of mutant:

A gamma-ray induced mutant in MFB 104 (PI 232921) (5).

Mutational events:

mul1.a (Alb Acc 321, GSHO 561) in Montcalm (Clho 7149) (8, 19); *vrs4.k* (Gamma III 3B, MFB-2, GSHO 775) in MFB 104 (PI 232921), *vrs4.l* (Xc 41.5, Piro-2) in Pirolina (PI 262210) (5); *vrs4.m* (X17 2497, Piro-7) in Pirolina (4, 5); *int-e.4* (trans) (NGB 115423) in Bonus (NGB 14657, PI 189763), *-e.20* (trans) (NGB 115438), *-e.23* (NGB 115441), *-e.26* (NGB 115444) in Foma (NGB 14659, Clho 11333), *-e.58* (NGB 115476, GSHO 1776), *-e.66* (NGB 115484) in Kristina (NGB 14661, NGB 1500), *-e.65* (NGB 115483) in Bonus (13); *int-e.72* (NGB 115490), *-e.87* (NGB 115505) in Bonus, *-e.89* (NGB 115507), *-e.90* (NGB 115508), *-e.91* (NGB 115509), *-e.92* (NGB 115510), *-e.101* (NGB 115519) in Hege (NGB 13692) (12); *hex-v.46* (NGB 115583), *hex-v.47* (NGB 115584), *hex-v.48* (NGB 115585) in Bonus (9, 12).

Mutant used for description and seed stocks:

vrs4.k (Gamma III 3B, GSHO 775) in MFB 104; *int-e.58* in Kristina (GSHO 1776, NGB 115476); *mul1.a* in Bowman (PI 483237)*6 (GSHO 1985); *mul1.a* in Bowman*7 (BW606, NGB 22172); *vrs4.k* in Bowman*6 (GSHO 1986); *vrs4.k* in Bowman*7 (BW903, NGB 22335); *int-e.58* in Bowman*6 (GSHO 1987); *int-e.58* in Bowman*7 (BW423, NGB 20656).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Fukuyama, T., J. Hayashi, I. Moriya, and R. Takahashi. 1972. A test for allelism of 32 induced six-rowed mutants. *Barley Genet. Newsl.* 2:25-27.
4. Fukuyama, T., J. Hayashi, and R. Takahashi. 1975. Genetic and linkage studies of the five types of induced 'six-row' mutants. *Barley Genet. Newsl.* 5:12-13.
5. Fukuyama, T., R. Takahashi, and J. Hayashi. 1982. Genetic studies on the induced six-rowed mutants in barley. *Ber. Ohara Inst. landw. Biol., Okayama Univ.* 18:99-113.
6. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
7. Gustafsson, Å., and U. Lundqvist. 1980. Hexastichon and intermedium mutants in barley. *Hereditas* 92:229-236.
8. Kasha, K.J., and G.W.R. Walker. 1960. Several recent barley mutants and their linkages. *Can. J. Genet. Cytol.* 2:397-415.
9. Komatsuda, T., M. Pourkheirandish, C. He, P. Azhaguvel, H. Kanamori, D. Perovic, N. Stein, A. Graner, T. Wicker, A. Tagiri, U. Lundqvist, T. Fujimura, M. Matsuoka, T. Matsumoto, and M. Yano. 2007. Six-rowed barley originated from a mutation in a homeodomain-leucine zipper I-class homeobox gene. *Proc. Natl. Acad. Sci. USA* 104 (4):1424-1429.
10. Koppolu, R., N. Anwar, S. Sakuma, A. Tagiri, U. Lundqvist, M. Pourkheirandish, T. Rutten, C. Seiler, A. Himmelbach, R. Ariyadasa, H.M. Youssef, N. Stein, N. Sreenivasulu, T. Komatsuda, and T. Schnurbusch. 2013. Six-rowed spike4 (*Vrs4*) controls spikelet determinacy and row-type in barley. *Proc. Natl. Acad. Sci. USA* 110 (32):13198-13203.
11. Liller, C.B., R. Neuhaus, M. von Korff, M. Koornneef, and W. van Esse. 2015. Mutations in barley row type genes have pleiotropic effects on shoot branching. *PLoS*

One 10: e0140246.

12. Lundqvist, U. (Unpublished).

13. Lundqvist, U. 1991. Coordinator's report: Ear morphology genes. *Barley Genet. Newsl.* 20:85-86.

14. Lundqvist, U., and A. Lundqvist. 1988. Induced intermedium mutants in barley: origin, morphology and inheritance. *Hereditas* 108:13-26.

15. Nötzel, H. 1952. Genetische Untersuchungen an röntgeninduzierten Gerstenmutanten. *Kühn-Archiv* 66:72-132.

16. Nybom, N. 1954. Mutation types in barley. *Acta Agric. Scand.* 4:430-456.

17. Persson, G. 1969. An attempt to find suitable genetic markers for dense ear loci in barley I. *Hereditas* 62:25-96.

18. Singh, R.J., and T. Tsuchiya. 1982. Identification and designation of telocentric chromosomes in barley by means of Giemsa N-banding technique. *Theor. Appl. Genet.* 64:13-24.

19. Walker, G.W., K. Kasha, and R.A. Miller. 1958. Recombination studies in barley. *Proc. Genet. Soc. Can.* 3:41-43.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:159-160.

Revised:

U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:101-102.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:76-78.

BGS 131, Granum-a, *gra-a*

Stock number: BGS 131
Locus name: Granum-a
Locus symbol: *gra-a*

Previous nomenclature and gene symbolization:

Many-noded and nodal-branched with no gene symbol (8).

Granum-a = *gran-a* (3).

Inheritance:

Monofactorial recessive (3, 7, 8).

Located in chromosome 7H (2); *gra-a.1* is associated with SNP markers 1_0861 to 1_0078 (positions 183.90 to 187.83 cM) in 7H bin 11 of the Bowman backcross-derived line BW398 (2); *gra-a.2* is associated with SNP markers 1_0078 to 1_0547 (position 187.83 to 231.00 cM) in 7H bins 11 to 13 of the Bowman backcross-derived line BW399 (2), likely in 7H bin 11. Previously the *gra-a.1* mutant was reported to be in chromosome 3HL (3, 7).

Description:

Plants have numerous, thin tillers with narrow leaves and short internodes (1/2 normal plant height). Spikes are less than half normal length and seeds are thin and small (3, 7). Spikes emerge from the sheath of the flag leaf prematurely, 4 to 7 days before anthesis (5). Compared to Bowman plants of Bowman, backcross-derived lines for *gra-a.1* (BW398) and *gra-a.2* (BW399) were about half of normal height and peduncles were about 1/3 normal length (4). Awns and rachis internodes of the BW398 spikes were slightly shorter than those of Bowman (1, 4). The tip of the spike has a fasciated appearance because spikelets are very close together (1, 4). The number of kernels per spike for BW398 and BW399 were often 4 to 7 less than the number for Bowman. Kernel weights were about 25% less (45 vs. 59 mg) than those of Bowman, but grain yields were almost normal (4). Combining the *gra-a.1* and *cul2.b* (uniculm 2) mutants produced short plants that occasionally produced two tillers (1).

Origin of mutant:

An X-ray induced mutant in Donaria (PI 161974) (3, 7).

Mutational events:

gra-a.1 (GSHO 1757) in Donaria (PI 161974) (3; 7); *gra-a.2* (GSHO 1758) from a Tokak (PI 178421) mutant in OR-SS-2 (5, 6, 9); *gra-a.3* (DWS1176) in HE2816 (likely from HE-MN-C 292) from Milan Váša (5, 6); many noded and nodal branched mutants in C. 167 (8).

Mutant used for description and seed stocks:

gra-a.1 (GSHO 1757) in Donaria; *gra-a.1* in Bowman (PI 483237)*7 (GSHO 1980, BW398, NGB 20635); *gra-a.2* in Bowman*7 (GSHO 1981, BW 399, NGB 20636).

References:

1. Babb, S., and G.J. Muehlbauer. 2003. Genetic and morphological characterization of the barley unculm 2 (*cul2*) mutant. Theor. Appl. Genet. 106:846–857.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. Plant Physiol. 155:617-627.
3. Fischbeck, G., and H. Häuser. 1976. Research notes. Barley Genet. Newsl. 6:28-29.
4. Franckowiak, J.D. (Unpublished).

5. Franckowiak, J.D. 1992. Allelism tests among selected semidwarf barleys. *Barley Genet. Newsl.* 21:17-23.
6. Franckowiak, J.D., and A. Pecio. 1992. Coordinator's report: Semidwarf genes: A listing of genetic stocks. *Barley Genet. Newsl.* 21:116-127
7. Häuser, J., and G. Fischbeck. 1976. Untersuchungen zur Lokalisierung einiger Mutationen von Gerste (*Hordeum sativum*). *Z. Pflanzenzücht.* 77:269-280.
8. Minogha, J.L., and D.S. Brar. 1969. Many-noded and nodal-branched mutants in barley. *Indian J. Genet. Plant Breed.* 29:446-447.
9. Sears, R.G., W.E. Kronstad, and R.J. Metzger. 1981. Inheritance of dwarf and semidwarf plant height in barley. *Crop Sci.* 21:828-833.

Prepared:

J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:167.

Revised:

J.D. Franckowiak. 2012. *Barley Genet. Newsl.* 42:216-217.

J.D. Franckowiak. 2014. *Barley Genet. Newsl.* 44:88-89.

J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:79-80.

BGS 155, Glossy leaf 1, *glf1*

Stock number: BGS 155
Locus name: Glossy leaf 1
Locus symbol: *glf1*

Previous nomenclature and gene symbolization:

Waxless bloom on leaves = *w1* (16).

Glossy = *gl* (14).

Glossy leaves = *gl* (21).

Glossy leaf = *gl* (20).

Glossy seedling 2 = *gl2* (6, 14).

Eceriferum-zh = *cer-zh* (9).

Inheritance:

Monofactorial recessive (14).

Located in chromosome 4HL (5, 14, 17, 19); *glf1.a* is about 1.5 cM distal from the *lbi2* (long basal rachis internode 2) locus (2, 6, 7); *glf1.a* is about 1.2 cM proximal from the *Mlg* (*Reg2*, reaction to *Erysiphe graminis* 2) locus (2, 6); *glf1.a* is associated with SNP markers 2_0820 to 1_0606 (positions 89.78 to 95.92 cM) in 4HL of the Bowman backcross-derived line BW385 (1); *glf1.a* co-segregated with molecular markers AK361226 and AK362558 (8), in 4H bin 06.

Description:

Surface wax coating on the leaf blade appears absent from the seedling stage to near maturity, and leaves have a shiny appearance (wax code ++ ++ -) (9). Plants are semidwarf, relatively weak, and late in heading. The stock in the Bonus is highly sterile (9), but the Bowman backcross-derived line has nearly complete fertility (3). The lack of surface waxes reduces the ability of growing germ tube of certain fungi to find the stomata openings (15). Plants of Bowman backcross-derived line BW385 were 5 to 15% shorter than Bowman and headed about 2 days later. Kernel weights were about 20% lower and grain yields were approximately 3/4 of normal (3). Seedling leaf waxes in the *cer-zh.54* mutant showed a 76% decrease in total wax load due to marked decreases of all wax classes except the esters (8). The gene mutated in *cer-zh.54* was identified an elongase component, β -ketoacyl-CoA synthase (CER-ZH/*HvKCS1*) (8).

Origin of mutant:

A radiation induced mutant in Himalaya (Clho 1312) (14, 18), an X-ray induced mutant in Bonus (NGB 14657, PI 189763) (9).

Mutational events:

glf1.a (GSHO 98), *glf1.b* (*gl2*, GSHO 22) in Himalaya (Clho 1312) (18); *glf1.f* in 34-119-1, *glf1.g* in Il-34-199-7-2 (GSHO 89) (4); *cer-zh.54* (NGB 110938, GSHO 455) in Bonus (NGB 14657, PI 189763) (9, 10); *cer-zh.266* (NGB 111153), *-zh.308* (NGB 111195), *-zh.357* (NGB 111244, NGB 117254), *-zh.366* (NGB 111253), *-zh.432* (NGB 111320), *-zh.433* (NGB 111321) in Foma (NGB 14659, Clho 11333) (10, 13); *cer-zh.325* (NGB 111212) in Foma (10); *cer-zh.373* (NGB 111260) in Foma (11); *cer-zh.865* (NGB 111753) in Bonus (12); *glf1.h* (MXM137) in Morex (Clho 15773) based on phenotype and SNP markers retained in the Bowman backcross-derived line BW384 (1, 3).

Mutant used for description and seed stocks:

glf1.a (GSHO 98) in Himalaya; *cer-zh.54* (NGB 110938, GSHO 455) in Bonus was used for allelism tests; *glf1.a* via R.I. Wolfe's chromosome 4 multiple recessive marker stock

in Bowman (PI 483237)*8 (BW385, NGB 20623, GSHO 2015); *cer-zh.54* in Bowman*3 (BW164, NGB 21996); *glf1.h* in Bowman*5 (BW384, NGB 22146).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Forster, B.P. 1992. Coordinator's report: Chromosome 4. *Barley Genet. Newsl.* 21:86-88.
3. Franckowiak, J.D. (Unpublished).
4. Haus, T.E., and T. Tsuchiya. 1972. Allelic relationships among glossy seedling mutants. *Barley Genet. Newsl.* 2:79-80.
5. Immer, F.R., and M.T. Henderson. 1943. Linkage studies in barley. *Genetics* 28:419-440.
6. Jensen, J. 1987. Linkage map of barley chromosome 4. p. 189-199. *In* S. Yasuda and T. Konishi (eds.) *Barley Genetics V. Proc. Fifth Int. Barley Genet. Symp.*, Okayama, 1986. Sanyo Press Co., Okayama.
7. Kasha, K.J., and G.W.R. Walker. 1960. Several recent barley mutants and their linkages. *Can. J. Genet. Cytol.* 2:397-415.
8. Li, C., T.M. Haslam, A. Krüger, L.M. Schneider, K. Mishina, L. Samuels, H. Yang, L. Kunst, U. Schaffrath, C. Nawrath, G. Chen, T. Komatsuda, and P. von Wettstein-Knowles. 2018. The β -Ketoacyl-CoA Synthase *HvKCS1*, encoded by *Cer-zh*, plays a key role in synthesis of barley leaf wax and germination of barley powdery mildew. *Plant Cell Physiol.* 59:806-822.
9. Lundqvist, U., and D. von Wettstein. 1962. Induction of eceriferum mutants in barley by ionizing radiations and chemical mutagens. *Hereditas* 48:342-362.
10. Lundqvist, U., and D. von Wettstein. 1971. Stock list for the eceriferum mutants. *Barley Genet. Newsl.* 1:97-102.
11. Lundqvist, U., and D. von Wettstein. 1973. Stock list for the eceriferum mutants II. *Barley Genet. Newsl.* 3:110-112.
12. Lundqvist, U., and D. von Wettstein. 1977. Stock list for the eceriferum mutants IV. *Barley Genet. Newsl.* 5:92-96.
13. Lundqvist, U., P. von Wettstein-Knowles, and D. von Wettstein. 1968. Induction of eceriferum mutants in barley by ionizing radiations and chemical mutagens. II. *Hereditas* 59:473-504.
14. Robertson, D.W., and O.H. Coleman. 1942. Location of glossy and yellow leaf seedlings in two linkage groups. *J. Am. Soc. Agron.* 34:1028-1034.
15. Rubiales, D., M.C. Ramirez, T.L.W. Carver, and R.E. Niks. 2001. Abnormal germling development by brown rust and powdery mildew on *cer* barley mutants. *Hereditas* 135:271-276.
16. Smith, L. 1951. Cytology and genetics of barley. *Bot. Rev.* 17:1-55, 133-202, 285-355.
17. Tsuchiya, T. 1981. Revised linkage maps of barley. 1981. *Barley Genet. Newsl.* 11:96-98.
18. Tsuchiya, T., and T.E. Haus. 1973. Allelism testing in barley. I. Analysis of ten mapped genes. *J. Hered.* 64:282-284.
19. Tsuchiya, T., and R.J. Singh. 1982. Chromosome mapping in barley by means of telotrisomic analysis. *Theor. Appl. Genet.* 61:201-208.
20. Walker, G.W.R., J. Dietrich, R. Miller, and K. Kasha. 1963. Recent barley mutants

and their linkages II. Genetic data for further mutants. Can. J. Genet. Cytol. 5:200-219.
21. Woodward, R.W. 1957. Linkages in barley. Agron. J. 49:28-32.

Prepared:

T.E. Haus and T. Tsuchiya. 1971. Barley Genet. Newsl. 1:141 as BGS 155, Glossy seedling, *gl*; and Barley Genet. Newsl. 1:145 as BGS 159, Glossy seedling 2, *gl2*.

U. Lundqvist. 1975. Barley Genet. Newsl. 5:144 as BGS 426, Eceriferum-zh, *cer-zh*.

Revised:

T. Tsuchiya. 1980. Barley Genet. Newsl. 10:114 as BGS 155, Glossy seedling, *gl*; and Barley Genet. Newsl. 10:116 as BGS 159, Glossy seedling 2, *gl2*.

U. Lundqvist and J.D. Franckowiak. 1997. Barley Genet. Newsl. 26:181-182.

U. Lundqvist and J.D. Franckowiak. 2007. Barley Genet. Newsl. 37:233-234.

U. Lundqvist and J.D. Franckowiak. 2010. Barley Genet. Newsl. 40:61-63.

U. Lundqvist and J.D. Franckowiak. 2017. Barley Genet. Newsl. 47:81-83.

Stock number: BGS 157
Locus name: Brachytic 2
Locus symbol: *brh2*

Previous nomenclature and gene symbolization:

Brachytic 2 = *br2* (11).
Breviaristatum-I = *ari-I* (6, 7).

Inheritance:

Monofactorial recessive (10).
Located in chromosome 4HL (10); *brh2.b* is about 1.5 cM proximal from the *glf3* (glossy leaf 3) locus (4, 10); *brh2.b* is over 22.8 cM proximal from the *Kap1* (Hooded lemma 1) locus (10); *brh2.b* is near AFLP marker E4140-7 in subgroup 38-40 of the Proctor/Nudinka map (9), *brh2.b* is about 15.9 cM distal from SSR marker Bmag0353 near the boundary between 4H bins 06 and 07 (2); *brh2.b* is associated with SNP markers 2_0012 to 1_1319 (positions 57.19 to 110.93 cM) in 4H bins 05 to 07 of the Bowman backcrossed-derived line BW090 (3); *ari-I.3* is associated with SNP markers 2_1374 to 2_1332 (positions 41.13 to 112.64 cM) in 4H bins 04 to 08 of the Bowman backcrossed-derived line BW050 (3), likely in 4H bin 06.

Description:

Plant height and vigor are reduced to about 2/3 normal; the awn is less than 1/4 normal length; the spike is semi-compact; and the leaf, kernel, glume and glume awn, rachilla, and coleoptile are shorter than in the original cultivar. Auricles are well developed and larger than those of the original cultivar (11). In the Bowman backcross-derived lines, the peduncle is about 1/2 normal length, kernel weights are slightly over 2/3 normal, yield is about 1/2 normal; however, rachis internode lengths are normal (2). The *ari-I.3* allele at the *brh2* locus is sensitive to gibberellic acid treatment (1). The Bowman backcross-derived for *brh2.b*, BW090, and *ari-I.3*, BW050, were morphologically similar to each other, shorter than Bowman, 65 vs. 90 cm. They also had short awns, 2.5 vs. 12 cm beyond the tip of the spike and small kernels. Compared to Bowman, kernels were shorter, 7.5 vs. 9.8 mm, and thinner, 3.61 vs. 3.83 mm, and weighed less, 40 vs. 56 mg. Grain yields for BW050 and BW090 were 1/2 to 2/3 of those produced by Bowman (5).

Origin of mutant:

An X-ray induced mutant in Svanhals (NGB 1482, PI 5474) (11).

Mutational events:

brh2.b (Kmut 28, OUM283, GSHO 573) in Svanhals (NGB 1482, PI 5474) (10); *ari-I.3* (NGB 115848) in Bonus (NGB14657, PI 189763) (7); *ari-I.132* (NGB 115942) in Foma (NGB 14659, CIho 11333) (8); *ari-I.135* (NGB 115945), *-I.145* (NGB 115956), *-I.214* (NGB 116023), *-I.237* (NGB 116047) in Foma, *-I.257* (NGB 116066) in Kristina (NGB 14661, NGB 1500) (7).

Mutant used for description and seed stocks:

brh2.b (GSHO 573) in Svanhals; *ari-I.3* (GSHO 1660) in Bonus; *brh2.b* in Bowman (PI 483237)*7 (GSHO 2016, BW090, NGB 20496); *ari-I.3* in Bowman*7 (GSHO 2017, BW050, NGB 20458).

References:

1. Börner, A. 1996. GA response in semidwarf barley. Barley Genet. Newsl. 25:24-26.
2. Dahleen, L.S., L.J. Vander Wal, and J.D. Franckowiak. 2005. Characterization and

- molecular mapping of genes determining semidwarfism in barley. J. Hered. 96:654-662.
3. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. Plant Physiol. 155:617-627.
4. Forster, B.P. 1993. Coordinator's report: Chromosome 4. Barley Genet. Newsl. 22:75-77.
5. Franckowiak, J.D. (Unpublished).
6. Franckowiak, J.D. 1995. The brachytic class of semidwarf mutants in barley. Barley Genet. Newsl. 24:56-59.
7. Kucera, J., U. Lundqvist, and Å. Gustafsson. 1975. Inheritance of brevistaratum mutants in barley. Hereditas 80:263-278.
8. Lundqvist, U. (Unpublished).
9. Pozzi, C., D. di Pietro, G. Halas, C. Roig, and F. Salamini. 2003. Integration of a barley (*Hordeum vulgare*) molecular linkage map with the position of genetic loci hosting 29 developmental mutants. Heredity 90:390-396.
10. Takahashi, R., J. Hayashi, and I. Moriya. 1971. Linkage studies in barley. Barley Genet. Newsl. 1:51-58.
11. Tsuchiya, T. 1962. Radiation breeding in two-rowed barley. Seiken Zihō 14:21-34.

Prepared:

T.E. Haus and T. Tsuchiya. 1971. Barley Genet. Newsl. 1:143.

Revised:

T. Tsuchiya. 1980. Barley Genet. Newsl. 10:115.

J.D. Franckowiak and U. Lundqvist. 1997. Barley Genet. Newsl. 26:184.

J.D. Franckowiak and L.S. Dahleen. 2007. Barley Genet. Newsl. 37:235-236.

J.D. Franckowiak and U. Lundqvist. 2014. Barley Genet. Newsl. 44:95-96.

J.D. Franckowiak and U. Lundqvist. 2017. Barley Genet. Newsl. 47:84-85.

BGS 165, Glossy leaf 3, *glf3*

Stock number: BGS 165
Locus name: Glossy leaf 3
Locus symbol: *glf3*

Previous nomenclature and gene symbolization:

Glossy leaf = *gl* (5).
Glossy leaf 3 = *gl3* (17).
Glossy leaves 4 = *gl4* (5, 20, 21, 22).
Eceriferum-j = *cer-j* (5, 8).

Inheritance:

Monofactorial recessive (17, 19).
Located in chromosome 4HL (2, 7, 17); *glf3.c* is about 1.5 cM distal from the *brh2* (brachytic 2) locus (6); *glf3.c* is about 13.7 cM from the *sid1* (single elongated internode 1) locus (6); *glf3.d* is associated with SNP markers 1_1180 to 1_1513 (positions 58.13 to 101.44 cM) in 4H bins 05 to 07 of the Bowman backcross-derived line BW386 (1); *cer-j.59* is associated with SNP markers 2_1122 to 2_1332 (positions 47.80 to 112.64 cM) in 4H bins 05 to 07 of the Bowman backcross-derived line BW114 (1); likely in 4H bin 06.

Description:

Surface wax coating on the leaf blades appears absent from the seedling stage to near maturity, and leaf blades have a shiny appearance (wax code ++ ++ -) (8). Concerning the chemical epicuticular wax composition, the *cer-j* mutant in Bonus produces 34% less wax compared to the wild type, the primary alcohols are the largest wax class. The wax occurs as many very thin plates and as few large irregularly shaped bodies (4, 16, 23). Compared to Bowman, plants of the Bowman backcross-derived line for *glf3.d*, BW386, had kernels that were slightly lighter in some environments. Plants of the Bowman backcross-derived line for *cer-j.59*, BW114, were slightly shorter, headed about 2 days later, and had slightly lighter kernel weights (3).

Origin of mutant:

A spontaneous mutant in Goseshikoku (OUJ128) (17).

Mutational events:

glf3.c (Goseshikoku-hen, OUL032, GSHO 577) in Goseshikoku (OUJ128) (18); *glf3.d* (*gl4*, GSHO 1376) in Gateway (Clho 10072) (22); *cer-j.59* (NGB 110943, GSHO 431), -*j.62* (NGB 110946) in Bonus (NGB 14657, PI 189763) (8, 9); *cer-j.71* (NGB 110955), -*j.142* (NGB 111028), -*j.148* (NGB 111034) in Bonus, -*j.274* (NGB 111161), -*j.288* (NGB 111175), -*j.301* (NGB 111188), -*j.311* (NGB 111198), -*j.340* (NGB 111227), -*j.346* (NGB 111233), -*j.349* (NGB 111236), -*j.358* (NGB 111245), -*j.370* (NGB 111257), -*j.390* (NGB 111277), -*j.429* (NGB 111317), -*j.447* (NGB 111335), -*j.470* (NGB 111358) in Foma (NGB 14659, Clho 11333) (9, 16); *cer-j.183* (NGB 111069), -*j.184* (NGB 111070), -*j.696* (NGB 111584) in Bonus, -*j.275* (NGB 111162), -*j.399* (NGB 111286), -*j.517* (NGB 111405), -*j.521* (NGB 111409), -*j.558* (NGB 111446) in Foma, -*j.1024* (NGB 111912), -*j.1081* (NGB 111969) in Carlsberg II (NGB 5085, Clho 10114) (11); *cer-j.458* (NGB 111346) in Foma, -*j.780* (NGB 111668), -*j.797* (NGB 111685), -*j.832* (NGB 111720) in Bonus, -*j.1090* (NGB 111978), -*j.1102* (NGB 111990) in Kristina (NGB 14661, NGB 1500) (12); *cer-j.909* (NGB 111797) in Bonus, -*j.1152* (NGB 112040) in Kristina (11); *cer-j.1204* (NGB 112092), -*j.1231* (NGB 112119), -*j.1233* (NGB 112121), -*j.1239* (NGB 112127), -*j.1251* (NGB 112139) in Kristina (12); *cer-j.997* (NGB 111885), -*j.1331* (NGB

112219), -j.1361 (NGB 112249), -j.1388 (NGB 112276), -j.1405 (NGB 112293), -j.1433 (NGB 112321), -j.1469 (NGB 112357) in Bonus (12); *cer-j.1712* (NGB 112525), -j.1744 (NGB 112558), -j.1761 (NGB 112576), -j.1803 (NGB 117351), -j.1808 (NGB 117356) in Bonus (12); *cer-j.1827* (NGB 117375) in Bonus, -j.1831 (NGB 117379), -j.1835 (NGB 117383), -j.1839 (NGB 117387), -j.1841 (NGB 117389), -j.1843 (NGB 117391), -j.1844 (NGB 117392), -j.1847 (NGB 117395), -j.1850 (NGB 117398), -j.1852 (NGB 117400), -j.1857 (NGB 117405), -j.1858 (NGB 117406), -j.1859 (NGB 117407), -j.1860 (NGB 117408), -j.1861 (NGB 117409), -j.1862 (NGB 117410) in Sv 79353 (15).

Mutant used for description and seed stocks:

glf3.c (GSHO 577) in Goseshikoku; *cer-j.59* (NGB 110943, GSHO 431) in Bonus is used for allelic localization tests and for wax chemistry and wax structure studies; *glf3.d* from Gateway in Bowman (PI 483237)*5 (GSHO 2019); *glf3.d* in Bowman*8 (BW386, NGB 20624); *cer-j.59* from Bonus in Bowman*3 (GSHO 2021); *cer-j.59* in Bowman*7 (BW114, NGB 20520).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Fester, T., and B. Sogaard. 1969. The localization of *eceriferum* loci in barley. *Hereditas* 61:327-337.
3. Franckowiak, J.D. (Unpublished).
4. Giese, B.N. 1976. Roles of the *cer-j* and *cer-p* loci in determining the epicuticular wax composition on barley seedling leaves. *Hereditas* 82:137-148.
5. Haus, T.E. 1974. Allelism tests of chromosome 4 mutants. *Barley Genet. Newsl.* 4:31-33.
6. Hayashi, J., T. Konishi, I. Moriya, and R. Takahashi. 1984. Inheritance and linkage studies in barley VI. Ten mutant genes located on chromosomes 1 to 7, except 3. *Ber. Ohara Inst. landw. Biol., Okayama Univ.* 18:227-250.
7. Jensen, J. 1987. Linkage map of barley chromosome 4. p. 189-199. *In* S. Yasuda and T. Konishi (eds.) *Barley Genetics V. Proc. Fifth Int. Barley Genet. Symp., Okayama, 1986.* Sanyo Press Co., Okayama.
8. Lundqvist, U., and D. von Wettstein. 1962. Induction of *eceriferum* mutants in barley by ionizing radiations and chemical mutagens. *Hereditas* 48:342-362.
9. Lundqvist, U., and D. von Wettstein. 1971. Stock list for the *eceriferum* mutants. *Barley Genet. Newsl.* 1:97-102.
10. Lundqvist, U., and D. von Wettstein. 1973. Stock list for the *eceriferum* mutants II. *Barley Genet. Newsl.* 3:110-112.
11. Lundqvist, U., and D. von Wettstein. 1975. Stock list for the *eceriferum* mutants III. *Barley Genet. Newsl.* 5:88-91.
12. Lundqvist, U., and D. von Wettstein. 1977. Stock list for the *eceriferum* mutants IV. *Barley Genet. Newsl.* 7:92-96.
13. Lundqvist, U., and D. von Wettstein. 1982. Stock list for the *eceriferum* mutants VI. *Barley Genet. Newsl.* 12:169-172.
14. Lundqvist, U., and D. von Wettstein. 1985. Stock list for the *eceriferum* mutants VII. *Barley Genet. Newsl.* 15:89-93.
15. Lundqvist, U., and D. von Wettstein. 1988. Stock list for the *eceriferum* mutants VIII. *Barley Genet. Newsl.* 18:88-91.
16. Lundqvist, U., P. von Wettstein-Knowles, and D. von Wettstein. 1968. Induction of

eceriferum mutants in barley by ionizing radiations and chemical mutagens. II. Hereditas 59:473-504.

17. Takahashi, R., J. Hayashi, and I. Moriya. 1962. Linkage studies. Barley Newsl. 5:41-42.

18. Takahashi, R., J. Hayashi, and I. Moriya. 1971. Linkage studies in barley. Barley Genet. Newsl. 1:51-58.

19. Takahashi, R., J. Hayashi, T. Konishi, and I. Moriya. 1972. Inheritance and linkage studies in barley V. Locating of seven new mutant genes. Ber. Ohara Inst. landw. Biol., Okayama Univ. 15:147-168.

20. Tsuchiya, T. 1973. Allelic relationship between two glossy seedling genes, *gl3* and *gl4* in barley. Barley Genet. Newsl. 3:66-67.

21. Tsuchiya, T., and T.E. Haus. 1973. Allelism testing in barley. I. Analysis of ten mapped genes. J. Hered. 64:282-284.

22. Walker, G.W.R., J. Dietrich, R. Miller, and K. Kasha. 1963. Recent barley mutants and their linkages II. Genetic data for further mutants. Can. J. Genet. Cytol. 5:200-219.

23. Wettstein-Knowles, P. von, 1971. The molecular phenotypes of the *eceriferum* mutants. p. 146-193. In R.A. Nilan (ed). Barley Genetics II. Proc. Second Int. Barley Genet. Symp., Pullman, WA, 1969. Washington State Univ. Press, Pullman.

Prepared:

R. Takahashi. 1972. Barley Genet. Newsl. 2:190.

T. Tsuchiya and T.E. Haus. 1971. Barley Genet. Newsl. 1:159 as BGS 256, Glossy leaf 4, *gl4*.

Revised:

T. Tsuchiya. 1980. Barley Genet. Newsl. 10:117 as BGS 165, Glossy seedling 3, *gl3*; and Barley Genet. Newsl. 10:123 as BGS 256, Glossy leaf 4, *gl4*.

U. Lundqvist and J.D. Franckowiak. 1997. Barley Genet. Newsl. 26:190-191.

U. Lundqvist and J.D. Franckowiak. 2013. Barley Genet. Newsl. 43:92-94.

U. Lundqvist and J.D. Franckowiak. 2017. Barley Genet. Newsl. 47:86-88.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 172, Short awn 5, *lks5*

Stock number: BGS 172
Locus name: Short awn 5
Locus symbol: *lks5*

Previous nomenclature and gene symbolization:

Short awn 5 = *lk5* (7).
Breviaristatum-2 = *ari-2* (5, 6).
Breviaristatum-c = *ari-c.2* (6, 13, 14).

Inheritance:

Monofactorial recessive (6, 7).
Located in chromosome 4HL (15); *lks5.f* is about 6.9 cM distal from the *Kap1* (Hooded lemma 1) locus (7, 9); *lks5.f* is near AFLP marker E4143-5 in subgroup 38 of the Proctor/Nudinka map (10); *lks5.f* is associated with SNP markers 1_0262 to 2_0072 (positions 76.26 to 95.92 cM) in 4H bin 06 of the Bowman backcrossed-derived line BW493 (2); *ari-c.2* is associated with SNP markers 2_0289 to 2_1322 (positions 69.62 to 112.64 cM) in 4H bins 05 to 08 of the Bowman backcrossed-derived line BW040 (2), in 4H bin 06; *lks5.p* was isolated in genetic stocks BW474 and BW475 from the *lel1* (leafy lemma 1) stock, G7118, *lks5.p* is associated with SNP markers 2_1122 to 1_1019 (positions 47.80 to 183.54) in 4H bins 05 to 13 of the Bowman backcrossed-derived line BW474 (2); *lks5.p* is associated with SNP markers 2_0422 to 2_0072 (positions 38.41 to 95.92 cM) in 4H bins 04 to 06 of the Bowman backcrossed-derived line BW475 (2); in 4H bin 06.

Description:

Awns on both central and lateral spikelets are reduced to 1/4 or less of normal length in six-rowed cultivars (7). Only the central spikelets exhibit reduced awn length in two-rowed cultivars (3, 6). The rachilla is often modified, and in extreme cases the rachilla may develop as a malformed additional floret (7). Awns are thin and brittle (6). Plants of the Bowman backcross-derived line for *lks5.f*, BW493, had awns that extended 3 to 4 cm beyond the tip of the spike, while those of Bowman were 12 to 13 cm beyond the tip. Kernels of BW493 were slightly shorter, narrower (3.4 vs. 3.9 mm), and lighter (4.6 vs. 5.5 mg). BW493 plants were slightly taller than Bowman and grain yields were slightly reduced (4). Presence of the mutant allele *lks5.p* is needed for expression of the leafy lemma 1 (*lel1.a*) gene (4, 10).

Origin of mutant:

A spontaneous mutant in an unknown cultivar (7).

Mutational events:

lks5.f (GSHO 1297) in Clho 5641 (7, 13); *lks5.g* (Kmut 218) in Asahi 5 (OUJ509) (13); *ari-c.2* (NGB 115847, GSHO 1651), -c.59 (NGB 115913) in Bonus (NGB 14657, PI 189763), -c.103 (NGB 115915) in Foma (NGB 14659, Clho 11333) (3); *ari-c.106* (NGB 115918) in Foma (5); *ari-c.109* (NGB 115921), -c.110 (NGB 115922), -c.111 (NGB 115923), -c.112 (NGB 115924), -c.120 (NGB 115932), -c.139 (NGB 115949), -c.157 (NGB 115967), -c.159 (NGB 115969), -c.179 (NGB 115989), -c.180 (NGB 115990), -c.199 (NGB 116008), -c.201 (NGB 116010), -c.203 (NGB 116012), -c.204 (NGB 116013), -c.206 (NGB 116014), -c.210a (NGB 116018), -c.210b (NGB 116019) -c.229 (NGB 116039) in Foma, -c.259 (NGB 116068), -c.262 (NGB 116071), -c.272 (NGB 116084), -c.276 (NGB 116089), -c.289 (NGB 116107), -c.291 (NGB 116111) in Kristina

(NGB 14661, NGB 1500) (6); *ari-c.307* (NGB 116135) in Kristina (8); two possible additional alleles have been reported, *lks5.h* (*lk,,f*) in two-rowed Glacier (3); and a mutant in Morex (CIho 15773) (12); *lks5.p* in G7118 (leafy lemma, GSHO 1780) mutant stock (1, 3, 4, 11).

Mutant used for description and seed stocks:

lks5.f (GSHO 1297) in CIho 5641; *ari-c.2* (NGB 115847, GSHO 1651) in Bonus; *lks5.f* in Bowman (PI 483237)*6 (GSHO 2014); *lks5.f* in Bowman*7 (BW493, NGB 20721); *ari-c.2* in Bowman*7 (GSHO 2013, BW040, NGB 20448); *lks5.p* from G7118 in Bowman*4 with *lel1.a* (BW474, NGB 20704); *lks5.p* from G7118 in Bowman*5 (BW475, NGB 20705).

References:

1. Bossinger, G., U. Lundqvist, W. Rohde, and F. Salamini. 1992. Genetics of plant development in barley. p. 989-1017. *In* L. Munck, K. Kirkegaard, and B. Jensen (eds.). Barley Genetics VI. Proc. Sixth Int. Barley Genet. Symp., Helsingborg, 1991. Munksgaard Int. Publ., Copenhagen.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
3. Eckhoff, J.L.A., and R.T. Ramage. 1984. Assignment of a short awn mutant to chromosome 4. *Barley Genet. Newsl.* 14:20-21.
4. Franckowiak, J.D. (Unpublished).
5. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
6. Kucera, J., U. Lundqvist, and Å. Gustafsson. 1975. Inheritance of brevistaratum mutants in barley. *Hereditas* 80:263-278.
7. Litzenberger, S.C., and J.M. Green. 1951. Inheritance of awns in barley. *Agron. J.* 43:117-123.
8. Lundqvist, U. (Unpublished).
9. Persson, G. 1969. An attempt to find suitable genetic markers for the dense ear loci in barley I. *Hereditas* 62:25-96.
10. Pozzi, C., D. di Pietro, G. Halas, C. Roig, and F. Salamini. 2003. Integration of a barley (*Hordeum vulgare*) molecular linkage map with the position of genetic loci hosting 29 developmental mutants. *Heredity* 90:390-396.
11. Pozzi, C., P. Faccioli, V. Terzi, A.M. Stanca, S. Cerioli, P. Castiglioni, R. Fink, R. Capone, K.J. Müller, G. Bossinger, W. Rohde, and F. Salamini. 2000. Genetics of mutations affecting the development of a barley floral bract. *Genetics* 154:1335-1346.
12. Ramage, T., and J.L.A. Eckhoff. 1985. Assignment of mutants in Morex to chromosomes. *Barley Genet. Newsl.* 15:22-25.
13. Tsuchiya, T. 1974. Allelic relationships of genes for short-awned mutants in barley. *Barley Genet. Newsl.* 4:80-81.
14. Tsuchiya, T. 1975. Characteristics and inheritance of radiation-induced mutations in barley II. Two short-awned mutations. *Genetica* 45:519-529.
15. Tsuchiya, T., and L.B. Hall. 1978. Telotrisomic analysis of four mutant genes in barley. *Barley Genet. Newsl.* 8:104-107.

Prepared:

T.E. Haus. 1978. *Barley Genet. Newsl.* 8:156.

Revised:

T. Tsuchiya. 1980. *Barley Genet. Newsl.* 10:119.

J.D. Franckowiak and U. Lundqvist. 1997. *Barley Genet. Newsl.* 26:197.

Barley Genetics Newsletter (2017): 46:50-187.
J.D. Franckowiak and U. Lundqvist. 2011. Barley Genet. Newsl. 41:110-111.
J.D. Franckowiak and U. Lundqvist. 2017. Barley Genet. Newsl. 47:89-91.

Stock number: BGS 178
Locus name: Intermedium spike-c
Locus symbol: *int-c*

Previous nomenclature and gene symbolization:

Intensifier for *Z* = *W* (25).
Infertile intermedium = *i* (14, 23, 24).
Allelic series *I^h*, *I*, *i* (14, 26).
Intermedium spike-c = *int-c* (8, 9, 19, 20).
Six-rowed spike 5 = *v5* (*vrs5*) (27).
Hordeum vulgare *TEOSINTE BRANCHED 1* = *HvTB1* (22).

Inheritance:

Monofactorial recessive (6, 7, 20, 24, 27).
Located in chromosome 4HS (5, 7, 20, 24, 27); *Int-c.a* is about 13.1 cM proximal from the *fch9* (chlorina seedling 9) locus (3, 5, 6, 7, 13); *Int-c.a* is about 14.5 cM distal from the *Kap1* (Hooded lemma 1) locus (3, 5, 6, 7, 13); *Int-c.a* is about 3.5 cM from AFLP marker E4143-5 in subgroup 8 of the Proctor/Nudinka map (21); *int-c.5* is associated with SNP markers 1_0113 to 2_1122 (positions 26.58 to 47.80 cM) in 4H bins 02 to 05 of the Bowman backcross-derived line BW421 (2); *Int-c.a* is associated with SNP markers 1_0312 to 2_1374 (positions 38.41 to 41.13 cM) in 4H bins 03 to 04 of the Bowman backcross-derived line BW911, with SNP markers 1_0409 to 2_1122 (positions 5.24 to 47.80 cM) in 4H bins 01 to 05 of the Bowman backcross-derived line BW889 and with SNP markers 1_0132 to 2_0109 (positions 38.41 to 39.48 cM) in 4H bins 03 to 04 of the Bowman backcross-derived line BW624 (2); near SNP marker 2_0302 (22), in 4H bin 04.

Description:

Alleles at the *int-c* (*vrs5*) locus alter the size of lateral spikelets (1, 6, 22). The lemma apex of lateral kernels is rounded or weakly pointed, awnless or short-awned (1, 11, 18). Lower lateral spikelets may develop poorly in some *int-c* mutants (6), while seed development may occur in all lateral spikelets of others (8, 17). Variability in lateral spikelet development exists among the *int-c* mutants and environmental conditions can alter expressivity. The *Int-c.a* (formerly *I*) allele in six-rowed barley increases the size of lateral spikelets, while the *int-c.b* (formerly *i*) allele in two-rowed barley prevents anther development in lateral spikelets (11, 25). The *int-c.5* mutant in Bonus produces fertile stamens in lateral spikelets (11). In the presence of the *Int-c.h* (formerly *I^h*) allele of Mortoni, lateral spikelets are male fertile and may occasionally set seed (10, 14). Spikes of *vrs5.n* plants appear similar to those of six-rowed barley, but lateral spikelets are smaller (less than half the size of the central spikelets) and broader (5, 6). The *Int-c* (*HvTB1*) gene is an ortholog of the maize domestication gene *TEOSINTE BRANCHED 1* (*TB1*) (22). Seventeen barley mutations in the coding region for *HvTB1* were correlated with lateral spikelet fertility phenotypes (22). Compared to Bowman the backcross-derived lines for *int-c.5*, BW421, had slightly smaller leaf blades and smaller, lighter kernels, 4.2 vs. 5.6 mg. In spite of more kernels per spike (lateral spikelet fertility), the grain yields of BW421 were slightly lower than those of Bowman. (4) Compared to Bowman, tillering was increased in BW421 (22) The increased size of sterile lateral spikelets in Bowman backcross-derived lines for *Int-c.a*, BW421 and BW889, seems to

be associated with a slight increase in leaf blade size (4).

Origin of mutant:

The dominant allele, *Int-c.a*, is present in six-rowed barley (22) and wild barley (28); the recessive allele, *int-c.b*, is present in many two-rowed cultivars (22, 28); an X-ray induced mutant in Gamma 4 (5, 7).

Mutational events:

int-c.b (*i*) in many two-rowed barleys (26); *Int-c.h* (*h*) in Mortoni (Clho 2210, GSHO 72) (10, 14); *vrs5.n* (*v5*) (38X-197, OUM338, GSHO 776) in Gamma 4 (5, 7, 16); *int-c.5* (NGB 115423, GSHO 1765) in Bonus (NGB 14657, PI 189763) (17, 20); *int-c.7* (NGB 115425), -c.62 (NGB 116835), -c.63 (NGB 115481) in Bonus, -c.13 (NGB 115431), -c.15 (NGB 115433), -c.16 (NGB 115434), -c.18 (NGB 115436), -c.25 (NGB 115443), -c.29 (NGB 115447) in Foma (NGB 14659, Clho 11333), -c.33 (NGB 115451), -c.38 (NGB 115456), -c.45 (NGB 115463), -c.48 (NGB 115466), -c.49 (NGB 115467), -c.53 (NGB 115471), -c.56 (NGB 115474), -c.60 (NGB 115478) in Kristina (NGB14661, NGB 1500) (15); *int-c.70* (NGB 115488), -c.76 (NGB 115494), -c.78 (NGB 115496), -c.84 (NGB 115502) in Bonus, -c.95 (NGB 115513) in Hege (NGB 13692) (15).

Mutant used for description and seed stocks:

vrs5.n (GSHO 776, OUM338) in Gamma 4; *int-c.b* in *Hordeum distichon* var. *nigrinudum* (GSHO 988); *int-c.5* (GSHO 1765) in Bonus; *int-c.b* from Compana (Clho 5438) in Bonneville (Clho 7248)*6 (Clho 16176) (10); *vrs5.n* from Manabe Island (OUL067, GSHO 766) in Bowman (PI 483237)*5 (GSHO 2102, BW904, NGB 20788); *int-c.5* in Bowman*6 (GSHO 2003); *int-c.5* in Bowman*7 (BW421, NGB 20654); *Int-c.a* from SEN "sib" from CIMMYT in Bowman*4 (GSHO 2363); *Int-c.a* from SEN "sib" in Bowman*7 (BW889, NGB22322). Note that based on SNP markers the Bowman backcross-derived lines from Manabe (GSHO 2002, BW904, NGB 20788); do not have the *vrn5,n* gene.

References:

1. Åberg, E., and G.A. Wiebe. 1945. Irregular barley, *Hordeum irregulare* sp. nov. J. Washington Acad. Sci. 35:161-164.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. Plant Physiol. 155:617-627.
3. Forster, B.P. 1993. Coordinator's report: Chromosome 4. Barley Genet. Newsl. 22:75-77.
4. Franckowiak, J.D. (Unpublished).
5. Fukuyama, T. 1982. Locating a six-rowed gene *v5* on chromosome 4 in barley. Barley Genet. Newsl. 12:29-31.
6. Fukuyama, T., J. Hayashi, and R. Takahashi. 1975. Genetic and linkage studies of the five types of induced 'six-row' mutants. Barley Genet. Newsl. 5:12-13.
7. Fukuyama, T., R. Takahashi, and J. Hayashi. 1982. Genetic studies on the induced six-rowed mutants in barley. Ber. Ohara Inst. landw. Biol., Okayama Univ. 18:99-113.
8. Gustafsson, Å., and U. Lundqvist. 1980. Hexastichon and intermedium mutants in barley. Hereditas 92:229-236.
9. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. Hereditas 62:409-414.
10. Gymer, P.T. 1977. Probable allelism of *li* and *int-c* genes. Barley Genet. Newsl. 7:34-35.
11. Gymer, P.T. 1978. The genetics of the six-row/two row character. Barley Genet. Newsl. 8:44-46.

12. Hockett, E.A. 1985. Registration of two- and six-rowed isogenic Bonneville barley germplasm. *Crop Sci.* 25:201.
13. Jensen, J. 1987. Linkage map of barley chromosome 4. p. 189-199. *In* S. Yasuda and T. Konishi (eds.) *Barley Genetics V. Proc. Fifth Int. Barley Genet. Symp.*, Okayama. 1986. Sanyo Press Co., Okayama.
14. Leonard, W.H. 1942. Inheritance of fertility in the lateral spikelets of barley. *Genetics* 27:299-316.
15. Lundqvist, U. (Unpublished).
16. Lundqvist, U. 1991. Coordinator's report: Ear morphology genes. *Barley Genet. Newsl.* 20:85-86.
17. Lundqvist, U., and A. Lundqvist. 1988. Induced intermedium mutants in barley: origin, morphology and inheritance. *Hereditas* 108:13-26.
18. Nötzel, H. 1952. Genetische Untersuchungen an röntgeninduzierten Gerstenmutanten. *Kühn-Archiv* 66:72-132.
19. Nybom, N. 1954. Mutation types in barley. *Acta Agric. Scand.* 4:430-456.
20. Persson, G. 1969. An attempt to find suitable genetic markers for dense ear loci in barley I. *Hereditas* 62:25-96.
21. Pozzi, C., D. di Pietro, G. Halas, C. Roig, and F. Salamini. 2003. Integration of a barley (*Hordeum vulgare*) molecular linkage map with the position of genetic loci hosting 29 developmental mutants. *Heredity* 90:390-396.
22. Ramsay, L., J. Comadran, A. Druka, D.F. Marshall, W.T.B. Thomas, M. Macaulay, K. MacKenzie, C. Simpson, J. Fuller, N. Bonar, P.M. Hayes, U. Lundqvist, J.D. Franckowiak, T.J. Close, G.J. Muehlbauer, and R. Waugh. 2011. *INTERMEDIUM.C*, a modifier of lateral spikelet fertility in barley, is an ortholog of the maize domestication gene *TEOSINTE BRANCHED 1*. *Nature Genet.* 43:169-173.
23. Robertson, D.W. 1929. Linkage studies in barley. *Genetics* 14:1-36.
24. Robertson, D.W. 1933. Inheritance in barley. *Genetics* 18:148-158.
25. Ubisch, G. von. 1916. Beitrag zu einer Faktorenanalyse von Gerste. *Z. Indukt. Abstammungs. Vererbungs.* 17:120-152.
26. Woodward, R.W. 1947. The I^h , I , i alleles in *Hordeum deficiens* genotypes of barley. *J. Am. Soc. Agron.* 39:474-482.
27. Woodward, R.W. 1957. Linkages in barley. *Agron. J.* 49:28-32.
28. Youssef, H.M., M. Mascher, M.A. Ayoub, N. Stein, B. Kilian, and T. Schnurbusch. 2017. Natural diversity of inflorescence architecture traces cryptic domestication genes in barley (*Hordeum vulgare* L.) *Genet. Res. Crop Evol.* 64:843-853.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:200-201.

Revised:

U. Lundqvist and J.D. Franckowiak. 2007. *Barley Genet. Newsl.* 37:237-239.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:92-94.

Stock number: BGS 184
Locus name: Zeocriton 3
Locus symbol: Zeo3

Revised locus symbol:

Since the zeocriton spike phenotype was studied by Hayes and Harlan (8), the recommended locus name is Zeocriton 1 (*Zeo1*). The semi-dominant dense spike variants at the complex *Zeo1* locus exhibit a range of phenotypes associated with restricted elongation of the rachis internodes. Based on DNA sequencing, three distinct phenotypic groups of dense spike variants are associated with the *Hordeum vulgare* *APELATA2* (*AP2*)-like transcription factor (*HvAP2*) locus (10, 11). The phenotypic expressions were historically assigned different locus symbols, but each group is now associated with specific molecular changes in the *HvAP2* transcript (10). Alleles at this locus are assigned locus symbols: the *Zeo1* (BGS 082) mutants exhibit the most extreme phenotypes; the *Zeo2* variants have intermediate phenotypes; and the *Zeo3* variants may show a lesser degree in shortening of rachis internodes (10). Molecular markers and/or pedigree information are necessary to separate variants assigned to the *Zeo2* (BGS 614) and *Zeo3* (BGS 184) groups (10).

Previous nomenclature and gene symbolization:

Zeocriton with no gene symbol (4, 8).
Semidwarf mutant in Morex = Mo1 (15).
Zeocriton 2 = *Zeo2* (6).
Cleistogamy 1 = *cly1.c* (10, 11, 14).
Hordeum vulgare *APELATA2* (*AP2*)-like transcription factor = *HvAP2* (10, 11).

Inheritance:

Monofactorial semi-dominant (9, 13).
Located in chromosome 2HL (2, 13); *Zeo3.d* is associated with SNP markers 1_1118 to 2_0494 (positions 180.85 to 221.70 cM) in 2HL bins 11 to 14 of Bowman backcross-derived line BW933; *Zeo3.h* is associated with SNP markers 2_1125 to 2_0943 (positions 206.17 to 237.13 cM) in 2HL bins 13 to 14 of the Bowman backcross-derived line BW940; *Zeo3.j* is associated with SNP markers 2_1370 to 1_1023 (positions 199.54 to 224.35 cM) in 2HL bins 12 to 14 of the Bowman backcross-derived line BW936; *Zeo3.av* (formerly named *dsp.av*) is associated with SNP markers 1_1236 to 2_0561 (positions 184.46 to 247.96 cM) in 2HL bins 11 to 15 of the Bowman backcross-derived line BW369 (2); the *HvAP2* locus is near marker BOPA2_12_10579 and these Bowman lines have the same single base pair change in the microRNA172 binding site (9), in 2H bin 13. Bowman backcross-derived lines BW660 (*Zeo3.g*) and BW661 (*Zeo3.i*) have the same change at the microRNA172 binding site (9) and SNP markers associated with dense spike mutants *ert-c.1* and *ert-d.7* in chromosomes 3HL and 7HS, respectively (2).

Description:

Hayes and Harlan (8) reported that three genes controlled rachis internode length in their cross to Zeocriton. One of these genes produced strap-shaped spikes with shortened rachis (8). Identification of Bowman backcross-derived lines with the *Zeo3* gene was based on DNA sequencing of the *HvAP2* gene (10). A similar degree of rachis internode shortening was observed in Bowman backcross-derived lines with either the *Zeo2* or the *Zeo3* sequence changes in the *HvAP2* gene (6, 10). Mutants at *Zeo3* are

apparently in some cultivars identified as having small lodicules (closed flowering or cleistogamy) (9, 13). The original *Zeo3.h* (Mo1) mutant differed from the parental cultivar Morex (CIho 15773) in plant height, tiller number, grain yield, and grain quality (12). The Bowman backcrossed lines for *Zeo2* and *Zeo3* have similar rachis internode lengths, but variability among cultivars with the same genes was observed (10). The rachis internode length of plants in Bowman backcross-derived lines for *Zeo3* alleles averaged 3.3 mm compared to 4.5 mm for Bowman. In some test environments, the BW lines with the *Zeo3* genes had 1 to 2 more kernels per spike than Bowman, and kernel weights were slightly lower. No effects of the *Zeo3* gene on plant height and grain yield were observed (5). DNA sequencing showed that the *Zeo1* mutants occur in a *Hordeum vulgare* APELATA2 (*AP2*)-like transcription factor, *HvAP2*. The dense spike and cleistogamous (small lodicules) phenotypes are a consequence of a perturbed interaction between microRNA 172 (*Hv-miR172*) and its corresponding binding site on the mRNA from the *HvAP2* gene, which acts early in spike development to regulate turnover of *HvAP2* mRNA (10, 11).

Origin of mutant:

A naturally and induced mutants occurring variant in the fourth exon of the *HvAP2* locus (10, 11). A sodium azide induced mutant in Morex (CIho 15773) (15).

Mutational events:

Based on haplotypes similarity for in the *HvAP2* gene, the potential *Zeo3* alleles can be placed in one group (10, 11). The *Zeo3* mutant is present in Tammi (OUU059, PI 175505) and a few other cultivars (11) and in Harry (NGB 2666, PI 491575) and in several other cultivars (10). *Zeo3.d* was isolated from line with a male sterile mutant in P11 (CIho 15836) from Finland (1). The Bowman backcross-derived stocks with *Zeo3.g*, *Zeo3.i*, and *Zeo3.av* were isolated from Finnish cultivars Aapo (PI 467771) and Pokko (PI 467770) (2). *Zeo3.h* (WA11094-81, GSHO 1611) identified as Mo1 from Morex (CIho 15773) (7, 15), and *Zeo3.j* (SA121-4-5, GSHO 1612) from Glenn (CIho 15769) (4) were isolated as spike density variants from North American six-rowed cultivars. The SNP markers retained in Bowman backcross-derived lines, BW936 with *Zeo3.h* and BW940 with *Zeo3.j*, suggest that these two variants originated independent of the Bowman backcross-derived lines having Finnish cultivars as donor parents (2, 5).

Mutant used for description and seed stocks:

Zeo3.d from P11 (CIho 15836) in Bowman*7 (BW933, NGB 22362); *Zeo3.av* from Aapo (PI 467771) via Pokko (PI 467770) in Bowman*4 (BW269, NGB 22094); *Zeo3.g* and *Zeo3.i*, both with another dense spike gene, from Aapo (PI 467771) via Pokko (PI 467770) in Bowman*7 (BW660, NGB 22225; and BW661, NGB 22226), respectively; *Zeo3.h* (Mo1, Wa11094-81, GSHO 1611) (formerly listed as *Zeo2.h*) in Morex (CIho 15773); *Zeo3.h* in Bowman*5 (GSHO 1999); *Zeo3.h* from Morex in Bowman*8 (BW940, NGB 22369); *Zeo3.j* (SA121-4-5, GSHO 1612) from Glenn (CIho 15769) in Bowman*7 (GSHO 1882, BW936, NGB 22365).

References:

1. Ahokas, H. 1976. Male sterile mutants of barley. III. Additional inaperturate mutants. Barley Genet. Newsl. 6:4-6.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. Plant Physiol. 155:617-627.
3. Engledow, F.L. 1924. Inheritance in barley. III. The awn and the lateral floret (cont'd): fluctuation: a linkage: multiple allelomorphs. J. Genet. 14:49-87.

4. Faue, A.C. 1987. Chemical mutagenesis as a breeding tool for barley. M.S. Thesis. North Dakota State Univ., Fargo.
5. Franckowiak, J.D. (Unpublished).
6. Franckowiak, J.D. 1999. Coordinator's report: Semidwarf genes. *Barley Genet. Newsl.* 29:74-79.
7. Franckowiak, J.D., and A. Pecio. 1992. Coordinator's report: Semidwarf genes. A listing of genetic stocks. *Barley Genet. Newsl.* 21:116-127.
8. Hayes, H.K., and H.V. Harlan. 1920. The inheritance of the length of internode in the rachis of the barley spike. U.S. Dept. Agr., Bull. 869. 26 pp.
9. Honda, I., Y. Turuspekov, T. Komatsuda, and Y. Watanabe. 2005. Morphological and physiological analysis of cleistogamy in barley (*Hordeum vulgare*). *Physiol. Plant* 124:524-531.
10. Houston, K., S.M. McKim, J. Comadran, N. Bonar, I. Druka, N. Uzrek, E. Cirillo, J. Guzy-Wrobeliska, N.C. Collins, C. Halpin, M. Hansson, C. Dockter, A. Druka, and R. Waugh. 2013. Variation in the interaction between alleles of *HvAPETALA2* and microRNA172 determines the density of grains on the barley inflorescence. *Proc. Natl. Acad. Sci. USA* 110:16675-16680.
11. Nair, S.K., N. Wang, Y. Turuspekov, M. Pourkheirandish, S. Sinsuwongwat, G. Chen, M. Sameri, A. Tagiri, I. Honda, Y. Watanabe, H. Kanamori, T. Wicker, N. Stein, Y. Nagamura, T. Matsumoto, and T. Komatsuda. 2010. Cleistogamous flowering in barley arises from the suppression of microRNA-guided *HvAP2* mRNA cleavage. *Proc. Natl. Acad. Sci. USA* 107:490-495.
12. Nedel, J.L., S.E. Ullrich, J.A. Clancy, and W.L. Pan. 1993. Barley semidwarf and standard isotype yield and malting quality response to nitrogen. *Crop Sci.* 33:258-263.
13. Sameri, M., K. Takeda, and T. Komatsuda. 2006. Quantitative trait loci controlling agronomic traits in recombinant inbred lines from a cross of oriental- and occidental-type barley cultivars. *Breed. Sci.* 56:243-252.
14. Turuspekov, Y., Y. Mano, I. Honda, N. Kawada, Y. Watanabe, and T. Komatsuda. 2004. Identification and mapping of cleistogamy gene in barley. *Theor. Appl. Genet.* 109:480-487.
15. Ullrich, S.E., and A. Aydin. 1988. Mutation breeding for semi-dwarfism in barley. p. 135-144. *In* Semi-dwarf Cereal Mutants and Their Use in Cross-breeding III. IAEA-TECDOC-455. IAEA, Vienna.

Prepared:

J.D. Franckowiak. 2002. *Barley Genet. Newsl.* 32:99.

Revised:

J.D. Franckowiak and U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:95-97.

Stock number: BGS 188
Locus name: Aluminum tolerance 1
Locus symbol: *Alp1*

Previous nomenclature and gene symbolization:

Aluminum tolerance (4, 17).
Aluminum tolerance = *Alp* (15).
pH tolerance = *Pht* (18).
Aluminum tolerance = *Alt* (14).
Hordeum vulgare Al-activated citrate transporter 1 = *HvAACT1* (7).

Inheritance:

Incomplete dominant with inheritance pattern dependent on Al concentration (15, 16). Location in chromosome 4HL (13, 18); *Alp1.a* is about 25 cM from the *Kap1* (Hooded awn 1) locus (18); *Alp1.a* is 2.1 cM proximal to the marker *Xbcd1117* and 2.1 cM distal to the markers *Xwg464* and *Xcdo1395* (19); the *Alp1* locus is flanked by SSR markers Bmag353 and HVM68 (21); *Alp1.a* is flanked by microsatellite markers Bmac310 and Bmag353 (9); the 4H QTL for Al-tolerance is 0.8 cM from DArT marker bPb-6949 (3).

Description:

Better performance in acid or aluminum-toxic soils was observed among spring (10, 11, 17) and winter (4, 15, 16) barley cultivars. An association between root and shoot yields in greenhouse studies and grain yield in field studies was reported (16). Recommended screening conditions in nutrient solutions with 4 ppm aluminum (Al) and adjusted twice daily to pH 4.8 (5, 15). Retarded root elongation was observed in 4 to 5 days with roots thickened and more branched. Al-tolerant cultivars tended to raise the pH while the opposite trend was reported for Al-sensitive cultivars (5). Aluminum sensitivity of cultivars was determined by staining seedling roots after aluminum treatment with a hematoxylin staining solution (12). An Al-tolerant cultivar secreted citrate from the roots in response to Al treatment (9, 22). The differential acid soil response caused by alleles at the *Alp1* locus was associated with the *HvAACT1* (*Hordeum vulgare* Al-activated citrate transporter 1) gene (7; 21). Increased expression of the *HvAACT1* gene in East Asian cultivars was associated with the presence of a 1023-bp transposable-element like insertion in the 5' untranslated region (6).

Origin of mutant:

Natural occurrence in Dayton (PI 539129) from Composite Cross X (CIho 6625) (4, 15) and in Charlottetown 80 (CIho 2732) (10, 11).

Mutational events:15

Alp1.a in Dayton (PI 539129) (15); *Alp1.a* in Murasakimochi (6, 9); *Alp1.a* in Svanhals (NGB 1482, PI 5474) (2); *Alp1.a* in Honen (PI 307495) (21); Al-tolerance is present in Tibetan wild barley at the *Alp1* locus (3); *Alp1.a* in Cevada BR2 (1); a new allele (*Alp1.c*) at the *Alp1* locus was identified in Chinese barley CXHKSL (8).

Mutant used for description and seed stocks:

Alp1.a in Dayton; *Alp1.a* in Svanhals; *Alp1.a* in Murasakimochi; *Alp1.c* in CXHKSL.

References:

1. Bian, M., X. Jin, S. Broughton, X.Q. Zhang, G. Zhou, M. Zhou, G. Zhang, D. Sun, and C. Li. 2015. A new allele of acid soil tolerance gene from a malting barley variety. BMC Genet. 16:92. DOI 10.1186/s12863-015-0254-4.

2. Bian, M., I. Waters, S. Broughton, X.Q. Zhang, M. Zhou, R. Lance, D. Sun, and C. Li. 2013. Development of gene-specific markers for acid soil/aluminium tolerance in barley (*Hordeum vulgare* L.). *Mol Breed.* 32:155-164.
3. Cai, S., D. Wu, Z. Jabreen, Y. Huang, Y. Huang, and G. Zhang. 2013. Genome-wide association analysis of aluminium tolerance in cultivated and Tibetan wild barley. *PLoS ONE* 8(7): e69776.
4. Foy, C.D., W.H. Armiger, L.W. Briggles, and D.A. Reid. 1965. Differential aluminum tolerance of wheat and barley varieties in acid soils. *Agron. J.* 57:413-417.
5. Foy, C.D., A.L. Fleming, G.R. Burns, and W.H. Armiger. 1967. Characterization of differential aluminum tolerance among varieties of wheat and barley. *Proc. Soil Sci. Soc. Amer.* 31:513-521.
6. Fujii, M., K. Yokosho, N. Yamaji, D. Saisho, M. Yamane, H. Takahashi, et al. 2012. Acquisition of aluminium tolerance by modification of a single gene in barley. *Nat. Commun.* 3:713.
7. Furukawa, J., N. Yamaji, H. Wang, N. Mitani, Y. Murata, K. Sato, M. Katsuhara, K. Takeda, and J.F. Mal. 2007. An aluminum-activated citrate transporter in barley. *Plant Cell Physiol.* 48:1081-1091.
8. Ma, Y., C. Li, P.R. Ryan, S. Shabala, J. You, J. Liu, C. Liu, and M. Zhou. 2016. A new allele for aluminium tolerance gene in barley (*Hordeum vulgare* L.). *BMC Genomics* 17:186.
9. Ma, J.F., S. Nagao, K. Sato, H. Ito, J. Furukawa, and K. Takeda. 2004. Molecular mapping of a gene responsible for Al-activated secretion of citrate in barley. *J. Exp. Bot.* 55:1335-1341.
10. MacLean, A.A., and T.C. Chiasson. 1966. Differential performance of two barley varieties to varying aluminum concentration. *Can. J. Soil Sci.* 46:147-153.
11. MacLeod, L.B., and L.P. Jackson. 1967. Aluminum tolerance of two barley varieties in nutrient solution, peat, and soil culture. *Agron. J.* 59:359-363.
12. Minella, E., and M.E. Sorrells. 1992. Aluminum tolerance in barley: genetic relationships among genotypes of diverse origin. *Crop Sci.* 40:329-337.
13. Minella, E., and M.E. Sorrells. 1997. Inheritance and chromosome location of *Alp*, a gene controlling aluminum tolerance in "Dayton" barley. *Plant Breed.* 116:465-469.
14. Raman, H., J.S. Moroni, K. Sato, B. Read, and B.J. Scott. 2002. Identification of AFLP and microsatellite markers linked with an aluminium tolerance gene in barley (*Hordeum vulgare* L.). *Theor. Appl. Genet.* 105:458-464.
15. Reid, D.A. 1970. Genetic control of reaction to aluminum in winter barley. p. 409-413. *In* R.A. Nilan (ed.) *Barley Genetics II*. Proc. Second Int. Barley Genet. Symp., Pullman, WA, 1969. Washington State Univ. Press, Pullman.
16. Reid, D.A., G.D. Jones, W.H. Armiger, C.D. Foy, E.J. Koch, and T.M. Starling. 1969. Differential aluminum tolerance of winter barley varieties and selections in associated greenhouse and field experiments. *Agron. J.* 61:218-222.
17. Stølen, O. 1965. Investigations on the tolerance of barley varieties to high hydrogen-ion concentrations in soil. p. 81-107 in Royal Vet. Agr. Coll. Yearbook, Copenhagen.
18. Stølen, O., and S. Anderson. 1978. Inheritance of tolerance to low soil pH in barley. *Hereditas* 88:101-105.
19. Tang, Y., M.E. Sorrells, L.V. Kochian, and D.F. Garvin. 2000. Identification of RFLP markers linked to the barley aluminum tolerance gene *Alp*. *Crop Sci.* 40:778-782.
20. Wang, J., H. Raman, M. Zhou, P.R. Ryan, E. Delhaize, D.M. Hebb, N. Coombes, and N. Mendham. 2007. High resolution mapping of the *Alp* locus and identification of a

Barley Genetics Newsletter (2017): 46:50-187.

candidate gene *HvMATE* controlling aluminium tolerance in barley (*Hordeum vulgare* L.).
Theor. Appl. Genet. 115:265-276.

21. Wang, J., H. Raman, B. Read, M. Zhou, N. Mendham, and S. Venkatanagappa.
2006. Validation of an *Alt* locus for aluminium tolerance scored with eriochrome cyanine
R staining method in barley cultivar Honen (*Hordeum vulgare* L.). Australian J. Agric.
Res. 57:113-118.

22. Zhou, G., E. Delhaize, M. Zhou, and P.R. Ryan. 2013. The barley MATE gene,
HvAACT1, increases citrate efflux and Al(3+) tolerance when expressed in wheat and
barley. Ann. Bot. 112:603-612.

Prepared:

J.D. Franckowiak. 2017. Barley Genet. Newsl. 47:98-100.

Stock number: BGS 189
Locus name: Accordion rachis 2
Locus symbol: *Acr2*

Previous nomenclature and gene symbolization:

Lax spike = lax (4, 9, 11).
Accordion rachis = *acr* (7, 9, 12).
Accordion rachis 1 = *acr1.a* (6).
Accordion rachis b and c = *acr.b*, *acr.c* (3, 5).
Laxatum spike stocks = *Lax.aa*, *Lax.ab*, *Lax.am*, *lax.ao*, (5, 12).

Inheritance:

Monofactorial incomplete dominant for lax spike, one of three factors required for expression of accordion rachis trait (5, 12).
Located in chromosome 4HL (3); *Acr2.ab* is associated with SNP markers 1_0123 to 2_1018 (positions 158.82 to 183.54 cM) in 4HL of the Bowman backcross-derived line BW445; Bowman backcross-derived lines BW007, BW008, BW009, BW439, BW444, and BW456 have identical heterogeneous SNP markers in 4HL, but their segments from the donor parent are longer (3), in 4H bins 11 or 12.

Description:

Rachis internodes are elongated and kernels appear longer than normal. The *Acr2* gene is in the second of three chromosomal regions associated with expression of the accordion rachis trait in Bowman backcross-derived lines BW009 and BW439. The long rachis internode trait was specifically added to the original accordion rachis stock by R.I. Wolfe (12). Characteristics associated with variants at the *Acr2* locus are difficult to determine because only the Bowman backcross-derived line BW444 (*Lax.aa* or *Acr2.aa*) did not have more than one chromosome segment from its donor parent (3). Rachis internodes of BW444 plants were about 30% longer, 5.7 vs. 4.4 mm, than those of Bowman. Leaf blades were slightly longer and wider. Kernels were slightly longer and heavier, 61 vs. 58 mg. Grain yields were equal or slightly more than those of Bowman. The donor parent (FN280) of the BW444 line traces to lines developed by Mary L. Martini and Harry V. Harlan (8) in Idaho via crosses to six-rowed barley made by D.C. Rasmusson (10). The Bowman backcross-derived lines BW007 (*Acr2.b2*) and BW008 (*Acr2.c2*) had the identical segment of chromosome 4HL plus a 1H segment retained from their donor parents; however, the 1H segments in these lines differed for SNP marker patterns (3). The Bowman lines BW007 and BW008 are 10 to 15% taller than Bowman. The rachis internodes of BW007 are about 25% longer than those of Bowman, 4.7 vs. 3.7 mm., Burma Girl (GSHO 1071, *Acr2.b2*), the original stock for BW007, has several sterile rachis nodes at the base of the spike before spikelet development is initiated (4). BW007 had kernel weights that varied across experiments from smaller to slightly larger than those of Bowman. The rachis internodes of BW008 were nearly twice as long as those of Bowman, 8.2 vs. 4.5 mm, but they did not show the accordion-like bending associated with the accordion phenotype. Kernels of the BW008 line were slightly longer and heavier than those of Bowman (5). Two other Bowman backcross-derived lines, BW439 and BW456, have the 4HL SNP pattern associated with the *Acr2* locus. Both originate from the same donor stock AB1398 (GSHO 1115) from Aberdeen, Idaho and are assigned the gene symbol *Acr2.am*. BW456 was selected based on lax

spikes while BW439 was selected based on tall plants (5).

Origin of mutant:

A naturally occurring deviant possibly in Long Rachis Internode (CIho 6164), developed by M.L. Martini and H.V. Harlan (8), or Burma Girl (CIho 14831), donated to the USDA Small Grains Collection by G.A. Wiebe. The *Acr2* gene has been incorporated into several genetic stocks from that source.

Mutational events:

Acr2.b2 in Burma Girl (CIho 14831, GSHO 1071) (5, 12); *Acr2.aa* in FN280 (GSHO 1572) (5, 10); *Acr2.ab* in ACBV89B229 (GSHO 1617) and ACBV89B232 (GSHO 1573) (12); *Acr2.c2* in T188 (GSHO 1480) from R.W. Woodward (2); *Acr2.am* in AB 1398 [GSHO 1115, likely CIho 14785 from G.A. Wiebe (1)]; all alleles may have been derived from one stock (3, 5).

Mutant used for description and seed stocks:

Acr2.aa in FN280 (GSHO 1572); *Acr2.b2* in Burma Girl (GSHO 1071); *Acr2.a2* in ACBV89B229 (GSHO 1617); *Acr2.c2* in T188 (GSHO 1480); *Acr2.am* in AB 1398 (GSHO 1115); *Acr2.a2* from ACBV89229 in Bowman*4 (GSHO 1899); *Acr2.a2* in Bowman*7 (BW009, NGB 20417 and BW439, NGB 20671); *Acr2.aa* from FN280 in Bowman*4 (GSHO 2276); *Acr2.aa* in Bowman*5 (BW444, NGB 20676); *Acr2.ab* from ACBV89B232 (GSHO 1573) in Bowman*4 (GSHO 2277); *Acr2.ab* in Bowman*6 (BW445, NGB 20677); *Acr2.b2* from Burma Girl in Bowman*2 (GSHO 1898); *Acr2.b2* in Bowman*5 (BW007, NGB 20415); *Acr2.c2* from T188 in Bowman*6 (BW008, NGB 20416); *Acr2.am* from AB 1398 in Bowman*4 (BW456, NGB 20688).

References:

1. Bockelman, H. <http://www.ars-grin.gov/cgi-bin/npgs/acc/display.pl?1068768>.
2. Doney, D.L. 1961. An inheritance and linkage study of barley with special emphasis on purple pigmentation of the auricle. M.S. Thesis. Utah State Univ., Logan.
3. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
4. Enns, H. 1961. Inheritance and linkage studies in barley using chromosomal interchanges and marker stocks. Ph.D. Thesis. Univ. Saskatchewan. pp. 70.
5. Franckowiak, J.D. (Unpublished).
6. Franckowiak J.D. 2001. BGS 97, Accordion rachis 1, *acr1*. *Barley Genet. Newsl.* 32:85.
7. Haus, T.E. 1957. Inheritance of rachis internode length and an abnormal rachis type in barley. *Dissertation Abstr.* 49:179-180.
8. Martini, M.L., and H.V. Harlan. 1942. Barley freaks. *J. Hered.* 33:339-343.
9. Nilan, R.A. 1964. The cytology and genetics of barley, 1951-1962. *Monogr. Suppl.* 3, *Res. Stud.* Vol. 32, No. 1. Washington State Univ. Press, Pullman.
10. Rasmusson, D.C. (Personal communications).
11. Smith, L. 1951. Cytology and genetics of barley. *Bot. Rev.* 17:1-55, 133-202, 285-355.
12. Wolfe, R.I. 1990. (Personal communications).

Prepared:

J.D. Franckowiak 2010. *Barley Genet. Newsl.* 40:65-66.

Revised:

J.D. Franckowiak 2017. *Barley Genet. Newsl.* 47:101-102.

BGS 229, Curly dwarf 2, *cud2*

Stock number: BGS 229
Locus name: Curly dwarf 2
Locus symbol: *cud2*

Previous nomenclature and gene symbolization:

None.

Inheritance:

Monofactorial recessive (3).

Located in chromosome 1HL (3); *cud2.b* is about 14.6 cM proximal from the *Blp1* (Black lemma and pericarp 1) locus (3, 4); no SNP markers different than those of Bowman were retained in the Bowman backcross-derived line BW199 for *cud2.b* (1).

Description:

Lemmas and awns are curly or strongly twisted, and stem internodes are curved. Culms are short (1/2 normal), spikes are semi-compact, awns are short (2/3 normal length), and kernels are globe-shaped. Most leaves are short and slightly twisted (3). In the Bowman backcross-derived line for *cud2.b*, BW199, plants were short (3/5 of Bowman), the twisted leaf blades were slightly shorter, peduncles were coiled and about 1/2 normal length, rachis internodes were slightly shorter, and awns were twisted and 2/3 normal length. Compared to Bowman, kernels were short (80% as long) and slightly wider and were about 3/4 normal weight (4.2 vs. 5.5 mg) and test weights were about 25% lower. Grain yields of BW199 were 1/4 to 1/2 of those for Bowman (2).

Origin of mutant:

An ethyl methanesulfonate induced mutant in Akashinriki (OUJ659, PI 467400) (3).

Mutational events:

cud2.b (OUM112, GSHO 1712) in Akashinriki (OUJ659, PI 467400) (3).

Mutant used for description and seed stocks:

cud2.b (OUM112, GSHO 1712) in Akashinriki; *cud2.b* in Bowman (PI 483237)*2 (GSHO 2062); *cud2.b* in Bowman*7 (BW199, NGB 22031).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Hayashi, J., T. Konishi, I. Moriya, and R. Takahashi. 1984. Inheritance and linkage studies in barley VI. Ten mutant genes located on chromosomes 1 to 7, except 3. *Ber. Ohara Inst. landw. Biol., Okayama Univ.* 18:227-250.
4. Jensen, J. 1986. Coordinator's report: Chromosome 5. *Barley Genet. Newsl.* 16:32-34.

Prepared:

T. Konishi and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:227.

Revised:

J.D. Franckowiak. 2014. *Barley Genet. Newsl.* 44:111.

J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:103.

Stock number: BGS 238
Locus name: Breviaristatum-t
Locus symbol: *ari-t*

Previous nomenclature and gene symbolization:

Breviaristatum-25 = *ari-25* (3).

Inheritance:

Monofactorial recessive (2, 4).

Located in chromosome 1H (1); *ari-t.25* is associated with SNP markers between 1_0744 to 1_1326 (positions 39.84 to 82.35 cM) in 1H of the Bowman backcross-derived line BW032 (1), in 1H bins 05 to 08.

Description:

The awns of *ari-t* plants are 2/3 of normal length and slightly curly. Peduncles are coiled and less than half normal length and plants are 1/2 to 2/3 of normal height (4, 5). In the Bowman backcross-derived line for *ari-t.25*, BW032, leaf blades were shorter and narrower than those of Bowman. Kernels of BW032 were shorter and narrower, seed weights were about 20% lower, and seed yields were about 1/4 of normal (2). A new locus name was assigned based on the unique phenotype and lack of other breviaristatum mutants located in 1H.

Origin of mutant:

An ethylene oxide induced mutant in Bonus (NGB 14657, PI 189763) (4, 5).

Mutational events:

ari-t.25 (NGB 115873) in Bonus (NGB 14657, PI 189763) (5).

Mutant used for description and seed stocks:

ari-t.25 in Bonus (NGB 115873); *ari-t.25* in Bowman (PI 483237)*5 (BW032, NGB 20440).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Gustafsson, Å, A. Hagberg, U. Lundqvist and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Kucera, J., U. Lundqvist, and Å. Gustafsson. 1975. Inheritance of breviaristatum mutants in barley. *Hereditas* 80:263-278.
5. Lundqvist, U. (Unpublished).

Prepared:

J.D. Franckowiak and U. Lundqvist. 2010. *Barley Genet. Newsl.* 40:82.

Revised:

J.D. Franckowiak and U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:104.

Stock number: BGS 241
Locus name: Accordion rachis 3
Locus symbol: *Acr3*

Previous nomenclature and gene symbolization:

Lax spike = Lax (7, 8)
Accordion rachis = *acr* (6).
Accordion rachis c = *acr.c* (4).
Accordion rachis 1 = *acr1.a* (5).

Inheritance:

Monofactorial incomplete dominant for lax spike, the third factor required for expression of accordion rachis 1 (*acr1*) trait (3, 4).
Located in chromosome 1HL (3); *Acr3.a3* is associated with SNP markers 2_1357 to 2_0997 (positions 76.96 to 94.86 cM) in 1HL of the Bowman backcross-derived lines BW009 and BW439 (3), likely in 1H bin 08. Bowman backcross-derived lines BW007 with *Acr3.b3*, BW009 with *Acr3.a3*, BW438 with *Acr3.a*, and BW456 with *Acr3.am3* contain overlapping segments of variable length and SNP marker patterns in 1H from their donor parents (3).

Description:

Mutant plants have slightly elongated rachis internodes (1). The *Acr3* gene is in the third chromosomal region associated with expression of the accordion rachis trait in Bowman backcross derived lines BW009 and BW439. The contribution of the 1HL region to characteristics of the accordion phenotype is unknown. Only the line BW438 (*Lax.an* or *Acr3.an*) has a 1HL donor chromosome segment in a line lacking both the 2H and 4HL segments identified as containing the *acr1* (see BGS 097) and *Acr2* (see BGS 189) genes, respectively. Plants of the Bowman derived line BW438 lines were ranged from slightly shorter to slightly taller than Bowman and had slightly longer rachis internodes (4). Kernels of BW438 were slightly longer than those of Bowman, but weighed slightly less (5.3 vs. 6.0 mg). Grain yield of BW003 was about 2/3 that of Bowman (4).

Origin of mutant:

A naturally occurring deviant for lax spike from several sources (3, 4).

Mutational events:

Acr3.a3 in ACBV89B229 (GSHO 1617) and ACBV89B232 (GSHO 1573) (3, 9); *Acr3.b3* in Burma Girl (CIho 14831, GSHO 1071) (4, 9); *Acr2.am2* and *Acr3.an* in AB 1398 [GSHO 1115, likely CIho 14785 from G.A. Wiebe (1)]. *Acr3.c3* in T188 (GSHO 1480) from R.W. Woodward (2).

Mutant used for description and seed stocks:

Acr3.a3 in ACBV89B229 (GSHO 1617); *Acr3.b3* in Burma Girl (GSHO 1071); *Acr2.am2* and *Acr3.an* in AB 1398 (CIho 14785); *Acr3.an* from AB 1398 in Bowman*5 (BW438, NGB 20670); *Acr3.b3* from Burma Girl in Bowman *2 (GSHO 1898); *Acr3.b3* in Bowman*5 (BW007, NGB 20415); *Acr3.am3* from AB 1398 in Bowman*4 (BW456, NGB 20688); *Acr3.a3* plus *acr1.a* and *Acr2.a2* in Bowman*7 (BW009, NGB 20417 and BW439, NGB 20671); *Acr3.c3* in T188 (GSHO 1480); *Acr3.c3* from T188 in Bowman*6 (BW008, NGB 20416).

References:

1. Bockelman, H. <http://www.ars-grin.gov/cgi-bin/npgs/acc/display.pl?1068768>.

2. Doney, D.L. 1961. An inheritance and linkage study of barley with special emphasis on purple pigmentation of the auricle. M.S. Thesis. Utah State Univ., Logan.
3. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
4. Franckowiak, J.D. (Unpublished).
5. Franckowiak J.D. 2001. BGS 97, Accordion rachis 1, *acr1*. *Barley Genet. Newsl.* 32:85.
6. Haus, T.E. 1957. Inheritance of rachis internode length and an abnormal rachis type in barley. *Dissertation Abstr.* 49:179-180.
7. Nilan, R.A. 1964. The cytology and genetics of barley, 1951-1962. *Monogr. Suppl.* 3, *Res. Stud.* Vol. 32, No. 1. Washington State Univ. Press, Pullman.
8. Smith, L. 1951. Cytology and genetics of barley. *Bot. Rev.* 17:1-55, 133-202, 285-355.
9. Wolfe, R.I. 1990. (Personal communications).

Prepared:

J.D. Franckowiak. 2010. *Barley Genet. Newsl.* 40:85-86.

Revised:

J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:105-106.

BGS 258, Dense spike 9, *dsp9*

Stock number: BGS 258
Locus name: Dense spike 9
Locus symbol: *dsp9*

Revised locus symbol:

The *dsp9.i* mutant is an allele at the *ert-e* (erectoides-e) locus based on allelism tests (2), phenotypic expression (2) and retained SNP markers in 6HL of the Bowman backcross-derived line (BW279) (1). See BGS 266 for more information on the alleles at the *ert-e* locus.

Previous nomenclature and gene symbolization:

Dense spike = *la* (3).
Dense spike 9 = *l9* (3).

Inheritance:

Monofactorial recessive (3).
Located in chromosome 6HL (3), *dsp9.i* is about 27.2 cM distal from the *xnt5* (xantha seedling 5) locus (4); *dsp9.i* is over 26.9 cM distal from the *Aat2* (aspartate aminotransferase 2) locus (7); *dsp9.i* is associated with SNP markers 1_0645 to 2_1425 (positions 190.39 to 207.0 cM) in 6H bins 11 to 15 of the Bowman backcross-derived line BW279 (1).

Description:

Extremely dense spikes are easily distinguished from normal types when grown in the field or greenhouse (3). A photo of the original spike phenotypes is presented in Saisho and Takeda (6). The spike often emerges from the side of the sheath. Compared to Bowman, plants of the Bowman backcross-derived line for *dsp9.i*, BW279, were about 2/3 as tall and had very short rachis internodes, 2.3 vs. 4.6 mm. Peduncles averaged about 3/4 normal length, kernels were 10 to 20% lighter, and grain yields were about 2/3 those of Bowman (2).

Origin of mutant:

An ethyl methanesulfonate induced mutant in Akashinriki (OUJ659, PI 467400) (3).

Mutational events:

dsp9.i (OUM113, GSHO 1774) in Akashinriki (OUJ659, PI 467400) (3); *dsp9.j* (OUM106), *dsp9.k* (OUM107), *dsp9.l* (OUM115), *dsp9.m* (OUM118) in Akashinriki (5).

Mutant used for description and seed stocks:

dsp9.i in Akashinriki (GSHO 1774, OUM113); *dsp9.i* in Bowman (PI 483237)*7 (GSHO 2090, BW279, NGB 20563).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Konishi, T. 1973. Genetic analyses of EMS-induced mutants in barley. *Barley Genet. Newsl.* 3:28-31.
4. Konishi, T. 1978. New linkage data on chromosome 6 of barley. *Barley Genet. Newsl.* 8:71-72.
5. Konishi, T. (Unpublished).
6. Saisho, D., and K. Takeda. 2011. Barley: Emergence of a new research material of

- Barley Genetics Newsletter (2017): 46:50-187.*
- crop science. Plant Cell Physiol. 52:724-727.
7. Yoshimi, R., and T. Konishi. 1995. Linkage analysis of several isozyme loci in barley. Barley Genet. Newsl. 24:35-37.
- Prepared:
- T. Konishi. 1976. Barley Genet. Newsl. 6:132.
- Revised:
- T. Konishi and J.D. Franckowiak. 1997. Barley Genet. Newsl. 26:239.
- J.D. Franckowiak. 2013. Barley Genet. Newsl. 43:114-115.
- J.D. Franckowiak. 2017. Barley Genet. Newsl. 47:107-108.

BGS 266, Erectoides-e, *ert-e*

Stock number: BGS 266
Locus name: Erectoides-e
Locus symbol: *ert-e*

Previous nomenclature and gene symbolization:

Erectoides-17 = *ert-17* (3).

Dense spike = *la* (5).

Dense spike 9 = *l9* (5).

Dense spike 9 = *dsp9* (8).

Inheritance:

Monofactorial recessive (3, 5, 10).

Located in chromosome 6HL (4, 9, 10), *ert-e* is about 27.2 cM distal from the *xnt5* (xantha seedling 5) locus (4), over 26.9 cM distal from the *Aat2* (aspartate aminotransferase 2) locus (13); *ert-e.17* is associated with SNP markers 2_0467 to 2_0733 (positions 169.88 to 180.69 cM) in 6H bin 11 of the Bowman backcross-derived line BW307 (1); *dsp9.i* is associated with SNP markers 1_0645 to 2_1425 (positions 190.39 to 207.0 cM) in 6H bins 11 to 15 of the Bowman backcross-derived line BW279 (1).

Description:

Spikes are very compact with rachis internode length values from 1.2 to 1.5 mm. Plants are about 2/3 normal height. Partial fertility and reduced vigor are noted among *ert-e* mutants. The peduncle is very short and spikes often emerge from the side of the flag sheath (9, 11). A large deficiency of mutant plants is frequently noted in segregating populations (9). Spike density decreases greatly when plants are treated with gibberellic acid (GA₃) as the flag leaf emerges (12). The mutant *ert-e.17* is allelic to mutant *dsp9.i* (dense spike 9, see BGS 258) (2). Compared to Bowman, plants of the Bowman backcross-derived line for *ert-e.17*, BW307, were about 2/3 as tall and had very short rachis internodes, 2.0 vs. 4.4 mm. Peduncles were 3/4 normal length, kernels were 10 to 20% lighter, and grain yields were 1/3 to 1/2 those of Bowman (2).

Origin of mutant:

An X-ray induced mutant in Bonus (NGB 14657, PI 189763) (3); an ethyl methanesulfonate induced mutant in Akashinriki (OUJ659, PI 467400) (5).

Mutational events:

ert-e.17 (GSHO 477, NGB112619), *-e.65* (NGB 112664) in Bonus (NGB 14657, PI 189763) (2); *ert-e.94* (NGB 112693), *-e.143* (NGB 112742) in Bonus, *-e.331* (NGB112846), *-e.396* (NGB 114150) in Foma (NGB 14659, CIho 11333) (11); *dsp9.i* (OUM113) in Akashinriki (OUJ659, PI 467400) (5); *dsp9.j* (OUM106), *dsp9.k* (OUM107), *dsp9.l* (OUM115), *dsp9.m* (OUM118, GSHO 1774) in Akashinriki (7).

Mutant used for description and seed stocks:

ert-e.17 (GSHO 477, NGB 112619) in Bonus; *ert-e.17* in Bowman (PI 483237)*6 (GSHO 2091); *ert-e.17* in Bowman*7 (BW307, NGB 22103); *dsp9.i* (GSHO 1774) in Akashinriki; *dsp9.i* in Bowman (PI 483237)*7 (GSHO 2090, BW279, NGB 20563).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.

2. Franckowiak, J.D. (Unpublished).
3. Hagberg, A., Å. Gustafsson, and L. Ehrenberg. 1958. Sparsely contra densely ionizing radiations and the origin of erectoides mutants in barley. *Hereditas* 44:523-530.
4. Hagberg, A., G. Persson, and A. Wiberg. 1963. Induced mutations in the improvement of self-pollinated crops. p. 105-124. *In* E. Åkerberg and A. Hagberg (eds.) *Recent Plant Breeding Research. Svalöf 1946-1961*. Almqvist & Wiksell, Stockholm.
5. Konishi, T. 1973. Genetic analyses of EMS-induced mutants in barley. *Barley Genet. Newsl.* 3:28-31.
6. Konishi, T. 1978. New linkage data on chromosome 6 of barley. *Barley Genet. Newsl.* 8:71-72.
7. Konishi, T. (Unpublished).
8. Konishi, T., and J.D. Franckowiak. 1997. BGS 258, Dense spike 9, *dsp9*. *Barley Genet. Newsl.* 26:239.
9. Persson, G. 1969. An attempt to find suitable genetic markers for the dense ear loci in barley I. *Hereditas* 62:25-96.
10. Persson, G., and A. Hagberg. 1962. Linkage studies with the erectoides loci. *Barley Newsl.* 5:46-47.
11. Persson, G., and A. Hagberg. 1969. Induced variation in a quantitative character in barley. Morphology and cytogenetics of *erectoides* mutants. *Hereditas* 61:115-178.
12. Stoy, V., and A. Hagberg. 1967. Effects of growth regulators on ear density mutants in barley. *Hereditas* 58:359-384.
13. Yoshimi, R., and T. Konishi. 1995. Linkage analysis of several isozyme loci in barley. *Barley Genet. Newsl.* 24:35-37.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:246.

Revised as BGS 258:

T. Konishi and J.D. Franckowiak. 1997. BGS 258, Dense spike 9, *dsp9*. *Barley Genet. Newsl.* 26:239.

Revised:

J.D. Franckowiak and U. Lundqvist. 2007. *Barley Genet. Newsl.* 37:257-258.

U. Lundqvist and J.D. Franckowiak. 2013. *Barley Genet. Newsl.* 43:118-119.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:109-110.

BGS 275, Leafless 1, *lfl*

Stock number: BGS 275
Locus name: Leafless 1
Locus symbol: *lfl*

Previous nomenclature:

Leafless = *lfl* (3).
Leafless 1.a = *lfs1.a* (1).
Leafless 2.b = *lfs2.b* (1).

Inheritance:

Monofactorial recessive (2, 3).
Located in chromosomes 6HL (1, 2); *lfl1.b* is associated with SNP markers 1_0297 to 1_0128 (positions 85.71 to 180.85 cM) in 2H bins 07 to 11, with SNP markers 1_0676 to 1_1534 (positions 44.96 to 180.69 cM) in 6H bins 03 to 11, and with SNP markers 2_0247 to 2_1223 (positions 159.73 to 198.70 cM) in 7H bins 09 to 12 of the Bowman backcross-derived line BW477 (1); the F₂-derived line for *lfl1.a* (BW476) contains many markers from GSHO 609, but donor parents SNP markers were retained in both BW lines only in the long arm of chromosome 6H (1, 2).

Description:

The leafless 1 mutants are characterized by a reduction in the size of leaf blades. Leaf blades on lower culm phytomers are normal while those associated with upper phytomers of the culm are gradually shortened until the blade of the flag leaf is commonly less than 2 to 3 cm in length. Other morphological traits appear unaffected by the *lfl1* mutant (2, 4). The original leafless mutant (GSHO 609) showed a much stronger reduction in leaf blade development with only a few partially developed leaf blades on lower culm leaf sheathes. The nearly complete loss of leaf blades was associated with numerous tillers of which only a few elongating and producing spikes (2). The second leafless mutant *lfl1.b* could be an allele based on phenotypic in the BW lines, but allelism tests have not been conducted (2).

Origin of mutant:

An induced mutant (KM 255, GSHO 609) in Kanto Bansei Gall [this cultivar likely is Golden Melon (OUJ818, Clho 958)] based on notes from T. Tsuchiya (3).

Mutational events:

lfl1.a in Golden Melon (OUJ818, Clho 958) from Okayama University, Kurashiki, Japan (3); *lfl1.b* in an unidentified breeding line, which likely originated from KM 255, from the Institute of Plant Science and Resources, Okayama University, Kurashiki, Japan (4).

Mutant used for description and seed stocks:

lfl1.a (KM 255, GSHO 609) in Golden Melon (OUJ818, Clho 958); *lfl1.b* in an unidentified breeding line; *lfl1.a* from GSHO 609 in Bowman (PI 483237)*1 (BW 476, NGB 22155); *lfl1.b* from an unidentified breeding line in Bowman*2 (BW 477, NGB 22156).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Hang, A. 1995. List of genetic stocks in barley arranged in numerical order of BGS (Barley Genetic Stock) numbers. USDA-ARS National Small Grains Germplasm

Barley Genetics Newsletter (2017): 46:50-187.

Research Facility, Aberdeen, ID 83210. (Notes from T. Tsuchiya on the collection of barley genetic stocks at Colorado State University.)

4. Takeda, K. (Personal communications).

Prepared:

J.D. Franckowiak 2017. Barley Genet. Newsl. 47:111-112.

Stock number: BGS 314
Locus name Six-rowed spike 2
Locus symbol: *vrs2*

Previous nomenclature and gene symbolization:

Six-rowed spike 2 = *v2* (10).
Intermedium-b = *int-b* (6, 7, 13).

Inheritance:

Monofactorial recessive (4, 5, 6, 8, 12).
Located in chromosome 5HL (10, 11); *vrs2.e* is over 19.0 cM proximal from the *srh1* (short rachilla hair 1) locus (10, 11, 12); *vrs2.e* is associated with SNP markers 2_1421 to 1_0477 (positions 159.51 to 173.49 cM) in 5H bins 09 to 10 of the Bowman backcross-derived line BW901 (1); *int-b.3* is near the *raw1* (smooth awn 1) locus based on linkage drag (3); *int-b.3* is associated with SNP markers 2_0134 to 2_1355 (positions 163.29 to 234.98 cM) in 5H bins 10 to 13 of the Bowman backcross-derived line BW420 (1), likely in 5H bin 10. A high-resolution genetic map of the *vrs2* locus was developed using recombinants and barley BACs (13).

Description:

The spike appears similar to the ordinary six-rowed spike, but the lateral spikelets on the upper and lower portion of the spike are reduced in size, less fertile, and more or less pedicellated. Lateral spikelets are pointed with reduced awn length or awnless (4, 11, 12). Kernels of the Bowman backcross-derived line for *vrs2.e*, BW901, were 15 to 20% lighter than kernels of Bowman. BW901 plants exhibited reduced tillering and grain yield was about half that of Bowman (2). Plants of the backcross derived line for *int-b.3*, BW420, headed 2 to 3 days earlier than Bowman, but more lodging was observed. The BW420 kernels were slightly longer, but weighed less, 52 vs. 58 mg, compared to those of Bowman. Grain yields of BW420 were 10 to 30% of those recorded for Bowman (3). Late heading and prolonged development of BW901 was reported in Germany (13), but this was not observed in field tests in Australia, England, New Zealand, or the USA (3). During barley inflorescence and shoot development, the gene at the *Vrs2* locus encodes for a SHORT INTERNODES (SHI) transcriptional regulator (13).

Origin of mutant:

vrs2.e is an X-ray induced mutant in Svanhals (PI 5474, GSHO 784, OUU032) (10, 12); *int-b.3* is a neutron induced mutant in Bonus (NGB 14657, PI 189763) (9).

Mutational events:

vrs2.e (Kmut 27, OUM284, GSHO 773) in Svanhals (PI 5474, GSHO 784, OUU032) (12); *int-b.3* (NGB 115421, GSHO 1764), *-b.6* (NGB 115424) in Bonus (NGB 14657, PI 189763) (6, 8); *int-b.75* (NGB 115493) in Bonus (8); HOR19366 in the Genebank of the Institute of Plant Genetics and Crop Plant Research (IPK), Gatersleben, Germany. (isolated as spontaneous mutant in a cultivated landrace collection from USA) (13).

Mutant used for description and seed stocks:

vrs2.e (Kmut 27, GSHO 773, OUM284) in Svanhals; *vrs2.e* in Bowman (PI 483237)*4 (GSHO 2111), *vrs2.e* in Bowman*6 (BW901, NGB 22333); *int-b.3* (NGB 115421, GSHO 1764) in Bonus; *int-b.3* in Bowman (PI 483237)*6 (GSHO 2129); *int-b.3* in Bowman*7 (BW420, NGB 20653).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Franckowiak, J.D. 1995. Notes on linkage drag in Bowman backcross derived lines of spring barley. *Barley Genet. Newsl.* 24:63-70.
4. Fukuyama, T., J. Hayashi, I. Moriya, and R. Takahashi. 1972. A test for allelism of 32 induced six-rowed mutants. *Barley Genet. Newsl.* 2:25-27.
5. Fukuyama, T., R. Takahashi, and J. Hayashi. 1982. Genetic studies on the induced six-rowed mutants in barley. *Ber. Ohara Inst. landw. Biol., Okayama Univ.* 18:99-113.
6. Gustafsson, Å., and U. Lundqvist. 1980. Hexastichon and intermedium mutants in barley. *Hereditas* 92:229-236.
7. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
8. Lundqvist, U. (Unpublished).
9. Lundqvist, U., and A. Lundqvist. 1988. Induced intermedium mutants in barley: origin, morphology and inheritance. *Hereditas* 108:13-26.
10. Takahashi, R., J. Hayashi, and C. Hirao. 1968. Linkage studies. *Barley Newsl.* 11:59.
11. Takahashi, R., J. Hayashi, and I. Moriya. 1971. Linkage studies in barley. *Barley Genet. Newsl.* 1:51-58.
12. Takahashi, R., J. Hayashi, T. Konishi, and I. Moriya. 1972. Inheritance and linkage studies in barley. V. Locating of seven new mutant genes. *Ber. Ohara Inst. landw. Biol., Okayama Univ.* 15:147-168.
13. Youssef, H.M., K. Eggert, R. Koppolu, A.M. Alqudah, N. Poursarebani, A. Fazeli, S. Sakuma, A. Tagiri, T. Rutten, G. Govind, U. Lundqvist, A. Graner, T. Komatsuda, N. Sreenivasulu, and T. Schnurbusch. 2017. VRS2 regulates hormone-mediated inflorescence patterning in barley. *Nat. Genet.* 49:157-161.

Prepared:

T. Fukuyama. 1983. *Barley Genet. Newsl.* 13:112.

Revised:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:263.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:113-114.

Stock number: BGS 315
Locus name: Six-rowed spike 3
Locus symbol: *vrs3*

Previous nomenclature and gene symbolization:

Six-rowed spike 3 = *v3* (5).
Intermedium spike-a = *int-a* (8, 9, 12, 17).
Intermedium spike-g = *int-g* (8, 9, 14).

Inheritance:

Monofactorial recessive (5, 7, 8, 12, 14).
Located in chromosome 1HL (6, 10, 18); *int-a.2* is near the centromere and proximal from the *ert-b* (erectoides-b) locus (10, 18); *int-a.1* is associated with SNP markers 2_0619 to 1_1361 (positions 50.96 to 82.35 cM) in 1H of the Bowman backcross-derived line BW419 (2); *vrs3.f* is associated with SNP markers 1_0575 to 1_0075 (positions 37.21 to 82.35 cM) in 1H of the Bowman backcross-derived line BW902 (2); *int-a.1* is located between molecular markers 1_0520 and 2_1217 [82.35 to 81.26 cM based on the scale in (2)] in 1H bin 08 (1); only one gene, MLOC_69611.1 located at 47.52 cM, carried a unique mutation in both BW419 (*int-a.1*) and BW902 (*vrs3.f*) (1, 3).

Description:

The upper half of the spike appears six-rowed, but lateral spikelets in the lower half are reduced in size. The basal portion of the spike appears two-rowed. Awns on the lateral spikelets range from normal near the top, to awnletted or pointed in the middle, to awnless at the base of the spike. Seeds may develop in lateral spikelets of only the upper two-thirds of the spike. Lateral spikelets may be pedicellated in some stocks. Central spikelets often have double awns, one on the lemma and one on the palea. The rachilla may be deformed (5, 6, 7, 8, 14, 16). In Bowman backcross-derived lines for *int-a.1* and *vrs3.f*, BW419 and BW902 respectively, plants were often slightly shorter, rachis internodes were slightly longer, and kernels were thinner than those of Bowman. Kernel weights for BW419 ranged from 70 to 85% of normal compared to only 60 to 75% for those of BW902. Grain yields of Bowman lines BW419 and BW902 were often lower than those of Bowman (4). Compared to Bowman, BW419 and BW902 had more kernels per spike, but tiller numbers and seed weights were reduced (11). The *VRS3* gene encodes a putative histone Lys demethylase with a conserved zinc finger and Jumonji C and N domain (1, 3). Comparative transcriptome analysis of the row-type mutants at the *Vrs3*, *Vrs4*, and *Int-c* (*Vrs5*) loci confirmed that all three genes act as transcriptional activators of *Vrs1* variants and produce quantitative variation in the expression levels of *VRS1* and differences in the number of developed lateral spikelets (3). The naturally occurring variants *Vrs3.w* (Haplotype 45) and *Vrs3.x* (Haplotype 46) were identified as present predominantly in two-rowed spring barleys and six-rowed winter barleys, respectively (1), and could explain the row-type associations with winter and spring growth habit (1, 15, 19). Combining a *vrs3* mutant allele (*int-a.1* or *vrs3.f*) with natural six-rowed alleles of *vrs1* and *vrs5* (*int-c*) loci leads to increased lateral grain size and greater grain uniformity (1).

Origin of mutant:

A gamma-ray induced mutant in Hakata 2 (OUJ807, PI 263407) (5, 7); an X-ray induced mutant in Bonus (NGB 14657, PI 189763) (12).

Mutational events:

vrs3.f (Kmut 213, OUM292, GSHO 774) in Hakata 2 (OUJ807, PI 263407), *vrs3.g* (Xb 388.8, Piro-1), *vrs3.h* (Xd 160.46, Piro-3), *vrs3.i* (X15 1463, Piro-5), *vrs3.j* (X17 3555, Piro-8) in Pirolina (PI 539132) (7); *int-a.1* (NGB 115419) in Bonus (NGB 14657, PI 189763) (13); *int-a.2* (NGB 115420) in Bonus (14, 17); *int-a.8* (NGB 115426), *-a.9* (NGB 115427), *-a.10* (NGB 115328), *-a.64* (NGB 115482) in Bonus, *-a.14* (NGB 115432), *-a.17* (NGB 115435), *-a.21* (NGB 115439), *-a.27* (NGB 115445), *-a.30* (NGB 115448), *-a.31* (NGB 115449), *-a.32* (NGB 115450), *-a.34* (NGB 115452), *-a.35* (NGB 115453), *-a.37* (NGB 115455) in Foma (NGB 14659, Clho 11333), *-a.46* (NGB 115464), *-a.51* (NGB 115469), *-a.52* (NGB 115470), *-a.54* (NGB 115472), *-a.55* (NGB 15473), *-a.59* (NGB 115477), *-a.61* (NGB 115479) in Kristina (NGB14661, NGB 1500) (14); *int-a.71* (NGB 115489), *-a.74* (NGB 115492), *-a.77* (NGB 115495), *-a.79* (NGB 115497), *-a.86* (NGB 115504), *-a.88* (NGB 115506) in Bonus, *-a.102* (NGB 115529), *-a.103* (NGB 115521) in Hege (NGB 13692) (13); wildtype alleles *Vrs3.w* [Haplotype 45 in Morex (Clho 15773)] and *Vrs3.x* [Haplotype 46 in Bowman (PI 483237)] were present predominantly in two-rowed spring barleys and six-rowed winter barleys, respectively (1).

Mutant used for description and seed stocks:

vrs3.f (GSHO 774) in Hakata 2; *int-a.1* (GSHO 170, NGB 115419) in Bonus; *vrs3.f* in Bowman (PI 483237)*7 (GSHO 2056, BW902, NGB 22334); *int-a.1* in Bowman*5 (GSHO 2055); *int-a.1* in Bowman*7 (BW419, NGB 20652).

References:

1. Bull, H., M.C. Casao, M. Zwirek, A.J. Flavell, W.T.B. Thomas, W. Guo, R. Zhang, P. Rapazote-Flores, S. Kyriakidis, J. Russell, A. Druka, S.M. McKim, and R. Waugh. 2017. Barley *SIX-ROWED SPIKE3* encodes a putative Jumonji C-type H3K9me2/me3 demethylase that represses lateral spikelet fertility. *Nat. Commun.* 8:936.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. 2. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
3. Esse, G.W. van, A. Walla, A. Finke, M. Koornneef, A. Pecinka, and M. von Korff. 2017. Six-rowed spike3 (VRS3) is a histone demethylase that controls lateral spikelet development in barley. *Plant Physiology* 174:2397-2408.
4. Franckowiak, J.D. (Unpublished).
5. Fukuyama, T., J. Hayashi, I. Moriya, and R. Takahashi. 1972. A test for allelism of 32 induced six-rowed mutants. *Barley Genet. Newsl.* 2:25-27.
6. Fukuyama, T., J. Hayashi, and R. Takahashi. 1975. Genetic and linkage studies of the five types of induced 'six-row' mutants. *Barley Genet. Newsl.* 5:12-13.
7. Fukuyama, T., R. Takahashi, and J. Hayashi. 1982. Genetic studies on the induced six-rowed mutants in barley. *Ber. Ohara Inst. landw. Biol., Okayama Univ.* 18:99-113.
8. Gustafsson, Å., and U. Lundqvist. 1980. Hexastichon and intermedium mutants in barley. *Hereditas* 92:229-236.
9. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
10. Jensen, J., and J.H. Jørgensen. 1975. The barley chromosome 5 linkage map. I. Literature survey and map estimation procedure. *Hereditas* 80:5-16.
11. Liller, C.B., R. Neuhaus, M. von Korff, M. Koornneef, and W. van Esse. 2015. Mutations in barley row type genes have pleiotropic effects on shoot branching. *PLoS One* 10: e0140246.

12. Lundqvist, U. 1991. Coordinator's report: Ear morphology genes. *Barley Genet. Newsl.* 20:85-86.
13. Lundqvist, U. (Unpublished).
14. Lundqvist, U., and A. Lundqvist. 1988. Induced intermedium mutants in barley: origin, morphology and inheritance. *Hereditas* 108:13-26.
15. Muñoz-Amatriaín, M., A. Cuesta-Marcos, J.B. Endelman, J. Comadran, J.M. Bonman, H.E. Bockelman, S. Chao, J. Russell, R. Waugh, P.M. Hayes. and G.J. Muehlbauer. 2014. The USDA barley core collection: genetic diversity, population structure, and potential for genome-wide association studies. *PLoS ONE* 9, e94688.
16. Nötzel, H. 1952. Genetische Untersuchungen an röntgeninduzierten Gerstenmutanten. *Kühn-Archiv* 66:72-132.
17. Nybom, N. 1954. Mutation types in barley. *Acta Agric. Scand.* 4:430-456.
18. Persson, G. 1969. An attempt to find suitable genetic markers for dense ear loci in barley I. *Hereditas* 62:25-96.
19. Ramsay, L., J. Comadran, A. Druka, D.F. Marshall, W.T.B. Thomas, M. Macaulay, K. MacKenzie, C. Simpson, J. Fuller, N. Bonar, P.M. Hayes, U. Lundqvist, J.D. Franckowiak, T.J. Close, G.J. Muehlbauer, and R. Waugh. 2011. *INTERMEDIUM.C*, a modifier of lateral spikelet fertility in barley, is an ortholog of the maize domestication gene *TEOSINTE BRANCHED 1*. *Nat. Genet.* 43:169-173.

Prepared:

T. Fukuyama. 1983. *Barley Genet. Newsl.* 13:113.

Revised:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:264-265.

J.D. Franckowiak and U. Lundqvist. 2010. *Barley Genet. Newsl.* 40:90-92.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:115-117.

Stock number: BGS 320
Locus name: Intermedium spike-b
Locus symbol: *int-b*

Revised locus symbol:

The *int-b* mutants are alleles at the *vrs2* (six-rowed spike 2) locus based on DNA sequencing (9). Although phenotypic expression of spike type is somewhat between *int-b* and *vrs2* in Bowman backcross-derived lines BW420 and BW901, respectively (4), SNP molecular marker are retained in the same region of 5HL in both lines (3). See BGS 314 for more information about the alleles at the *vrs2* locus.

Previous nomenclature and gene symbolization:

None.

Inheritance:

Monofactorial recessive (6, 7).

Located in chromosome 5HL (5); *int-b.3* is near the *raw1* (smooth awn 1) locus based on linkage drag (5); *int-b.3* is associated with SNP markers 2_0134 to 2_1355 (positions 163.29 to 234.98 cM) in 5H bins 10 to 13 of the Bowman backcross-derived line BW420 (3), likely in 5H bin 10.

Description:

The spike appears similar to the six-rowed spike, but developmental irregularities occur commonly in the lower portion of the spike. All lateral spikelets are reduced in size, and their lemma awns are short or reduced to a pointed tip. Commonly, only lateral spikelets in the middle of the spike set seed (6, 9). Plants of the Bowman backcross-derived line for *int-b.3*, BW420, showed reduced vigor and tillering (1). Plants of BW420 headed 2 to 3 days earlier than Bowman, and more lodging was observed. The BW420 kernels were slightly longer, but weighed less, 52 vs. 58 mg, compared to those of Bowman. Grain yields of BW420 were 10 to 30% of those recorded for Bowman (4). Combining the *int-b.3* and *cul2.b* (uniculm 2) mutants produced unculm plants having spikes without spikelets (1). The double mutant combination, *int-b.3* and low number of tillers 1 (*Int1.a*), produced unculm plants (2). The *Vrs2* (*Int-b*) locus encodes for a SHORT INTERNODES (SHI) transcriptional regulator during barley inflorescence and shoot development (10).

Origin of mutant:

A neutron induced mutant in Bonus (NGB 14657, PI 189763) (9).

Mutational events:

int-b.3 (NGB 115421, GSHO 1764), *-b.6* (NGB 115424) in Bonus (NGB 14657, PI 189763) (6, 9); *int-b.75* (NGB 115493) in Bonus (8).

Mutant used for description and seed stocks:

int-b.3 (NGB 115421, GSHO 1764) in Bonus; *int-b.3* in Bowman (PI 483237)*6 (GSHO 2129); *int-b.3* in Bowman*7 (BW420, NGB 20653).

References:

1. Babb, S., and G.J. Muehlbauer. 2003. Genetic and morphological characterization of the barley unculm2 (*cul2*) mutant. *Theor. Appl. Genet.* 106:846-857.
2. Dabbert, T., R.J. Okagaki, S. Cho, S. Heinen, J. Boddu, and G.J. Muehlbauer. 2010. The genetics of barley low-tillering mutants: *low number of tillers-1* (*Int1*). *Theor. Appl. Genet.* 121:705-715.

3. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
4. Franckowiak, J.D. (Unpublished).
5. Franckowiak, J.D. 1995. Notes on linkage drag in Bowman backcross derived lines of spring barley. *Barley Genet. Newsl.* 24:63-70.
6. Gustafsson, Å., and U. Lundqvist. 1980. Hexastichon and intermedium mutants in barley. *Hereditas* 92:229-236.
7. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
8. Lundqvist, U. (Unpublished).
9. Lundqvist, U., and A. Lundqvist. 1988. Induced intermedium mutants in barley: origin, morphology and inheritance. *Hereditas* 108:13-26.
10. Youssef, H.M., K. Eggert, R. Koppolu, A.M. Alqudah, N. Poursarebani, A. Fazeli, S. Sakuma, A. Tagiri, T. Rutten, G. Govind, U. Lundqvist, A. Graner, T. Komatsuda, N. Sreenivasulu, and T. Schnurbusch. 2017. VRS2 regulates hormone-mediated inflorescence patterning in barley. *Nature Genet.* 49:157-161.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:268.

Revised:

U. Lundqvist and J.D. Franckowiak. 2014. *Barley Genet. Newsl.* 44:118-119.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:118-119.

Stock number: BGS 328
Locus name: Breviaristatum-e
Locus symbol: *ari-e*

Previous nomenclature and gene symbolization:

Breviaristatum-1 = *ari-1* (6, 7).
Short awn 9 = *lk9* (20).
Golden Promise erectoides = *GPert* (19).
Hordeum vulgare dense and erect panicle 1 = *Hvdep1* (21).

Inheritance:

Monofactorial recessive (6).
Located in chromosome 5HL (12, 13, 14, 15, 19); *ari-e.1* is about 5.6 cM proximal from the *cer-zj* (eceriferum-zj) locus (17, 18); *ari-e.GP* is near SSR molecular marker Bmag337 (3); *ari-e.GP* is close to SNP marker 2_1239 (11); *ari-e.1* is associated with SNP markers 2_1480 and 3_0285 (position 137.96 cM) in 5H bin 08 of the Bowman backcross-derived line BW042 (1); *ari-e.GP* is associated with SNP markers 1_1198 to 2_0449 (positions 73.70 to 154.37 cM) in 5H bins 02 to 09 of the Bowman backcross-derived line BW043 (1), in 5H bin 06. The *ari-e* locus is positioned at 52.30 cM on the barley physical map (9, 22). The *ari-e.GP* gene was fine mapped to a 0.58 Mb interval on the POPSEQ physical map (7).

Description:

Awns are about 2/3 normal length, plants are 3/4 to 5/6 normal height, and kernels are smaller (8, 19). Awns and spikes are more erect than those of normal sibs (4). Mutant alleles at the *ari-e* locus are associated with salt tolerance, lower accumulation of Na⁺ (2, 3, 12), and show relative insensitivity to GA₃ (12). The *ari-e.GP* mutant is described as an elongation (*elo*) type of semidwarf mutant based on cell size reduction in leaf blades (2). Plants of the Bowman backcross-derived lines for *ari-e.1* (BW042) and *ari-e.GP* (BW043) are about 3/4 as tall as Bowman and have awns that are about half as long. Rachis internodes and leaf blades are slightly shorter. Kernels of BW042 and BW043 were about 10% shorter than those of Bowman and weighed about 10% less. Grain yields of BW042 and BW043 were often less than those of Bowman (5). The *ari-e.GP* mutant compared to Maythorpe headed 1.5 days earlier. The effect of the *ari-e.GP* mutant on grain yield per plant was greater in Bowman genetic background than in Maythorpe background (22). The *ari-e* mutants are loss-of-function mutations in the *Hordeum vulgare* dense and erect panicle 1 (*HvDep1*) gene, an Arabidopsis heterotrimeric G protein (AGG3)-type γ-subunit encoding gene that regulates culm elongation and seed size (22). The *HvDep1* coding sequence consists of five exons and four introns (22).

Origin of mutant:

A neutron induced mutant in Bonus (NGB 14657, PI 189763) (8).

Mutational events:

ari-e.1 (NGB 115846, GSHO 1653) in Bonus (NGB 14657, PI 189763) (8); *ari-e.30* (NGB 115879), *-e.39* (NGB 115889) in Bonus (10); *ari-e.119* (NGB 115931), *-e.156* (NGB 115966) in Foma (NGB 14659, Clho 11333) (8); *ari-e.166* (NGB 115976), *-e.178* (NGB 115988), *-e.222* (NGB 116301) in Foma (10); *ari-e.228* (NGB 116038) in Foma (8); *ari-e.GP* (Golden Promise, PI 343079, GSHO 1733) in Maythorpe (PI 260893) (3, 4, 16),

but PI 260893 has three genomic regions that differ from Golden Promise (21). Based on DNA sequencing of the *HvDep1* gene, the *ari-e.178*, *ari-e.222*, and *ari-e.228* mutants could not be confirmed as being *ari-e* mutants (22).

Mutant used for description and seed stocks:

ari-e.1 (NGB 115846, GSHO 1653) in Bonus; *ari-e.GP* in Maythorpe (GSHO 1733); *ari-e.1* in Bowman (PI 483237)*7 (GSHO 2104, BW042, NGB 20450); *ari-e.GP* from Golden Promise in Bowman*7 (GSHO 2105, BW043, NGB 20451).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Ellis, R.P., B.P. Forster, D.C. Gordon, L.L. Handley, R.P. Keith, P. Lawrence, R. Meyer, W. Powell, D. Robinson, C.M. Scrimgeour, G. Young and W.T.B. Thomas. 2002. Phenotype/genotype associations for yield and salt tolerance in a barley mapping population segregating for two dwarfing genes. *J. Exp. Bot.* 53:1163-1176.
3. Forster, B.P., H. Pakniyat, M. Macaulay, W. Matheson, M.S. Phillips, W.T.B. Thomas, and W. Powell. 1994. Variation in the leaf sodium content of the *Hordeum vulgare* (barley) cultivar Maythorpe and its derived mutant c.v. Golden Promise. *Heredity* 73:249-253.
4. Franckowiak, J.D. 1992. Allelism tests among selected semidwarf barleys. *Barley Genet. Newsl.* 21:17-23.
5. Franckowiak, J.D. (Unpublished).
6. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
7. Jia, Q., C. Tan, J. Wang, X.-Q. Zhang, J. Zhu, H. Luo, J. Yang, S. Westcott, S. Broughton, D. Moody, and C. Li. 2016. Marker development using SLAF-seq and whole-genome shotgun strategy to fine-map the semi-dwarf gene *ari-e* in barley. *BMC Genomics* 17:911.
8. Kucera, J., U. Lundqvist, and Å. Gustafsson. 1975. Inheritance of *breviaristatum* mutants in barley. *Hereditas* 80:263-278.
9. Liu, H., M. Bayer, A. Druka, J.R. Russell, C.A. Hackett, J. Poland, L. Ramsay, P.E. Hedley, and R. Waugh. 2014. An evaluation of genotyping by sequencing (GBS) to map the *Breviaristatum-e* (*ari-e*) locus in cultivated barley. *BMC Genomics* 15:104.
10. Lundqvist, U. (Unpublished).
11. Malosetti, M., F.A. van Eeuwijk, M.P. Boer, A.M. Casas, M. Elía, M. Moralejo, P.R. Bhat, L. Ramsay, and J.-L. Molina-Cano. 2011. Gene and QTL detection in a three-way barley cross under selection by a mixed model with kinship information using SNPs. *Theor. Appl. Genet.* 122:1605-1616.
12. Pakniyat, H., E. Baird, W.T.B. Thomas, P.D.S. Caligari, W. Powell, and B.P. Forster. 1996. Effect of semi-dwarf mutants on salt tolerance in barley. p. 660-661. *In* A.E. Slinkard, G.J. Scoles, and B.G. Rosnagel (eds.) *Proc. Fifth Int. Oat Conf. & Seventh Int. Barley Genet. Symp.*, Saskatoon. Univ. of Saskatchewan, Saskatoon.
13. Persson, G. 1969. An attempt to find suitable genetic markers for the dense ear loci in barley I. *Hereditas* 62:25-96.
14. Persson, G. 1969. An attempt to find suitable genetic markers for the dense ear loci in barley II. *Hereditas* 63:1-28.
15. Persson, G., and A. Hagberg. 1965. Localization of nine induced mutations in the barley chromosomes. *Barley Newsl.* 8:52-54.

16. Sigurbjörsson, B., and A. Micke. 1969. Progress in mutation breeding. p. 673-697. *In* Proceedings of an international symposium on the nature, induction and utilization of mutation in plants. Vienna: International Atomic Energy Agency.
17. Søggaard, B. 1974. Three-point tests on chromosome 1 and 7. *Barley genet. Newsl.* 4:70-73.
18. Søggaard, B. 1977. The localization of eceriferum loci in barley. IV. Three point tests of genes on chromosome 7 in barley. *Carlsberg Res. Commun.* 42:35-43.
19. Thomas, W.B.T., W. Powell, and W. Wood. 1984. The chromosomal location of the dwarfing gene present in Golden Promise. *Heredity* 53:177-183.
20. Tsuchiya, T. 1974. Allelic relationships of genes for short-awned mutants in barley. *Barley Genet. Newsl.* 4:80-81.
21. Walia, H., C. Wilson, P. Condamine, A.M. Ismail, J. Xu, X. Cui, and T.J. Close. 2007. Array-based genotyping and expression analysis of barley cv. Maythorpe and Golden Promise. *BMC Genomics.* 2007; 8:87.
22. Wendt, T., I. Holme, C. Dockter, A. Preuß, W. Thomas, A. Druka, R. Waugh, M. Hansson, and I. Braumann. 2016. *HvDep1* Is a positive regulator of culm elongation and grain size in barley and impacts yield in an environment-dependent manner. *PLoS ONE* 11(12):e0168924.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:276.

Revised:

U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:131-132.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:120-122.

Stock number: BGS 332
Locus name: Erectoides-r
Locus symbol: *Ert-r*

Revised locus symbol:

The *Ert-r.52* mutant is an allele at the *Zeo1* (Zeocriton 1) locus based on phenotypic expression and retained SNP markers in 2HL of the Bowman backcross-derived lines BW 322 with *Ert-r.52*, the BW937 line with *Zeo1.a*, and the BW938 line with *Zeo1.b* (1) and DNA sequencing of the *HvAP2* gene (5). Historically, the recommended locus symbol is *Zeo1* based on descriptions by Hayes and Harlan (4). See BGS 082 for more information the alleles at the *Zeo1* locus.

Previous nomenclature and gene symbolization:

Erectoides-52 = *ert-52* (2).

Hordeum vulgare APELATA2 (*AP2*)-like transcription factor = *HvAP2* (5).

Inheritance:

Monofactorial incomplete dominant (3).

Located in chromosome 2HL (1); *Ert-r.52* is associated with SNP markers 2_0715 to 1_0551 (positions 213.08 to 221.70 cM) in 2HL of the Bowman backcross-derived line BW322, in 2H bin 13 (1).

Description:

Spikes are compact in heterozygotes and very compact in homozygotes, with rachis internode length values from 1.4 to 1.8 mm. Homozygotes are about 2/3 normal height with excellent vigor. The glumes associated with lateral spikelets are 3 to 4 times larger than normal. Lodicule size is reduced (8). Heterozygotes are intermediate in plant height, have slightly more lax spikes, and have normal glumes in lateral spikelets (8). GA₃ treatment of plants as the flag leaf emerges decreases spike density (9). DNA sequencing showed that the *Zeo1* mutants occur in a *Hordeum vulgare* APELATA2 (*AP2*)-like transcription factor, *HvAP2*. The dense spike and cleistogamous (small lodicules) phenotypes are a consequence of a perturbed interaction between microRNA 172 (*Hv-miR172*) and its corresponding binding site on the mRNA from the *HvAP2* gene, which acts early in spike development to regulate turnover of *HvAP2* mRNA (5, 7).

Origin of mutant:

A neutron induced mutant in Bonus (NGB 14657, PI 189763) (2).

Mutational events:

Ert-r.52 (NGB 112651, GSHO 492), *-r.67* (NGB 112666) in Bonus (NGB 14657, PI 189763), *-r.329* (NGB 112844) in Foma (NGB 14659, CIho 11333) (8); *Ert-r.453* (NGB 112968) in Foma (6). *Ert-r.52* and *Ert-r.67* have the same basepair change in the *HvAP2* locus and are identified as *Zeo1.b* mutants (5).

Mutant used for description and seed stocks:

Ert-r.52 (GSHO 492, NGB 112651) in Bonus; *Ert-r.52* in Bowman (PI 483237)*8 (GSHO 2123, BW322, NGB 22117).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Hagberg, A., Å. Gustafsson, and L. Ehrenberg. 1958. Sparsely contra densely ionizing

radiations and the origin of erectoid mutants in barley. *Hereditas* 44:523-530.

3. Hagberg, A., G. Persson, and A. Wiberg. 1963. Induced mutations in the improvement of self-pollinated crops. p. 105-124. *In* E. Åkerberg and A. Hagberg (eds.) *Recent Plant Breeding Research*. Svalöf 1946-1961. Almqvist & Wiksell, Stockholm.

4. Hayes, H.K., and H.V. Harlan. 1920. The inheritance of the length of internode in the rachis of the barley spike. U.S. Dept. Agr., Bull. 869. 26 pp.

5. Houston, K., S.M. McKim, J. Comadran, N. Bonar, I. Druka, N. Uzrek, E. Cirillo, J. Guzy-Wrobelska, N.C. Collins, C. Halpin, M. Hansson, C. Dockter, A. Druka, and R. Waugh. 2013. Variation in the interaction between alleles of *HvAPETALA2* and

microRNA172 determines the density of grains on the barley inflorescence. *Proc. Natl. Acad. Sci. USA* 110:16675-16680.

6. Lundqvist, U. (Unpublished).

7. Nair, S.K., N. Wang, Y. Turuspekova, M. Pourkheirandish, S. Sinsuwongwat, G. Chen, M. Sameri, A. Tagiri, I. Honda, Y. Watanabe, H. Kanamori, T. Wicker, N. Stein, Y. Nagamura, T. Matsumoto, and T. Komatsuda. 2010. Cleistogamous flowering in barley

arises from the suppression of microRNA-guided *HvAP2* mRNA cleavage. *Proc. Natl. Acad. Sci. USA* 107:490-495.

8. Persson, G., and A. Hagberg. 1969. Induced variation in a quantitative character in barley. Morphology and cytogenetics of *erectoides* mutants. *Hereditas* 61:115-178.

9. Stoy, V., and A. Hagberg. 1967. Effects of growth regulators on ear density mutants in barley. *Hereditas* 58:359-384.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:280.

Revised:

U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:135-136.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:123-124.

BGS 352, Glossy sheath 2, *gsh2*

Stock number: BGS 352
Locus name: Glossy sheath 2
Locus symbol: *gsh2*

Previous nomenclature and gene symbolization:

Glossy sheath 2 = *gs2* (21, 23).

Eceriferum-b = *cer-b* (9, 23).

Inheritance:

Monofactorial recessive (9, 20).

Located in chromosome 3HL (2, 4, 22); *gsh2.f* is about 5.1 cM proximal from the *als1* (absent lower laterals 1) locus (5, 6); *gsh2.f* is over 32.5 cM distal from the *uzu1* (*uzu* 1) locus (2, 5, 6); *gsh2.f* is associated with SNP markers 2_1161 to 1_0584 (positions 169.94 to 178.12 cM) in 3H bin 11 of the Bowman backcross-derived line BW405; *cer-b.2* is associated with SNP markers 22_1513 to 1_1172 (positions 190.97 cM) in 3H bins 11 and 12 of the Bowman backcross-derived line BW107 (1); *gsh2* variants mapped at position 96.6 cM in 3H (19) in the POPSEQ map (17); the *cer-b.2* mutant mapped to a 1.3-cM interval flanked by SNP markers MLOC_10972 and MLOC_69561 (24), in 3H bin 11.

Description:

Surface wax coating appears absent on the spike, leaf sheath, and stem (wax code - - ++) (9, 20). The nodes of mutant plants are covered with surface wax (7). The Bowman backcross-derived lines for the *gsh2* mutants, BW107 with *cer-b.2* and BW405 with *gsh2.f*, had agronomic traits similar to those of Bowman, but grain yields were slightly lower (3). The *cer-b.2* mutant caused major changes in the composition of surface waxes on the leaf sheath and deficiency in the cuticular wax component 14,16-hentriacontanedione (24).

Origin of mutant:

Spontaneous occurrence in Vantage (Clho 7324) (18); an X-ray induced mutant in Maja Abed (NGB 8815, PI 184884) (9).

Mutational events:

gsh2.d in Vantage (Clho 7324), *gsh2.e* in Ymer (NGB 14860, Clho 7275), *gsh2.f* (GSHO 736), *gsh2.g* in Atlas (Clho 4118), *gsh2.h* in Klargin (18); *cer-b.2* (NGB 110886, GSHO 434; GSHO 1081) in Maja Abed (NGB 8815, PI 184884) (7, 8, 9, 22); *cer-b.4* (NGB 110888), *-b.64* (NGB 110948), *-b.66* (NGB 110950), *-b.109* (NGB 110994 in Bonus (NGB 14657, PI 189763) (9, 10); *cer-b.79* (NGB 110963), *-b.94* (NGB 110978), *-b.96* (NGB 110980) in Bonus, *-b.209* (NGB 111096), *-b.222* (NGB 111109), *-b.319* (NGB 111206), *-b.336* (NGB 111223), *-b.337* (NGB 111224), *-b.377* (NGB 111264), *-b.418* (NGB 111306), *-b.424* (NGB 111312), *-b.427* (NGB 111315) in Foma (NGB 14659, Clho 11333) (10, 16); *cer-b.169* (NGB 111055), *-b.193* (NGB 111079) in Bonus, *-b.412* (NGB 111300), *-b.531* (NGB 111419), *-b.549* (NGB 111437) in Foma, *-b.1037* (NGB 111925), *-b.1059* (NGB 111947) in Carlsberg II (NGB 5085, Clho 10114) (10); *cer-b.751* (NGB 111639), *-b.769* (NGB 111657), *-b.791* (NGB 111679), *-b.799* (NGB 111687), *-b.800* (NGB 111688) in Bonus, *-b.1129* (NGB 112017) in Kristina (NGB 1500, NGB 14661) (10); *cer-b.910* (NGB 111798) in Bonus (12); *cer-b.932* (NGB 111820) in Bonus (13); *cer-b.147* (NGB 111033), *-b.182* (NGB 111068), *-b.623* (NGB 111511) in Bonus, *-b.1099* (NGB 111987), *-b.1267* (NGB 112155), *-b.1269* (NGB 112157) in Kristina (14);

cer-b.1699 (NGB 119343) in Bonus (15); *gsh2.ab* (OUM019), *gsh2.ah* (OUM025) in Akashinriki (OUJ659, PI 467400) (4, 6); *gsh2.bb* in the cross NDB112/Fr926-77 (3); *gsh2.bd* in Ms13551 and *gsh2.be* in PI 282610 both in *Hordeum vulgare* subsp. *spontaneum* (1, 3); *gsh2* variants in *H. vulgare* ssp. *spontaneum* accessions WBDC032, WBDC035, and WBDC348 (19).

Mutant used for description and seed stocks:

gsh2.f (GSHO 736) in Atlas; *cer-b.2* (NGB 110886, GSHO 434; GSHO 1081) in Maja Abed (NGB 8815, PI 184884); *gsh2.f* in Bowman (PI 483237)*7 (GSHO 1988); *gsh2.f* in Bowman*8 (BW405, NGB 20678); *cer-b.2* in Bowman*6 (GSHO 1989); *cer-b.2* in Bowman*7 (BW107, NGB 20513); *gsh2.bd* from Ms13551 in Bowman*4 (BW402, NGB 22150); *gsh2.be* from PI 282610 in Bowman*7 (BW403, NGB 22151).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Eslick, R.F., and W.L. McProud. 1974. Positioning of genetic male sterile 5 (*msg5*) on chromosome 3. *Barley Genet. Newsl.* 4:16-23.
3. Franckowiak, J.D. (Unpublished).
4. Konishi, T. 1970. Genetic analyses of two kinds of EMS-induced mutants of barley. *Barley Newsl.* 13:57-58.
5. Konishi, T. 1973. Genetic analyses of EMS-induced mutants in barley. *Barley Genet. Newsl.* 3:28-31.
6. Konishi, T., J. Hayashi, I. Moriya, and R. Takahashi. 1984. Inheritance and linkage studies in barley. VII. Location of six new mutant genes on chromosome 3. *Ber. Ohara Inst. landw. Biol., Okayama Univ.* 18:251-264.
7. Lundqvist, U. (Unpublished).
8. Lundqvist, U. 1992. Coordinator's report: *Eceriferum* genes. *Barley Genet. Newsl.* 21:99-101.
9. Lundqvist, U., and D. von Wettstein. 1962. Induction of *eceriferum* mutants in barley by ionizing radiations and chemical mutagens. *Hereditas* 48:342-362.
10. Lundqvist, U., and D. von Wettstein. 1971. Stock list for the *eceriferum* mutants. *Barley Genet. Newsl.* 1:97-102.
11. Lundqvist, U., and D. von Wettstein. 1973. Stock list for the *eceriferum* mutants II. *Barley Genet. Newsl.* 3:110-112.
12. Lundqvist, U., and D. von Wettstein. 1975. Stock list for the *eceriferum* mutants III. *Barley Genet. Newsl.* 5:88-91.
13. Lundqvist, U., and D. von Wettstein. 1977. Stock list for the *eceriferum* mutants IV. *Barley Genet. Newsl.* 7:92-96.
14. Lundqvist, U., and D. von Wettstein. 1982. Stock list for the *eceriferum* mutants VI. *Barley Genet. Newsl.* 12:169-172.
15. Lundqvist, U., and D. von Wettstein. 1985. Stock list for the *eceriferum* mutants VII. *Barley Genet. Newsl.* 15:89-93.
16. Lundqvist, U., P. von Wettstein-Knowles, and D. von Wettstein. 1968. Induction of *eceriferum* mutants in barley by ionizing radiations and chemical mutagens. II. *Hereditas* 59:473-504.
17. Mascher, M., G.J. Muehlbauer, D.S. Rokhsar, J. Chapman, J. Schmutz, K. Barry, M. Munoz-Amatria, T.J. Close, R.P. Wise, A.H. Schulman, A. Himmelbach, K.F.X. Mayer,

- U. Scholz, J.A. Poland, N. Stein, and R. Waugh. 2013. Anchoring and ordering NGS contig assemblies by population sequencing (POPSEQ). *Plant J.* 76:718-727.
18. McProud, W.L., and R.F. Eslick. 1971. Allelism within a series of glossy sheath mutants. *Barley Genet. Newsl.* 1:31-34.
19. Nice, L.M., B.J. Steffenson, G.L. Brown-Guedira, E.D. Akhunov, C. Liu, T.J.Y. Kono, P.L. Morrell, T.K. Blake, R.D. Horsley, K.P. Smith, and G.J. Muehlbauer. 2016. Development and genetic characterization of an advanced backcross-nested association mapping (AB-NAM) population of wild x cultivated barley. *Genetics* 203:1453-1467.
20. Rasmusson, D.C., and J.W. Lambert. 1965. Inheritance of the glossy-sheath character in barley, *Hordeum vulgare* L. *Crop Sci.* 5:251-253.
21. Robertson, D.W., G.A. Wiebe, R.G. Shands, and A. Hagberg. 1965. A summary of linkage studies in cultivated barley, *Hordeum* species: Supplement III, 1954-1963. *Crop Sci.* 5:33-43.
22. Singh, R.J., and T. Tsuchiya. 1974. Further information on telotrisomic analysis in barley. *Barley Genet. Newsl.* 4:66-69.
23. Wettstein-Knowles, P. von. 1991. New alleles of *Cer-yy* and *cer-b*. *Barley Genet. Newsl.* 20:66-68.
24. Zhou, Q., C. Li, K. Mishina, J. Zhao, J. Zhang, R. Duan, X. Ma, A. Wang, Q. Meng, T. Komatsuda, and G. Che. 2017. Characterization and genetic mapping of the β -diketone deficient *eceriferum-b* barley mutant. *Theor. Appl. Genet.* 130:1169-1178.

Prepared:

- W.L. McProud and R.F. Eslick. 1971. *Barley Genet. Newsl.* 1:169.
- U. Lundqvist. 1975. *Barley Genet. Newsl.* 5:116 as BGS 398, *Eceriferum-b*, *cer-b*.

Revised:

- U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:294-295.
- U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:141-142.
- U. Lundqvist and J.D. Franckowiak. 2014. *Barley Genet. Newsl.* 44:124-125.
- U. Lundqvist and J.D. Franckowiak. 2016. *Barley Genet. Newsl.* 46:98-100.
- U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:125-127.

BGS 355, Glossy sheath 5, *gsh5*

Stock number: BGS 355
Locus name: Glossy sheath 5
Locus symbol: *gsh5*

Previous nomenclature and gene symbolization:

Glossy sheath 5 = *gs5* (15).
Eceriferum-s = *cer-s* (5, 13).
Glossy sheath 8 = *gs8* (17).

Inheritance:

Monofactorial recessive (5).
Located in chromosome 2HL (15, 16, 17, 19); *gsh5.m* is located in the centromeric region about 2.5 cM proximal from the *eog1* (elongated outer glume 1) locus (15, 17); *gsh5.m* is about 32.0 cM proximal from the *vrs1* (six-rowed spike 1) locus (15, 17); *gsh5.m* is associated with SNP markers 1_0851 to 2_1437 (positions 81.43 to 113.28 cM) in 2H bins 06 to 08 of the Bowman backcross-derived line BW408 (2); *cer-s.31* is associated with SNP markers 1_0796 to 2_0528 (positions 95.53 to 118.78 cM) in 2H bins 07 to 08 of the Bowman backcross-derived line BW122 (2), in 2H bin 07.

Description:

Mutants have a reduced surface wax covering on the spikes and stems, and a bright glossy green color on all leaf sheaths (15) (wax code + - ++) (5). The nodes appear to have an excessive wax covering (4). The Bowman backcross-derived line with the *cer-s.31* allele, BW122, headed 2 to 4 days later than BW408 with the *gsh5.m* allele and Bowman (4). BW122 also had 3 to 6 more kernels per spike. Kernels of BW122 and BW408 were about 10% lighter than those of Bowman. The grain yields of BW408 were often slightly lower than those of Bowman or BW408 (4). The differences in maturity and number of kernels per spike between BW122 and BW408 were attributed to close linkage between alleles at the *gsh5* locus and those at the *Eam6* [Early maturity 6, Earliness per se 2S (*eps2S*), Praematurum-c (*mat-c*), or *Hordeum vulgare* *CENTRORADIALIS* (*HvCEN*)] locus (1, 4).

Origin of mutant:

An X-ray induced mutant in Bonus (NGB 14657, PI 189763) (5); a radiation induced mutant in Jotun (PI 467357) (15); a spontaneous mutant in Okaiku 3 (OUL011) (18).

Mutational events:

cer-s.24 (NGB 110908) in Bonus (NGB 14657, PI 189763) (5); *cer-s.31* (NGB 110915, GSHO 440) in Bonus (3, 5); *cer-s.41* (NGB 110925) in Bonus (5); *cer-s.287* (NGB 111174), -s.303 (NGB 111190), -s.372 (NGB 111259), -s.444 (NGB 111332) in Foma (NGB 14659, Clho 11333) (13); *cer-s.1047* (NGB 111935) in Carlsberg II (NGB 5085, Clho 10114) (6); *cer-s.137* (NGB 111022) in Bonus, -s.591 (NGB 111479) in Foma, -s.803 (NGB 111691) in Bonus, -s.1132 (NGB 112020) in Kristina (NGB 14661, NGB 1500) (7); *cer-s.914* (NGB 111802) in Bonus (8); *cer-s.622* (NGB 111510) in Bonus, -s.1054 (NGB 111942) in Carlsberg II, -s.1153 (NGB 117264) in Kristina (9); *cer-s.1323* (NGB 112211) in Bonus (10); *cer-s.1134* (NGB 112022) in Kristina (11); *cer-s.1754* (NGB 112568) in Bonus (12); *gsh5.m* (GSHO 739) in Jotun (PI 467357) (15); *gsh5.n* (GSHO 832) in Okaiku 3 (OUL011) (14, 18).

Mutant used for description and seed stocks:

gsh5.m (GSHO 739) in Jotun; *cer-s.31* (NGB 110915, GSHO 440) in Bonus; *cer-s.31* in

Bowman (PI 483237)*5 (GSHO 1888); *cer-s.31* in Bowman*8 (BW122, NGB 20528); *gsh5.m* in Bowman*8 (GSHO 1889, BW408, NGB 20641).

References:

1. Comadran, J., B. Kilian, J. Russell, L. Ramsay, N. Stein, M. Ganai, P. Shaw, M. Bayer, W. Thomas, D. Marshall, P. Hedley, A. Tondelli, N. Pecchioni, E. Francia, V. Korzun, A. Walther, and R. Waugh. 2012. Natural variation in a homolog of *Antirrhinum CENTRORADIALIS* contributed to spring growth habit and environmental adaptation in cultivated barley. *Nature Genet.* 44:1388-1392.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
3. Eslick, R.F. 1974. Allele tests of *gs* and *cer* mutants. *Barley Genet. Newsl.* 4:9-11.
4. Franckowiak, J.D. (Unpublished).
5. Lundqvist, U., and D. von Wettstein. 1962. Induction of eceriferum mutants in barley by ionizing radiations and chemical mutagens. *Hereditas* 48:342-362.
6. Lundqvist, U., and D. von Wettstein. 1971. Stock list for the eceriferum mutants. *Barley Genet. Newsl.* 1:97-102.
7. Lundqvist, U., and D. von Wettstein. 1973. Stock list for the eceriferum mutants II. *Barley Genet. Newsl.* 3:110-112.
8. Lundqvist, U., and D. von Wettstein. 1975. Stock list for the eceriferum mutants III. *Barley Genet. Newsl.* 5:88-91.
9. Lundqvist, U., and D. von Wettstein. 1977. Stock list for the eceriferum mutants IV. *Barley Genet. Newsl.* 7:92-96.
10. Lundqvist, U., and D. von Wettstein. 1979. Stock list for the eceriferum mutants V. *Barley Genet. Newsl.* 9:135-137.
11. Lundqvist, U., and D. von Wettstein. 1982. Stock list for the eceriferum mutants VI. *Barley Genet. Newsl.* 12:169-172.
12. Lundqvist, U., and D. von Wettstein. 1985. Stock list for the eceriferum mutants VII. *Barley Genet. Newsl.* 15:89-93.
13. Lundqvist, U., P. von Wettstein-Knowles, and D. von Wettstein. 1968. Induction of eceriferum mutants in barley by ionizing radiations and chemical mutagens. II. *Hereditas* 59:473-504.
14. McProud, W.L., and R.F. Eslick. 1971. Allelism within a series of glossy sheath mutants. *Barley Genet. Newsl.* 1:31-34.
15. Rasmusson, D.C., and J.W. Lambert. 1965. Inheritance of the glossy-sheath character in barley, *Hordeum vulgare* L. *Crop Sci.* 5:251-253.
16. Singh, R.J., and T. Tsuchiya. 1974. Further information on telotrisomic analysis in barley. *Barley Genet. Newsl.* 4:66-69.
17. Takahashi, R., J. Hayashi, T. Konishi, and I. Moriya. 1972. Inheritance and linkage studies in barley. V. Locating seven new mutant genes. *Ber. Ohara Inst. landw. Biol., Okayama Univ.* 15:147-168.
18. Takahashi, R., J. Hayashi, and I. Moriya. 1971. Linkage studies in barley. *Barley Genet. Newsl.* 1:51-58.
19. Tsuchiya, T. 1972. Cytogenetics of telotrisomics in barley. *Barley Genet. Newsl.* 2:93-98.

Prepared:

W.L. McProud and R.F. Eslick. 1971. *Barley Genet. Newsl.* 1:171-172.

Revised:

Barley Genetics Newsletter (2017): 46:50-187.

- J.D. Franckowiak and U. Lundqvist. 1997. *Barley Genet. Newsl.* 26:300-301.
U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:149-150.
U. Lundqvist and J.D. Franckowiak. 2014. *Barley Genet. Newsl.* 44:126-127.
U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:128-130.

BGS 402, Eceriferum-g, *cer-g*

Stock number: BGS 402
Locus name: Eceriferum-g
Locus symbol: *cer-g*

Previous nomenclature and gene symbolization:

None.

Inheritance:

Monofactorial recessive (4).

Located in chromosome 2HL (2, 14); *cer-g.10* is about 15.9 cM distal from the *vrs1* (six-rowed spike 1) locus (14, 15); *cer-g.10* is associated with SNP markers 1_0317 to 2_0374 (positions 98.35 to 104.81 cM) in 2H bin 08 and SNP markers 1_0619 to 1_0533 (positions 133.59 to 141.56 cM) in 2H bin 09 of the Bowman backcross-derived line BW111 (1), in 2H bin 08.

Description:

Mutants have reduced surface wax coating on spike, leaf sheath, and stem (wax code + + +) with the wax coating of the leaf sheath and stem wax present in broad horizontal bands (4). Double and triple stomatal complexes are produced during stomatal development (4, 16). All *cer-g* mutants have globe-shaped or globosum kernels (4). Plants of the Bowman backcross-derived line for *cer-g.10*, BW111, headed 1 to 2 days later than Bowman and were about 10% shorter. Kernels of BW111 were shorter (8.2 vs. 9.5 mm), narrower (3.5 vs. 3.8 mm) and weighed 15 to 20% less. Grain yields of BW111 were 2/3 to 3/4 of those for Bowman (3).

Origin of mutant:

An X-ray induced mutant in Bonus (NGB 14657, PI 189763) (4).

Mutational events:

cer-g.10 (NGB 110894, GSHO 428), *-g.11* (NGB 110895), *-g.23* (NGB 110907), *-g.47* (NGB 110931), *-g.55* (NGB 110939) in Bonus (NGB 14657, PI 189763) (4); *cer-g.106* (NGB 110991) in Bonus, *-g.202* (NGB 111089), *-g.208* (NGB 111095), *-g.210* (NGB 111097), *-g.494* (NGB 111382) in Foma (NGB 14659, Clho 11333) (13); *cer-g.166* (NGB 111052) in Bonus, *-g.553* (NGB 111441), *-g.568* (NGB 111456), *-g.700* (NGB 111588), *-g.709* (NGB 111597), *-g.710* (NGB 111598) in Foma, *-g.1026* (NGB 111914) in Carlsberg II (NGB 5085, Clho 10114) (5); *cer-g.746* (NGB 111634), *-g.818* (NGB 111706), *-g.875* (NGB 111763) in Bonus, *-g.1033* (NGB 111921), *-g.1071* (NGB 111959) in Carlsberg II, *-g.1097* (NGB 111985), *-g.1116* (NGB 112004) in Kristina (NGB 14661, NGB 1500) (6); *cer-g.893* (NGB 111781), *-g.976* (NGB 111864) in Bonus, *-g.1172* (NGB 112060) in Kristina (7); *cer-g.1161* (NGB 117265), *-g.1169* (NGB 112057) in Kristina (8); *cer-g.1038* (NGB 111926) in Carlsberg II, *-g.1289* (NGB 112177) in Kristina, *-g.1332* (NGB 112220) in Bonus (9); *cer-g.1410* (NGB 112298), *-g.1451* (NGB 112339), *-g.1454* (NGB 112342) in Bonus (10); *cer-g.1495* (NGB 112383) in Bonus, *-g.1510* (NGB 112398) in Nordal (NGB 13680, NGB 4704), *-g.1736* (NGB 112549), *-g.1740* (NGB 112553), *-g.1741* (NGB 112554) in Bonus (11); *cer-g.1573* (NGB 112461) in Nordal (12).

Mutant used for description and seed stocks:

cer-g.10 (NGB 110894, GSHO 428) in Bonus; *cer-g.10* in Bowman (PI 483237)*6 (GSHO 1906); *cer-g.10* in Bowman*7 (BW111, NGB 20517).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Fester, T., and B. Søgaard. 1969. The localization of eceriferum loci in barley. *Hereditas* 61:327-337.
3. Franckowiak, J.D. (Unpublished).
4. Lundqvist, U., and D. von Wettstein. 1962. Induction of eceriferum mutants in barley by ionizing radiations and chemical mutagens. *Hereditas* 48:342-362.
5. Lundqvist, U., and D. von Wettstein. 1971. Stock list for the eceriferum mutants. *Barley Genet. Newsl.* 1:97-102.
6. Lundqvist, U., and D. von Wettstein. 1973. Stock list for the eceriferum mutants II. *Barley Genet. Newsl.* 3:110-112.
7. Lundqvist, U., and D. von Wettstein. 1975. Stock list for the eceriferum mutants III. *Barley Genet. Newsl.* 5:88-91.
8. Lundqvist, U., and D. von Wettstein. 1977. Stock list for the eceriferum mutants IV. *Barley Genet. Newsl.* 7:92-96.
9. Lundqvist, U., and D. von Wettstein. 1979. Stock list for the eceriferum mutants V. *Barley Genet. Newsl.* 9:135-137.
10. Lundqvist, U., and D. von Wettstein. 1982. Stock list for the eceriferum mutants VI. *Barley Genet. Newsl.* 12:169-172.
11. Lundqvist, U., and D. von Wettstein. 1985. Stock list for the eceriferum mutants VII. *Barley Genet. Newsl.* 15:89-93.
12. Lundqvist, U., and D. von Wettstein. 1988. Stock list for the eceriferum mutants VIII. *Barley Genet. Newsl.* 18:88-91.
13. Lundqvist, U., P. von Wettstein-Knowles, and D. von Wettstein. 1968. Induction of eceriferum mutants in barley by ionizing radiations and chemical mutagens. II. *Hereditas* 59:473-504.
14. Søgaard, B. 1971. Linkage studies on eceriferum mutants in barley. *Barley Genet. Newsl.* 1:41-47.
15. Søgaard, B. 1973. Continued linkage studies on eceriferum mutants in barley. *Barley Genet. Newsl.* 3:57-61.
16. Zeiger, E., and G.L. Stebbins. 1972. Developmental genetics in barley: a mutant for stomatal development. *Am. J. Bot.* 59:143-148.

Prepared:

U. Lundqvist. 1975. *Barley Genet. Newsl.* 5:120.

Revised:

J.D. Franckowiak and U. Lundqvist. 1997. *Barley Genet. Newsl.* 26:349-350.
U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:155-156.
U. Lundqvist and J.D. Franckowiak. 2014. *Barley Genet. Newsl.* 44:140-141.
U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:131-132.

BGS 439, Eceriferum-zv, *cer-zv*

Stock number: BGS 439
Locus name: Eceriferum-zv
Locus symbol: *cer-zv*

Previous nomenclature and symbolization:

None.

Inheritance:

Monofactorial recessive (6).

Located in chromosome 4HL (1, 3, 4); *cer-zv.268* is associated with SNP markers 2_1122 to 1_0568 (positions 47.80 to 78.18 cM) in 4H bins 05 to 06 of the Bowman backcross-derived line BW177 (1); *cer-ym.753*, a possible allele (4), is associated with SNP markers 2_0289 to 1_1513 (positions 69.62 to 101.44 cM) in 4H bins 05 to 07 of the Bowman backcross-derived line BW144 (1); *cer-zv.268* and *cer-ym.753* are in a 0.8 cM interval between EST markers AK370363 and AK251484 in pericentromeric region of 4H (3, 4).

Description:

Surface wax coating on the spike, the leaf sheath and stem, and the leaf blade appears absent (wax code - - -) (6). Plants are semidwarf, very weak, and head late, and the hull is poorly attached to the kernel. The Bowman backcross-derived line for *cer-zv.268*, BW177, is a semidwarf (2/3 normal height) with a kinked upper peduncle and a strong wax coating on the nodes (2). Compared to Bowman, BW177 plants headed 2 to 10 days later in various environments. Plant height, peduncle length, and leaf blade length measurements were 1/2 to 2/3 those for Bowman. Awns were slightly shorter, but rachis internodes were slightly longer. Kernels of BW177 were shorter and thinner and weighed less, 3.5 vs. 5.8 mg, compared to Bowman kernels. Grain yields of BW177 averaged 10% those of Bowman (2). The *cer-ym.753* plants are weak, culms have a kinked peduncle, nodes are wax covered, and kernels are small, globe shaped, and nearly hullless (2, 5). Plants of the Bowman backcross-derived line for *cer-ym.753*, BW144, were phenotypically similar to BW177 plants, but they were more vigorous (2, 3, 4). Detached leaves of the mutant *cer-ym.753* lost 41.8% of their initial weight after 1 hour of dehydration under laboratory conditions, while that of the wild type Bowman plants lost only 7.1%. Stomata function was not affected by the mutation, but the mutant leaves showed increased cuticular permeability to water, suggesting a defective leaf cuticle, which was confirmed by toluidine blue staining. The *cer-ym.753* leaf blades showed a substantial reduction in the amounts of the major cutin monomers and a slight increase in the main wax component, suggesting that the enhanced cuticle permeability was a consequence of cutin deficiency (3, 4). Allelism tests demonstrated that mutants at the *cer-zv* and *cer-ym* loci are not alleles (5).

Origin of mutant:

An ethylene imine induced mutant in Foma (NGB 14659, CIho 11333) (5).

Mutational events:

cer-zv.268 (NGB 111155, GSHO 1529), *-zv.342* (NGB 111229), *-zv.576* (NGB 111464) in Foma (NGB 14659, CIho 11333) (6); *cer-zv.1256* (NGB 112144) in Kristina (NGB 14661, NGB 1500) (8); and based on location possibly *cer-ym.130*, (NGB 111015, GSHO 1546), *-ym.753* (NGB 111641) in Bonus (NGB 14657, PI 189763) (7).

Mutant for description and seed stocks:

cer-zv.268 (NGB 111155, GSHO 1529) in Foma; *cer-zv.268* in Bowman (PI 483237)*5 (GSHO 2207); *cer-zv.268* in Bowman *7 (BW177, NGB 22009); *cer-ym.753* (NGB 111641, GSHO 1546) in Bonus; *cer-ym.753* in Bowman (PI 483237)*5 (GSHO 2217); *cer-ym.753* in Bowman*7 (BW144, NGB 20550).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendrarnin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Li, C., C. Liu, X. Ma, A. Wang, R. Duan, C. Nawrath, T. Komatsuda, and G. Chen. 2015. Characterization and genetic mapping of *eceriferum-ym* (*cer-ym*), a cutin deficient barley mutant with impaired leaf water retention capacity. *Breed. Sci.* 65:327-332.
4. Li, C., A. Wang, X. Ma, M. Pourkheirandish, S. Sakuma, N. Wang, S. Ning, E. Nevo, C. Nawrath, T. Komatsuda, and G. Chen. 2013. An *eceriferum* locus, *cer-zv*, is associated with a defect in cutin responsible for water retention in barley (*Hordeum vulgare*) leaves. *Theor. Appl. Genet.* 126:637-646.
5. Lundqvist, U. (Unpublished).
6. Lundqvist, U., and D. von Wettstein. 1973. Stock list for the *eceriferum* mutants II. *Barley Genet. Newsl.* 3:110-112.
7. Lundqvist, U., and D. von Wettstein. 1977. Stock list for the *eceriferum* mutants IV. *Barley Genet. Newsl.* 7:92-96.
8. Lundqvist, U., and D. von Wettstein. 1979. Stock list for the *eceriferum* mutants V. *Barley Genet. Newsl.* 9:135-137.

Prepared:

U. Lundqvist. 1975. *Barley Genet. Newsl.* 5:157.

Revised:

- U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:391.
U. Lundqvist and J.D. Franckowiak. 2016. *Barley Genet. Newsl.* 46:109-110.
U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:133-134.

Stock number: BGS 473
Locus name: Compositum 1
Locus symbol: *com1*

Previous nomenclature and gene symbolization:

Laxatum-270 = *lax*-270 (5, 7, 9).
Short-haired, inflorescent rachilla = *sⁱ* (7).
Branching, inflorescent rachilla = *bir1a* (8).
Compositum = *com1a* (9, 10, 11).

Inheritance:

Monofactorial recessive (7, 8, 9).
Located in chromosome 5HL (11, 12); *com1.a* is near the *srh1* (short rachilla hair 1) locus (12); *com1.a* is between the *ert-n* (erectoides-n) and the *lax-a* (laxatum-a) loci (11), but widely different estimates of their distance from the *srh1* (short rachilla hair 1) locus are reported (6, 11); *com1.a* is associated with SNP markers 1_0974 to 1_0641 (positions 41.99 to 108.33 cM) in 5H of the Bowman backcross-derived line BW189 (1); *com1.i* is associated with SNP markers 3_0745 to 1_0641 (positions 99.51 to 108.33 cM) in 5HL of the Bowman backcross-derived line BW605 (1), likely in 5H bin 06.

Description:

The spike is branched from the rachis of the bottom 1 to 10 spikelets. Awns, which vary from normal to thread-like and protruding branches, are bent in various directions because of "packing" problems in the boot (7). More spike branches develop from rachis nodes under favorable environmental conditions. Plants of Bowman backcross-derived lines BW189, BW190, and BW605 had slightly lower kernel weight and test weights compared to Bowman. The average kernel width was slightly reduced (3).

Origin of mutant:

An ethyl methanesulfonate and neutron induced mutant in Foma (NGB 14659, Clho 11333) (2, 7).

Mutational events:

com1.a (*lax*.270, NGB 116495, GSHO 1702), *com1.b* (*lax*.271, NGB 116496), *com1.c* (*lax*.272, NGB 116497), *com1.d* (*lax*.206, NGB 116433), *com1.e* (*lax*.213, NGB 116442) in Foma (NGB 14659, Clho 11333) (9); *com1.i* (*lax*.296, NGB 116520, GSHO 1699) in Foma (3, 9); *com1.l* (*mul.c*, Mut. 2227, GSHO 2412) in Donaria (PI 161974) (3, 13).

Mutant used for description and seed stocks:

com1.a (NGB 116495, GSHO 1702) in Foma; *com1.a* in Bowman (PI 483237)*6 (GSHO 2098); *com1.a* in Bowman*7 (BW189, NGB 22021); *com1.i* (NGB 116520, GSHO 1699) in Foma; *com1.i* in Bowman*3 (GSHO 2232); *com1.i* in Bowman*5 (BW190, NGB 22022); *com1.l* (*mul.c*) in Donaria (GSHO 2412); *com1.l* in Bowman*6 (GSHO 2312); *com1.l* in Bowman*7 (BW605, NGB 22171).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
3. Franckowiak, J.D. (Unpublished).

4. Franckowiak, J.D., B.P. Forster, U. Lundqvist, J. Lyon, I. Pitkethly, and W.T.B. Thomas. 2010. Developmental mutants as a guide to the barley phytomer. pp. 46-60. *In*: S. Ceccarelli and S. Grando (eds.), Proc. 10th International Barley Genetics Symposium, 5-10 April 2008, Alexandria Egypt. ICARDA, PO Box 5466, Aleppo, Syria.
5. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
6. Jensen, J. 1981. Construction of a barley chromosome 7 linkage map. p. 927-939. *In* M.J.C. Asher, R.P. Ellis, A.M. Hayter, and R.N.H. Whitehouse (eds.) *Barley Genetics IV*. Proc. Fourth Int. Barley Genet. Symp., Edinburgh. Edinburgh Univ. Press, Edinburgh.
7. Larsson, H.E.B. 1981. Branching spike mutants from two loci in two-row barley. *Barley Genet. Newsl.* 11:25-28.
8. Larsson, H.E.B. 1983. New data about a gene for branching inflorescent rachilla located on barley chromosome 7. *Barley Genet. Newsl.* 13:82-83.
9. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
10. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
11. Larsson, H.E.B. 1985. Linkage studies with genetic markers and some *laxatum* barley mutants. *Hereditas* 103:269-279.
12. Persson, G. 1969. An attempt to find suitable genetic markers for dense ear loci in barley I. *Hereditas* 62:25-96.
13. Scholz, F., and O. Lehmann. 1958. Die Gaterslebener Mutanten der Saatgerste in Beziehung zur Formenmannigfaltigkeit der Art *Hordeum vulgare* L.s.l.l. *Kulturpflanze* 6:123-166.

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:56.

Revised:

- U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:420.
U. Lundqvist and J.D. Franckowiak. 2010. *Barley Genet. Newsl.* 40:118-119.
U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:135-136.

Stock number: BGS 475
Locus name: Laxatum-c
Locus symbol: *lax-c*

Previous nomenclature and gene symbolization:

Laxatum-21 = *lax-21* (3, 4, 5).

Inheritance:

Monofactorial recessive (5).

Located in chromosome 6HL (6); *lax-c.21* is linked to a translocation breakpoint in chromosome 6HL (6); *lax-c.21* is over 27.3 cM distal from the *lax-b* (laxatum-b) locus (6); *lax-c.21* is associated with SNP markers 1_0015 to 1_1534 (positions 160.38 to 180.69 cM) in 6H bin 11 of the Bowman backcross-derived line BW460 (1); *lax-c.231* (*sld.l*) is associated with SNP markers 2_0783 to 1_1534 (positions 146.43 to 180.69 cM) in 6H bins 09 to 11 of the Bowman backcross-derived line BW858 (1).

Description:

The grain is thin and angular and caryopses are exposed between the lemma and palea. The awn has a very wide base. Rachis internodes are about 7% longer than normal. In the original Bonus stock, the tip of the spike is frequently sterile, and the tiller number is reduced (4). Plants of the Bowman backcross-derived line for *lax-c.21*, BW460, were 3/4 the height of Bowman. Rachis internodes of BW460 were slightly longer in some trials. The awns and peduncles of BW460 were slightly shorter than those of Bowman. Kernels were about 10% shorter and thinner and their weights were about 20% lighter, 4.8 mg vs. 5.7 mg, than those of Bowman. Grain yields were about 3/4 of those for Bowman (2). Plants of the Bowman backcross-derived line for *lax-c.231* (*sld.l*), BW858, were similar to those of BW460, except they had more pronounced spiral coiling of the neck and occasionally formed small branches on the awn (2).

Origin of mutant:

An X-ray induced mutant in Bonus (NGB 14657, PI 189763) (4, 7).

Mutational events:

lax-c.21 (NGB 116355, GSHO 1777), *-c.23* (NGB 116358) in Bonus (NGB 14657, PI 189763) (4, 5); *lax.231* (*sld.l*, NGB 116459, GSHO 2478) in Foma (NGB 14659, Clho 11333) may be an allele based on phenotype and SNP markers retained in 6HL of its Bowman backcross-derived line, BW858 (NGB 22295) (1, 2)

Mutant used for description and seed stocks:

lax-c.21 (NGB 116355, GSHO 1777) in Bonus; *lax-c.21* in Bowman (PI 483237)*6 (GSHO 2086); *lax-c.21* in Bowman*7 (BW460, NGB 20691); *lax-c.231* (*sld.l*) from Foma in Bowman*3 (GSHO 2359); *lax-c.231* in Bowman*6 (BW858, NGB 22295).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.

5 Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.

6. Larsson, H.E.B. 1985. Linkage studies with genetic markers and some *laxatum* barley mutants. *Hereditas* 103:269-279.

7. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:58.

Revised:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:423.

U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:174.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:137-138.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 476, Laxatum-d, *lax-d*

Stock number: BGS 476
Locus name: Laxatum-d
Locus symbol: *lax-d*

Previous nomenclature and gene symbolization:

Laxatum-83 = *lax-83* (3, 4).

Inheritance:

Monofactorial incomplete dominant (5).

Location is unknown.

Description:

Rachis internodes of *lax-d.83* were about 6% longer than those of Bonus (4). Plants were taller and relatively vigorous; and the kernels were comparatively long (4).

Origin of mutant:

An X-ray induced mutant in Bonus (NGB 14657, PI 189763) (4, 6).

Mutational events:

lax-d.2 (NGB 116336), *-d.27* (NGB 116362), *-d.83* (NGB 116415) in Bonus (NGB 14657, PI 189763) (1, 2, 5, 6, 7).

Mutant used for description and seed stocks:

lax-d.2 (NGB 116336) and *lax-d.83* (NGB 116415) in Bonus.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:59.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:139.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 477, Laxatum-f, *lax-f*

Stock number: BGS 477
Locus name: Laxatum-f
Locus symbol: *lax-f*

Previous nomenclature and gene symbolization:

Laxatum-61 = *lax-61* (3, 4).

Inheritance:

Monofactorial dominant (5).

Location is unknown.

Description:

Rachis internodes of *lax-f.61* were about 14% longer than those of Bonus, and the mutants had lower kernel weights (4).

Origin of mutant:

An ethylene imine induced mutant in Bonus (NGB 14657, PI 189763) (4, 6).

Mutational events:

lax-f.7 (NGB 116341), *-f.61* (NGB 116395), in Bonus (NGB 14657, PI 189763) (1, 2, 5, 6, 7).

Mutant used for description and seed stocks:

lax-f.7 (NGB 116341) and *lax-f.61* (NGB 116395) in Bonus.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).
7. Stoy, V., and A. Hagberg. 1967. Effects of growth regulators on ear density mutants in barley. *Hereditas* 58:359-384.

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:60.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:140.

BGS 478, Laxatum-g, *lax-g*

Stock number: BGS 478
Locus name: Laxatum-g
Locus symbol: *lax-g*

Previous nomenclature and gene symbolization:

Laxatum-25 = *lax-25* (3, 4).

Inheritance:

Monofactorial incomplete dominant (5).

Location is unknown.

Description:

Rachis internodes of *lax-g.25* were about 8% longer than those of Bonus; and mutant plants exhibited tip sterility and reduced tiller numbers (4).

Origin of mutant:

An X-ray induced mutant in Bonus (NGB 14657, PI 189763) (4, 6).

Mutational events:

lax-g.25 (NGB 116360), *-g.59* (NGB 116393) in Bonus (NGB 14657, PI 189763) (1, 2, 5, 6, 7).

Mutant used for description and seed stocks:

lax-g.25 (NGB 116360) in Bonus.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).
7. Stoy, V., and A. Hagberg. 1967. Effects of growth regulators on ear density mutants in barley. *Hereditas* 58:359-384.

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:61.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:141.

Stock number: BGS 479
Locus name: Laxatum-h
Locus symbol: *lax-h*

Previous nomenclature and gene symbolization:

Laxatum-26 = *lax-26* (5, 6).

Inheritance:

Monofactorial recessive (7).

Located possibly likely in chromosome 3H (1); *lax-h.26* is associated with SNP markers 2_1398 to 1_1172 (positions 13.67 to 190.87 cM) in 3H bins 01 to 12, with SNP markers 1_1061 to 1_1205 (positions 70.15 to 81.35 cM in 6H bins 05 to 06, and with SNP markers 2_1241 to 2_0320 positions 159.51 to 166.29 cM) in 5H bins 09 to 10 of the Bowman backcross-derived line BW463 (1). The original *lax-h.26* mutant was previously associated with translocation testers involving chromosome arms 5HS and 6HL (8, 10).

Description:

Rachis internodes of *lax-h.26* were about 20% longer than those of Bonus (6). Spikes had some sterile spikelets; and kernels were relatively thin and angular (6). Plants of the Bowman backcross-derived line for *lax-h.26*, BW463, had rachis internodes that averaged 20% longer (5.4 vs. 4.5 mm). Compared to Bowman, BW463 had one to two more kernels per spike, and leaf blades were shorter and narrower. Kernels were longer (12.0 vs. 9.3 mm), wider (4.0 vs. 3.8 mm), and weighted about the same, but the test weight was much lower (5). Grain yields of BW463 were less than 1/4 those of Bowman (5).

Origin of mutant:

An X-ray induced mutant in Bonus (NGB 14657, PI 189763) (6, 9).

Mutational events:

lax-h.26 (NGB 116361), *-h.38* (NGB 116373) in Bonus (NGB 14657, PI 189763) (2, 3, 6, 7).

Mutant used for description and seed stocks:

lax-h.26 (NGB 116361) in Bonus; *lax-h.26* in Bowman (PI 483237)*3 (BW463, NGB20693).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
3. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
4. Franckowiak, J.D. (Unpublished).
5. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
6. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
7. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.

8. Larsson, H.E.B. 1985. Linkage studies with genetic markers and some *laxatum* barley mutants. *Hereditas* 103:269-279.
9. Lundqvist, U. (Unpublished).
10. Marthe, F., and G. Künzel. 1994. Localization of translocation breakpoints in somatic metaphase chromosomes of barley. *Theor. Appl. Genet.* 89:240-248.

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:62.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:142-143.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 480, Laxatum-i, *lax-i*

Stock number: BGS 480
Locus name: Laxatum-i
Locus symbol: *lax-i*

Previous nomenclature and gene symbolization:

Laxatum-50 = *lax-50* (3, 4).

Inheritance:

Monofactorial recessive (5).

Location is unknown.

Description:

Rachis internodes of *lax-i.50* were about 18% longer than those of Bonus. The *lax-i.50* plants showed partial sterility, had coiled awns, and had small angular kernels (4).

Origin of mutant:

An ethylene imine induced mutant in Bonus (NGB 14657, PI 189763) (4, 6).

Mutational events:

lax-i.28 (NGB 116363), *-i.50* (NGB 116384) in Bonus (NGB 14657, PI 189763) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-i.50 (NGB 116363) in Bonus.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:63.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:144.

BGS 481, Laxatum-j, *lax-j*

Stock number: BGS 481
Locus name: Laxatum-j
Locus symbol: *lax-j*

Previous nomenclature and gene symbolization:

Laxatum-49 = *lax-49* (6, 7).

Inheritance:

Monofactorial recessive (5).

Located in chromosome 2H (2); *lax-j.49* is associated with SNP markers 1_0399 to 2_0887 (positions 66.78 to 103.73 cM) in 2H bins 05 to 08 and a small segment of 3HL between markers 2_0944 and 1_1172 (positions 185.12 and 190.87 cM) in 3H bin 12 of the Bowman backcross-derived line BW465 (2).

Description:

Rachis internodes of *lax-j.49* were about 24% longer than those of Bonus (7). Plants produced few tillers, awns were slightly twisted, seed set was reduced, and kernels were angular (7). Plants of the Bowman backcross-derived line for *lax-j.49*, BW465, headed eight days later than Bowman and had rachis internodes that were about 30% longer (6.0 vs. 4.6 mm). Compared to Bowman, BW465 plants were slightly shorter, had slightly shorter awns and peduncles, had slightly longer and wider leaf blades, and had about three more kernels per spike (5). Kernels were long (10.3 vs. 9.2 mm), thin (3.4 vs. 3.9 mm), and weighed less (5.1 vs. 6.0 mg). The caryopsis was partially exposed between the lemma and palea. Grain yields of BW465 were about 1/3 those of Bowman (5). A portion of the delayed heading of BW465 could be attributed to the late allele at the Early maturity 6 (*Eam6*) or Praematurum-c (*mat-c*) locus, which was retained in the 2H segment from the donor parent Bonus (1, 2).

Origin of mutant:

An ethylene oxide induced mutant in Bonus (NGB 14657, PI 189763) (7, 9).

Mutational events:

lax-j.29 (NGB 116364), *-j 33* (NGB 116368), *-j.49* (NGB 116383) in Bonus (NGB 14657, PI 189763) (3, 4, 7, 8).

Mutant used for description and seed stocks:

lax-j.49 (NGB 116383) in Bonus; *lax-j.49* in Bowman (PI 483237)*3 (BW465, NGB20695).

References:

1. Comadran, J., B. Kilian, J. Russell, L. Ramsay, N. Stein, M. Ganai, P. Shaw, M. Bayer, W. Thomas, D. Marshall, P. Hedley, A. Tondelli, N. Pecchioni, E. Francia, V. Korzun, A. Walther, and R. Waugh. 2012. Natural variation in a homolog of *Antirrhinum CENTRORADIALIS* contributed to spring growth habit and environmental adaptation in cultivated barley. *Nature Genet.* 44:1388-1392.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
3. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
4. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.

5. Franckowiak, J.D. (Unpublished).
6. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
7. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
8. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
9. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:64.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:145-146.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 482, Laxatum-k, *lax-k*

Stock number: BGS 482
Locus name: Laxatum-k
Locus symbol: *lax-k*

Previous nomenclature and gene symbolization:

Laxatum-84 = *lax-84* (3, 4).

Inheritance:

Monofactorial incomplete dominant (5).

Located in chromosomes 2H or 3H (6); *lax-k.84* is linked to the translocation breakpoint in translocation stock T2-3f (6).

Description:

Rachis internodes of *lax-k.84* were about 4% longer than those of Bonus. The *lax-k.84* plants had narrow, dark green leaf blades and very thin kernels (4).

Origin of mutant:

A gamma-ray induced mutant in Bonus (NGB 14657, PI 189763) (4, 7).

Mutational events:

lax-k.42 (NGB 116376), *-k.44* (NGB 116378), *k.84* (NGB 116416) in Bonus (NGB 14657, PI 189763) (1, 2, 5, 7).

Mutant used for description and seed stocks:

lax-k.84 (NGB 116416) in Bonus.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Larsson, H.E.B. 1985. Linkage studies with genetic markers and some *laxatum* barley mutants. *Hereditas* 103:269-279.
7. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:65.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:147.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 483, Laxatum-I, *lax-I*

Stock number: BGS 483
Locus name: Laxatum-I
Locus symbol: *lax-I*

Previous nomenclature and gene symbolization:

Laxatum-60 = *lax-60* (5, 6).

Inheritance:

Monofactorial recessive (7).

Located likely in chromosome 1H (1); *lax-I.60* is associated with SNP markers 1_0744 to 2_1126 (positions 39.84 to 110.10 cM) in 1H bins 03 to 10 and with small groups of SNP markers in chromosome arms 3HL and 7HS of the Bowman backcross-derived line BW468 (1).

Description:

Rachis internodes of *lax-I.60* were about 23% longer than those of Bonus (6). Spikes exhibited tip sterility, awns were slightly twisted, leaf blades were relatively broad, and kernels were large and angular (6). Plants of the Bowman backcross-derived line for *lax-I.60*, BW468, had visually longer rachis internodes (4).

Origin of mutant:

An ethylene imine induced mutant in Bonus (NGB 14657, PI 189763) (6, 8).

Mutational events:

lax-I.60 (NGB 116394), *-I.78* (NGB 116411) in Bonus (NGB 14657, PI 189763) (2, 3, 6).

Mutant used for description and seed stocks:

lax-I.60 (NGB 116394) in Bonus; *lax-I.60* in Bowman (PI 483237)*2 (BW468, NGB 20698).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
3. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
4. Franckowiak, J.D. (Unpublished).
5. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
6. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
7. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
8. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:66.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:148.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 484, Laxatum-m, *lax-m*

Stock number: BGS 484
Locus name: Laxatum-m
Locus symbol: *lax-m*

Previous nomenclature and gene symbolization:

Laxatum-80 = *lax-80* (3, 4).

Inheritance:

Monofactorial recessive (5).

Location is unknown.

Description:

Rachis internodes of *lax-m.80* were about 16% longer than those of Bonus. The *lax-m.80* plants had light-green leaf blades, slightly coiled awns, and reduced kernel weights (4).

Origin of mutant:

An ethylene imine induced mutant in Bonus (NGB 14657, PI 189763) (4, 6).

Mutational events:

lax-m.65 (NGB 116398), *-m.80* (NGB 116413) in Bonus (NGB 14657, PI 189763) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-m.80 (NGB 116413) in Bonus.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:67.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:149.

Stock number: BGS 485
Locus name: Laxatum-n
Locus symbol: *lax-n*

Previous nomenclature and gene symbolization:

Laxatum-67 = *lax-67* (6, 7).

Inheritance:

Monofactorial recessive (8).

Located in chromosomes 2H, 4HL, or 5H (2); *lax-n.67* is associated with SNP markers 2_1187 to 2_0887 (positions 51.62 to 103.73 cM) in 2H bins 05 to 08, with SNP markers 2_0496 to 2_11332 (positions 72.70 to 112.64 cM in 4H bins 05 to 07, and with SNP markers 2_0010 to 2_1244 (positions 28.11 to 91.91 cM) in 5H bins 02 to 05 of the Bowman backcross-derived line BW469 (2).

Description:

Rachis internodes of *lax-n.67* were about 4% longer than those of Bonus, the awns were coiled, leaf blades were narrow, and kernel weights were low (7). Plants of the Bowman backcross-derived line for *lax-n.67*, BW469, headed about three days later than Bowman and had rachis internodes that were slightly longer (4.8 vs. 4.5 mm). Compared to Bowman, BW469 plants had slightly shorter awns and had two to four more kernels per spike (5). Kernels weighed slightly less than those of Bowman (5.5 vs. 6.0 mg). The awns of BW469 plants were slightly twisted (5). The delayed heading of BW469 and greater number of fertile rachis could be caused by the presence of the late allele at the Early maturity 6 (*Eam6*) or Praematurum-c (*mat-c*) locus, which was retained in the 2H segment from the donor parent Bonus (1, 2).

Origin of mutant:

An ethylene imine induced mutant in Bonus (NGB 14657, PI 189763) (7, 9).

Mutational events:

lax-n.67 (NGB 116400), *-n.68* (NGB 116401) in Bonus (NGB 14657, PI 189763) (7, 8).

Mutant used for description and seed stocks:

lax-n.67 (NGB 116400) in Bonus; *lax-n.67* in Bowman (PI 483237)*3 (BW469, NGB20699).

References:

1. Comadran, J., B. Kilian, J. Russell, L. Ramsay, N. Stein, M. Ganai, P. Shaw, M. Bayer, W. Thomas, D. Marshall, P. Hedley, A. Tondelli, N. Pecchioni, E. Francia, V. Korzun, A. Walther, and R. Waugh. 2012. Natural variation in a homolog of *Antirrhinum CENTRORADIALIS* contributed to spring growth habit and environmental adaptation in cultivated barley. *Nature Genet.* 44:1388-1392.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
3. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
4. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
5. Franckowiak, J.D. (Unpublished).
6. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system

Barley Genetics Newsletter (2017): 46:50-187.

of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.

7. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.

8. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.

9. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:68.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:150-151.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 486, Laxatum-o, *lax-o*

Stock number: BGS 486
Locus name: Laxatum-o
Locus symbol: *lax-o*

Previous nomenclature and gene symbolization:

Laxatum-79 = *lax-79* (3, 4).

Inheritance:

Monofactorial recessive (5).

Location is unknown.

Description:

Rachis internodes of *lax-o.79* were about 14% longer than those of Bonus (4). Plants were relatively vigorous, rachis nodes in the upper part of the spike had enlarged sterile lateral spikelets, some sterile spikelets were observed, awn length was reduced, and kernels were relatively long (4).

Origin of mutant:

An ethylene imine induced mutant in Bonus (NGB 14657, PI 189763) (4, 6).

Mutational events:

lax-o.79 (NGB 116412), -o.88 (NGB 116421) in Bonus (NGB 14657, PI 189763) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-o.79 (NGB 116412) in Bonus.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:69.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:152.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 487, Laxatum-df, *lax-df*

Stock number: BGS 487
Locus name: Laxatum-df
Locus symbol: *lax-df*

Previous nomenclature and gene symbolization:

Laxatum-204 = *lax-204* (3, 4).

Inheritance:

Monofactorial recessive (5).

Location is unknown.

Description:

Rachis internodes of *lax-df.204* were about 2% longer than those of Foma. Plants were somewhat earlier, had relatively good vigor and leaf blades were darker green (4).

Origin of mutant:

A glycidol induced mutant in Foma (NGB 14659, Clho 11333) (4, 6).

Mutational events:

lax-df.204 (NGB 116431), *-df.241* (NGB 116471) in Foma (NGB 14659, Clho 11333) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-df.204 (NGB 115431) in Foma.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:70.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:153.

Stock number: BGS 488
Locus name: Laxatum-ef
Locus symbol: *lax-ef*

Previous nomenclature and gene symbolization:

Laxatum-225 = *lax-225* (6, 7).

Laxatum-238 = *lax-238* (6, 7).

Inheritance:

Monofactorial recessive (5).

Located likely in chromosome 2H (2); *lax-ef.225* is associated with SNP markers 2_1187 to 2_0528 (positions 51.62 to 118.78 cM) in chromosome 2H bins 05 to 08 and with small segments of chromosomes 3HS, 5HS, 5HL, and 6HS of the Bowman backcross-derived line BW462 (2).

Description:

Rachis internodes of *lax-ef.238* spikes were about 23% longer than those of Bonus (7). Leaf blades were relatively wide and darker green; spikes had more fertile rachis nodes, and the straw was very weak when dry (7). Plants of the Bowman backcross-derived line for *lax-ef.225*, BW462, headed three days later than Bowman and had rachis internodes that were nearly 30% longer (5.7 vs. 4.4 mm). Compared to Bowman, BW462 plants had slightly shorter peduncles and awns; and spikes had about 4 to 6 more kernels (5). Kernels were slightly longer and thinner and weighed less (5.3 vs. 6.0 mg). Grain yields of BW462 were 10 to 20% lower than those of Bowman (5). The delayed heading of BW462 could be attributed to the late allele at the Early maturity 6 (*Eam6*) or Praematurum-c (*mat-c*) locus, which was retained in the 2H segment from the donor parent Foma (1, 2).

Origin of mutant:

lax-ef.225 is a glycidol induced mutant in Foma (NGB 14659, CIho 11333) (2, 9); *lax-ef.238* is an ethylene imine induced mutant in Foma (7, 9).

Mutational events:

lax-ef.205 (NGB 116432), *-ef.225* (NGB 116453), *-ef.236* (NGB 116463), *-ef.237* (NGB 116466), *-ef.238* (NGB 116467), *-ef.263* (NGB 116488), *-ef.287* (NGB 116512), *-ef.290* (NGB 116514), *-ef.292* (NGB 116516), *-ef.309* (NGB 116532) in Foma (NGB 14659, CIho 11333) (3, 4, 7, 8).

Mutant used for description and seed stocks:

lax-ef.225 (NGB 116453) in Foma; *lax-ef.225* in Bowman (PI 483237)*3 (BW462, NGB20692).

References:

1. Comadran, J., B. Kilian, J. Russell, L. Ramsay, N. Stein, M. Ganai, P. Shaw, M. Bayer, W. Thomas, D. Marshall, P. Hedley, A. Tondelli, N. Pecchioni, E. Francia, V. Korzun, A. Walther, and R. Waugh. 2012. Natural variation in a homolog of *Antirrhinum CENTRORADIALIS* contributed to spring growth habit and environmental adaptation in cultivated barley. *Nature Genet.* 44:1388-1392.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
3. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of

ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.

4. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.

5. Franckowiak, J.D. (Unpublished).

6. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.

7. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.

8. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.

9. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:71.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:154-155.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 489, Laxatum-ff, *lax-ff*

Stock number: BGS 489
Locus name: Laxatum-ff
Locus symbol: *lax-ff*

Previous nomenclature and gene symbolization:

Laxatum-216 = *lax-216* (3, 4).

Inheritance:

Monofactorial recessive (5).

Location is unknown.

Description:

Rachis internodes of *lax-ff.216* were about equal to those of Foma at normal sowing distances, but rachis internodes were a little longer at lower sowing densities (4). Kernels were relatively round and weighed about 25% less than Foma kernels (4).

Origin of mutant:

An ethyle neimine induced mutant in Foma (NGB 14659, Clho 11333) (4, 6).

Mutational events:

lax-ff.216 (NGB 116444), -ff.233 (NGB 116461) in Foma (NGB 14659, Clho 11333) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-ff.216 (NGB 116444) in Foma.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:72.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:156.

BGS 490, Laxatum-gf, *lax-gf*

Stock number: BGS 490
Locus name: Laxatum-gf
Locus symbol: *lax-gf*

Previous nomenclature and gene symbolization:

Laxatum-217 = *lax-217* (3, 4).

Inheritance:

Monofactorial recessive (5).

Location is unknown.

Description:

Rachis internodes of *lax-gf.217* were 22% longer than those of Foma (4). Plants were relatively vigorous, spikes exhibited irregular rachis internode elongation, and sterile lateral spikelets were enlarged (4).

Origin of mutant:

An ethylene imine induced mutant in Foma (NGB 14659, Clho 11333) (4, 6).

Mutational events:

lax-gf.217 (NGB 116445), *-gf.288* (NGB 116512) in Foma (NGB 14659, Clho 11333) (1, 2, 5, 6,).

Mutant used for description and seed stocks:

lax-gf.217 (NGB 116445) in Foma.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:73.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:157.

Stock number: BGS 491
Locus name: Laxatum-hf
Locus symbol: *lax-hf*

Previous nomenclature and gene symbolization:

Laxatum-244 = *lax-244* (6, 7).

Inheritance:

Monofactorial recessive (5).

Located in chromosomes 2H, 3HL or 5H (2); partially sterile F1 plants indicated that a translocation was present in *lax-hf.244* (6); *lax-hf.244* is associated with SNP markers 2_1338 to 2_1242 (positions 74.97 to 133.59 cM) in chromosome 2H bins 05 to 08, with SNP markers 2_0931 to 2_0009 (positions 104.39 to 164.68 cM) in chromosome 3H bins 07 to 11, and with SNP markers 1_1381 to 2_0392 (positions 11.26 to 109.27 cM) in chromosome 5H bins 01 to 06 of the Bowman backcross-derived line BW464 (2). Based on translocation testers, *lax-hf.244* was previously associated with chromosome 5HL (9).

Description:

Rachis internodes of *lax-hf.244* plants were about 21% longer than those of Bonus (7). Leaf blades were relatively narrow and darker green; spikes were long, and kernels were relatively thin (7). A translocation could be present in the original *lax-hf.244* stock based on semisterility of the F1s following a cross to Foma (8). Plants of the Bowman backcross-derived line for *lax-hf.244*, BW464, headed two to three days later than Bowman and had rachis internodes that were about 15% longer (5.2 vs. 4.5 mm). Compared to Bowman, BW464 plants had slightly shorter peduncles and awns and had about four more kernels per spike (5). Kernels were longer (10.5 vs. 9.3 mm), thinner (3.3 vs. 3.8 mm) and weighed less (5.3 vs. 6.0 mg). Grain yields of BW464 were 15 to 20% lower than those of Bowman (5). The delayed heading of BW464 could be attributed to the late allele at the Early maturity 6 (*Eam6*) or Praematurum-c (*mat-c*) locus, which was retained in the 2H segment from the donor parent Foma (1, 2).

Origin of mutant:

lax-hf.244 is an ethyl methanesulfonate induced mutant in Foma (NGB 14659, Clho 11333) (7, 10).

Mutational events:

lax-hf.232 (NGB 116460), *-hf.242* (NGB 116472), *-hf.244* (NGB 116474), *-hf.251* (NGB 116478) in Foma (NGB 14659, Clho 11333) (3, 4, 8, 10).

Mutant used for description and seed stocks:

lax-hf.244 (NGB 116474) in Foma; *lax-hf.244* in Bowman (PI 483237)*4 (BW464, NGB20694).

References:

1. Comadran, J., B. Kilian, J. Russell, L. Ramsay, N. Stein, M. Ganai, P. Shaw, M. Bayer, W. Thomas, D. Marshall, P. Hedley, A. Tondelli, N. Pecchioni, E. Francia, V. Korzun, A. Walther, and R. Waugh. 2012. Natural variation in a homolog of *Antirrhinum CENTRORADIALIS* contributed to spring growth habit and environmental adaptation in cultivated barley. *Nature Genet.* 44:1388-1392.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011.

Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.

3. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.

4. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.

5. Franckowiak, J.D. (Unpublished).

6. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.

7. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.

8. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.

9. Larsson, H.E.B. 1985. Linkage studies with genetic markers and some *laxatum* barley mutants. *Hereditas* 103:269-279.

10. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:74.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:158-159.

Stock number: BGS 492
Locus name: Laxatum-if
Locus symbol: *lax-if*

Previous nomenclature and gene symbolization:

Laxatum-246 = *lax-246* (3, 4).

Inheritance:

Monofactorial recessive (5), but partially sterile F1 plants indicated that a translocation was present in *lax-if.246* (5).

Location is unknown.

Description:

Rachis internodes of *lax-if.246* were about equal to those of Foma at normal sowing distances, but rachis internodes were a little longer at lower sowing densities (4). Spikes that emerged earlier had slightly longer rachis internodes than later emerging spikes (4).

Origin of mutant:

A propyl methanesulfonate induced mutant in Foma (NGB 14659, Clho 11333) (4, 6).

Mutational events:

lax-if.246 (NGB 116475), *-if.248* (NGB 116477) in Foma (NGB 14659, Clho 11333) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-if.246 (NGB 116475) in Foma.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:75.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:160.

BGS 493, Laxatum-jf, *Lax-jf*

Stock number: BGS 493
Locus name: Laxatum-jf
Locus symbol: *Lax-jf*

Previous nomenclature and gene symbolization:

Laxatum 253 = *Lax-253* (5, 6).

Laxatum 255 = *Lax-255* (5, 6).

Inheritance:

Monofactorial dominant (4, 7).

Located in chromosome 4HS (1); *Lax-jf.253* is associated with SNP markers 2_0145 to 2_1558 (positions 2.58 to 5.42 cM) in chromosome 4H bin 01 of the Bowman backcross-derived line BW466 (1). Based on translocation breakpoints, *Lax-jf.255* was previously reported to have a weak linkage to chromosome 5H (8).

Description:

Rachis internodes of *Lax-jf.255* were about 12% longer than those of Bonus (6). Spikes had a few sterile spikelets and the kernels were thin and angular (6). Plants of the Bowman backcross-derived line for *Lax-jf.253*, BW466, were slightly taller and had rachis internodes that were 5 to 10% longer (4.8 vs. 4.5 mm) (4). Compared to Bowman, the *Lax-jf.253*, BW466, plants had slightly shorter peduncles and 2 to 4 more kernels per spike. Kernels of BW466 were of similar size and weight as those of Bowman. Grain yields of BW466 were on average equal to those of Bowman (4).

Origin of mutant:

Lax-jf.255 is a propyl methanesulfonate induced mutant in Foma (NGB 14659, Clho 11333) (6, 9). *Lax-jf.253* is an ethyl methanesulfonate induced mutant in Foma (1, 9).

Mutational events:

Lax-jf.253 (NGB 116480, GSHO 1571), *-jf.254* (NGB 116481), *-jf.255* (NGB 116482 in Foma (NGB 14659, Clho 11333) (2, 3, 6, 7).

Mutant used for description and seed stocks:

Lax-jf.253 (NGB 116480, GSHO 1571) in Foma; *Lax-jf.255* (NGB 116482) in Foma; *Lax-jf.253* in Bowman (PI 483237)*3 (GSHO 2138); *Lax-jf.253* in Bowman (PI 483237)*5 (BW466, NGB20696).

References;

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
3. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
4. Franckowiak, J.D. (Unpublished).
5. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
6. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
7. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.

Barley Genetics Newsletter (2017): 46:50-187.

8. Larsson, H.E.B. 1985. Linkage studies with genetic markers and some *laxatum* barley mutants. *Hereditas* 103:269-279.

9. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:76.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:160-162.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 494, Laxatum-kf, *lax-kf*

Stock number: BGS 494
Locus name: Laxatum-kf
Locus symbol: *lax-kf*

Previous nomenclature and gene symbolization:

Laxatum-295 = *lax*-295 (3, 4).

Inheritance:

Monofactorial recessive (5), but partially sterile F1 plants indicated that a translocation was present in *lax-kf.295* (5).

Location is unknown.

Description:

Rachis internodes of *lax-kf.295* were about equal to those of Foma at normal sowing distances, but rachis internodes were a little longer at lower sowing densities (4). Spikes that emerged earlier had slightly longer rachis internodes than later emerging spikes (4).

Origin of mutant:

An ethyl methanesulfonate induced mutant in Foma (NGB 14659, Clho 11333) (4, 6).

Mutational events:

lax-kf.268 (NGB 116493), *-kf.295* (NGB 116519) in Foma (NGB 14659, Clho 11333) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-kf.295 (NGB 116493) in Foma.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:77.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:163.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 495, Laxatum-lf, *lax-lf*

Stock number: BGS 495
Locus name: Laxatum-lf
Locus symbol: *lax-lf*

Previous nomenclature and gene symbolization:

Laxatum-274 = *lax-274* (3, 4).

Inheritance:

Monofactorial recessive (5).

Location is unknown.

Description:

Rachis internodes of *lax-lf.274* were about 16% longer than those of Foma (4). Compared to Foma, spikes were long and kernels were large (4).

Origin of mutant:

An ethyl methanesulfonate induced mutant in Foma (NGB 14659, Clho 11333) (4, 6).

Mutational events:

lax-lf.273 (NGB 116499), *-lf.274* (NGB 116500) in Foma (NGB 14659, Clho 11333) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-lf.274 (NGB 116500) in Foma.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:78.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:164.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 496, Laxatum-mf, *lax-mf*

Stock number: BGS 496
Locus name: Laxatum-mf
Locus symbol: *lax-mf*

Previous nomenclature and gene symbolization:

Laxatum-302 = *lax-302* (3, 4).

Inheritance:

Monofactorial dominant (5), but partially sterile F1 plants indicated that a translocation was present in *lax-mf.302* (5).

Location is unknown.

Description:

Rachis internodes of *lax-mf.302* were about 15% longer than those of Foma (4). Plants had few tillers and matured late. Compared to Foma, spikes were short and kernels weighed less (4).

Origin of mutant:

An ethyl methanesulfonate induced mutant in Foma (NGB 14659, Clho 11333) (4, 6).

Mutational events:

lax-mf.300 (NGB 116524), *-mf.301* (NGB 116525), *-mf.302* (NGB 116526) in Foma (NGB 14659, Clho 11333) (1, 2, 5, 6).

Mutant used for description and seed stocks:

lax-mf.302 (NGB 116526) in Foma.

References:

1. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
3. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
4. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
5. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas* 103:255-267.
6. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:79.

Revised:

U. Lundqvist. 2017. *Barley Genet. Newsl.* 47:165.

Stock number: BGS 497
Locus name: Laxatum-nf
Locus symbol: *lax-nf*

Previous nomenclature and gene symbolization:

Laxatum-322 = *lax*-322 (5, 6).

Laxatum-323 = *lax*-323 (5, 6).

Inheritance:

Monofactorial recessive (7).

Located likely in chromosomes 2HL or 4H (1); *lax-nf.322* is associated with SNP markers 1_0786 to 2_1474 (positions 133.59 to 218.47 cM) in chromosome 2H bins 09 to 13, with SNP markers 1_0113 to 1_0568 (positions 26.58 to 78.18 cM) in chromosome 4H bins 02 to 06, and with SNP markers in small segments of chromosomes 3HL, 5HL, 6HS, and 6HL of the Bowman backcross-derived line BW470 (1).

Description:

Rachis internodes of *lax-nf.322* were about 29% longer than those of Foma (6). Kernels were relatively thin and weighed less than half those of Foma. Spikes showed tip sterility (6). Compared to Bowman the Bowman backcross-derived line for *lax-nf.322*, BW470, had rachis internodes that were about 25% longer (5.6 vs. 4.5 mm). BW470 plants were 5 to 10% taller, had slightly shorter awns, and 1 to 3 more fertile spikelets (4). Kernels were longer (10.5 vs. 9.2 mm) and thinner (3.3 vs. 3.8 mm) and weighed less (5.2 vs. 6.0 mg). Grain yields of BW470 were about 25% lower than those of Bowman (4).

Origin of mutant:

lax-nf.322 and *lax-nf.323* are ethyl methanesulfonate induced mutants in Foma (NGB 14659, CIho 11333) (7, 8).

Mutational events:

lax-nf.322 (NGB 116542), *-nf.323* (NGB 116543) in Foma (NGB 14659, CIho 11333) (2, 3, 7, 8).

Mutant used for description and seed stocks:

lax-nf.322 (NGB 116542) in Foma; *lax-nf.323* NGB 116543) in Foma; *lax-nf.322* in Bowman (PI 483237)*3 (BW470, NGB20700).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1959. The mutagenic effects of ionizing radiations and reactive ethylene derivatives in barley. *Hereditas* 45:351-368.
3. Ehrenberg, L., Å. Gustafsson, and U. Lundqvist. 1961. Viable mutants induced in barley by ionizing radiations and chemical mutagens. *Hereditas* 47:243-282.
4. Franckowiak, J.D. (Unpublished).
5. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.
6. Larsson, H.E.B. 1985. Morphological analysis of *laxatum* barley mutants. *Hereditas* 103:239-253.
7. Larsson, H.E.B. 1985. Genetic analysis of *laxatum* barley mutants. *Hereditas*

Barley Genetics Newsletter (2017): 46:50-187.

103:255-267.

8. Lundqvist, U. (Unpublished).

Prepared:

H.E.B. Larsson and U. Lundqvist. 1986. *Barley Genet. Newsl.* 16:80.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:166-167.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 518, Semidwarf 1, *sdw1*

Stock number: BGS 518
Locus name: Semidwarf 1
Locus symbol: *sdw1*

Previous nomenclature and gene symbolization:

Denso dwarf = *denso* (7, 18).

Hordeum vulgare gibberellin 20-oxidase = *Hv20ox₂* (10, 11, 21).

Inheritance:

Monofactorial recessive (7, 19); some F₁'s are intermediate in height (1, 13).

Location in chromosome 3HL (2, 14); *sdw1.d* is probably proximal from the *gsh2* (glossy sheath 2) locus and near RFLP marker PSR170 (14); *sdw1.d* is in bin 3H-11 (12); *sdw1.d* is near RFLP marker R1545 (22); *sdw1.d* is close to SNP marker 1_0867 (15); *sdw1.a* is associated with SNP markers 1_0044 to 2_0650 (positions about 190 to 192.00 cM) in 3H bin 12 of the Bowman backcross-derived line BW827 (4); *sdw1.d* is associated with SNP markers 2_0023 to 1_0821 (positions 169.94 to 190.87 cM) in 3H bins 11 and 12 of the Bowman backcross-derived line BW828 (4), in 3H bin 12.

Description:

Plants homozygous for the *sdw1.a* gene ranged from 10 to 30 cm shorter than normal sibs, with expression partial dependent on environment (1, 18, 20). Spike length was variable, but fully as long as normal barley. The stock used for description of the *sdw1.a* gene, M21, has the short straw and long spike of the original 'Jotun Mutant' as well as a large culm diameter from its parent 'Vantage' (1, 20). The semidwarf mutants, 'Diamant' and 'Abed Denso', are alleles at the *sdw1* locus (7, 16). Alleles at the *sdw1* locus are associated with semi-prostrate juvenile growth (7, 18), delayed maturity (5, 7, 8, 18, 22), smaller grain size (7), and reduced malt quality (5, 8, 18). The *sdw1* mutants are GA sensitive (3, 23). They are very likely mutants in an orthologue of the rice *sd1* gene (23), which encodes a GA-oxidase that produces lower levels of GA and reduced levels of cause the dwarf phenotype (9, 17). A gibberellin 20-oxidase gene (*Hv20ox₂*) was identified as the candidate gene for *sdw1* (10, 11). The reduced expression of *Hv20ox₂* increased the number of effective tillers and enhanced grain yield (11, 21). The reduction in *Hv20ox₂* levels was 4-fold in the *sdw1.d* mutant and 60-fold in the *sdw1.a* mutant, which causes a reduced endogenous GAs concentration and delayed heading (11). Plants of the Bowman backcross-derived line for *sdw1.a*, BW827, were 15 to 30% shorter than Bowman and heading was delayed 1 to 3 days. Plants of the Bowman backcross-derived line for *sdw1.d*, BW828, were 10 to 20% shorter than Bowman, heading was delayed 1 to 3 days, and spikes had 1 to 2 more kernels in some trials. Kernels of BW827 and BW828 were up to 10% lighter than Bowman kernels in some trials. Grain yields of BW828 were similar to those of Bowman, while those of BW827 were about 85% of the Bowman yields (6). The QTL for *sdw1.d* co-located with QTL for yield, plant height, development score, hectoliter weight and grain plumpness (11).

Origin of mutant:

An X-ray induced mutant in the Norwegian cultivar Jotun (PI 467357) isolated as Jotun 22 by Knut Mikaelson (1, 13).

Mutational events:

sdw1.a (66/86, GSHO 1414) in Jotun (PI 467357) (20); *sdw1.c* in Abed Denso (PI 361639) (7); *sdw1.d* (*denso*) (Diamant, PI 330397, PI 467775) in Valticky (PI 268173)

(7); *sdw1.e* (Riso no. 9265) in Abed Bomi (NGB 5096, PI 43371) (7, 9). Some cultivars demonstrated to have the *sdw1.a* and *sdw1.d* alleles are listed in Xu et al. (21). Note that the *denso* name was originally assigned to the *sdw1.c* mutant (7), but *sdw1.d* mutant from Diamant is the only allele present in cultivars described as having the *denso* semidwarf (21).

Mutant used for description and seed stocks:

sdw1.a in M21 (Clho 15481, GSHO 2513) from the cross Jotun Mutant/Kindred//Vantage (20); *sdw1.d* in Trumpf (Triumph, PI 548762, GSHO 2465) from Diamant; *sdw1.a* from a Jotun derivative in Bowman (PI 483237)*7 (GSHO 1978), *sdw1.a* in Bowman*8 (BW827, NGB 22264); *sdw1.d* from Trumpf in Bowman*4 (GSHO 1979), *sdw1.d* in Bowman*5 (BW828, NGB 22265).

References:

1. Ali, M.A.M., O. Okiror, and D.C. Rasmusson. 1978. Performance of semidwarf barley. *Crop Sci.* 18:418-422.
2. Barau, U.M., K.J. Chambers, W.T.B. Thomas, C.A. Hackett, V. Lea, P. Jack, B.P. Forster, R. Waugh, and W. Powell. 1994. Molecular mapping of genes determining height, time to heading, and growth habit in barley (*Hordeum vulgare*). *Genome* 36:1080-1087.
3. Boulger, M.C., R.G. Sears, and W.E. Kronstad. 1982. An investigation of the association between dwarfing sources and gibberellic acid response in barley. p. 550-553. *In* M.J.C. Asher, R.P. Ellis, A.M. Hayter, and R.N.H. Whitehouse (eds.) *Barley Genetics IV. Proc. Fourth Int. Barley Genet. Symp.*, Edinburgh. Edinburgh Univ. Press, Edinburgh.
4. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
5. Foster, A.E., and A.P. Thompson. 1987. Effects of a semidwarf gene from Jotun on agronomic and quality traits of barley. p. 979-982. *In* S. Yasuda and T. Konishi (eds.) *Barley Genetics V. Proc. Fifth Int. Barley Genet. Symp.*, Okayama, 1986. Sanyo Press Co., Okayama.
6. Franckowiak, J.D. (Unpublished).
7. Haahr, V., and D. von Wettstein. 1976. Studies of an induced, high-yielding dwarf-mutant of spring barley. p. 215-218. *In* H. Gaul (ed.) *Barley Genetics III, Proc. Third Int. Barley Genet. Symp.*, Garching, 1975. Verlag Karl Thieme, München.
8. Hellewell, K.B., D.C. Rasmusson, M. Gallo-Meagher. 2000. Enhancing yield of semidwarf barley. *Crop Sci.* 40:352-358.
9. Jia, Q., C. Li, Y. Shang, J. Zhu, W. Hua, J. Wang, J. Yang, and G. Zhang. 2015. Molecular characterization and functional analysis of barley semi-dwarf mutant Riso no. 9265. *BMC Genomics* 16: 927.
10. Jia, Q.J., J. Zhang, S. Westcott, X.Q. Zhang, M. Bellgard, R. Lance, and C.G. Li. 2009. GA-20 oxidase as a candidate for the semidwarf gene *sdw1/denso* in barley. *Funct. Integr. Genomics* 9:255-262.
11. Jia, Q., X.Q. Zhang, S. Westcott, S. Broughton, M. Cakir, J. Yang, R. Lance, and C. Li. 2011. Expression level of a gibberellin 20-oxidase gene is associated with multiple agronomic and quality traits in barley. *Theor. Appl. Genet.* 122:1451-1460.
12. Kleinhofs, A. 2006. Integrating molecular and morphological/physiological marker maps. *Barley Genet. Newsl.* 36:66-82.
13. Lambert, J.W., and M. Shafi. 1959. Inheritance and heritability of height in three

barley crosses. *Barley Newsl.* 3:7-8. (Abstr.)

14. Laurie, D.A., N. Pratchett, C. Romero, E. Simpson, and J.W. Snape. 1993. Assignment of the *denso* dwarfing gene to the long arm of chromosome 3 (3H) of barley by use of RFLP markers. *Plant Breed.* 111:198-203.

15. Malosetti, M., F.A. van Eeuwijk, M.P. Boer, A.M. Casas, M. Elía, M. Moralejo, P.R. Bhat, L. Ramsay, and J.-L. Molina-Cano. 2011. Gene and QTL detection in a three-way barley cross under selection by a mixed model with kinship information using SNPs. *Theor. Appl. Genet.* 122:1605-1616.

16. Mickelson, H.R., and D.C. Rasmusson. 1994. Genes for short stature in barley. *Crop Sci.* 34:1180-1183.

17. Murai, M., T. Komazaki, and S. Sato. 2004. Effects of *sd1* and *Ur1* (Undulate rachis - 1) on lodging resistance and related traits in rice. *Breed. Sci.* 54: 333-340.

18. Powell, W., P.D.J. Caligari, W.T.B. Thomas, and J.L. Jinks. 1985. The effects of major genes on quantitatively varying characters in barley. 2. The *denso* and day length response loci. *Heredity* 54:349-352.

19. Powell, W., P.D.J. Caligari, W.T.B. Thomas, and J.L. Jinks. 1991. The effects of major genes on quantitatively varying characters in barley. 4. The *GPert* and *denso* loci and quality characters. *Heredity* 66:381-389.

20. Rasmusson, D.C., E.E. Bantari, and J.W. Lambert. 1973. Registration of M21 and M22 semidwarf barley. *Crop Sci.* 13:777.

21. Xu, Y., Q. Jia, G. Zhou, X.-Q. Zhang, T. Angessa, S. Broughton, G. Yan, W. Zhang, and C. Li. 2017. Characterization of the *sdw1* semi-dwarf gene in barley. *BMC Plant Biology* 17:11.

22. Yin, X., P.C. Struik, F.A. van Eeuwijk, P. Stam, and J. Tang. 2005. QTL analysis and QTL-based prediction of flowering phenology in recombinant inbred lines of barley. *J. Exp. Bot.* 56:967-976.

23. Zhang, J., X. Yang, P. Moolhuijzen, C. Li, M. Bellgard, R. Lance, and R. Appels. 2005. Towards isolation of the barley green revolution gene. *Proc. Australian Barley Tech. Symp* 2005.

[http://www.cdesign.com.au/proceedings_abts2005/posters%20\(pdf\)/poster_li.pdf](http://www.cdesign.com.au/proceedings_abts2005/posters%20(pdf)/poster_li.pdf).

Prepared:

D.C. Rasmusson. 1988. *Barley Genet. Newsl.* 18:87 as BGS 468.

Revised:

J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:444-445.

J.D. Franckowiak. 2007. *Barley Genet. Newsl.* 37:277-278.

J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:176-178.

J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:168-170.

BGS 525, Eceriferum-yl, *cer-yl*

Stock number: BGS 525
Locus name: Eceriferum-yl
Locus symbol: *cer-yl*

Previous nomenclature and gene symbolization:

None.

Inheritance:

Monofactorial recessive (4).

Located in chromosome 4HL (1); *cer-yl.187* is associated with SNP markers 2_0411 to 1_0062 (positions 52.11 to 91.93 cM) in 4H bins 04 to 06 of the Bowman backcross-derived line BW143 (1).

Description:

Surface wax coating on the spike, the leaf sheath, and the stem appears absent (wax code - - ++) (4). The original mutant stock produces weak plants (about 3/4 normal height) with a high level of sterility (4). Plants of the Bowman backcross-derived line for *cer-yl.187*, BW143, are semidwarf with waxy nodes, coiled peduncles, globose shaped seeds, and poor hull attachment. Reduced seed set can be caused by adverse environmental conditions (2). The BW143 plants headed 1 to 2 days later than Bowman and were shorter, 67 v. 89 cm. Peduncles and leaf blades were about 2/3 normal length. Kernels were thinner and weighed less, 4.4 vs. 5.9 mg and grain yields were 1/4 to 1/3 those of Bowman (2). Although not as extreme, the phenotypic characteristics of BW143 are similar to those of the Bowman backcross-derived lines for *cer-zv.268* (BW177) and *cer-ym.753* (BW144). Since all three loci retain donor parent markers in the same region of 4HL, they might have mutations in the same gene (1, 2). However, allelism tests demonstrated that the mutants at these loci are not alleles (3).

Origin of mutant:

A propane disulfonic acid diethyl ester induced mutant in Bonus (NGB 14657, PI 189763) (3).

Mutational events:

cer-yl.187 (NGB 111073, GSHO 1545), *-yl.188* (NGB 111074) in Bonus (NGB 14657, PI 189763) (4); *cer-yl.407* (NGB 111295) in Foma (NGB 14659, Clho 11333), *-yl.821* (NGB 111709) in Bonus (5).

Mutant used for description and seed stocks:

cer-yl.187 (GSHO 1545, NGB 111073) in Bonus; *cer-yl.187* in Bowman (PI 483237)*6 (GSHO 2216); *cer-yl.187* in Bowman*7 (BW143, NGB 20549). GSHO 1545 and GSHO 2216 do not show the phenotype of *cer-yl.187*, but rather the phenotype of *cer-zv.268* or *cer-ym.753*. NGB 111073 and NGB 20549 have the correct phenotypes (3).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendrarnin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Lundqvist, U. (Unpublished).
4. Lundqvist, U., and D. von Wettstein. 1977. Stock list for the eceriferum mutants IV. *Barley Genet. Newsl.*:92-96.
5. Lundqvist, U., and D. von Wettstein. 1982. Stock list for the eceriferum mutants VI.

Barley Genetics Newsletter (2017): 46:50-187.

Barley Genet. Newsl. 12:169-172.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. Barley Genet. Newsl. 26:452.

Revised:

U. Lundqvist and J.D. Franckowiak. 2016. Barley Genet. Newsl. 46:116.

U. Lundqvist and J.D. Franckowiak. 2017. Barley Genet. Newsl. 47:171-172.

Stock number: BGS 526
Locus name: Eceriferum-ym
Locus symbol: *cer-ym*

Previous nomenclature and gene symbolization:

None.

Inheritance:

Monofactorial recessive (7).

Located in chromosome 4HL (1, 3, 4); *cer-ym.753* is associated with SNP markers 2_0289 to 1_1513 (positions 69.62 to 101.44 cM) in 4H bins 05 to 07 of the Bowman backcross-derived line BW144 (1); *cer-zv.268*, a possible allele (4), is associated with SNP markers 2_1122 to 1_0568 (positions 47.80 to 78.18 cM) in 4H bins 05 to 06 of the Bowman backcross-derived line BW177 (1); *cer-ym.753* and *cer-zv.268* are in a 0.8 cM interval between EST markers AK370363 and AK251484 in pericentromeric region of 4H (3, 4).

Description:

Surface wax coating on the spike, the leaf sheath and stem, and the leaf blade appears absent (wax code - - -) (7). Plants are weak, culms have a kinked peduncle, nodes are wax covered, and kernels are small, globe shaped, and nearly hull-less (2, 5). Plants of the Bowman backcross-derived line for *cer-ym.753*, BW144, averaged about 2/3 the height of Bowman with peduncles and leaf blades about 2/3 normal length. Awns of BW144 were slightly shorter and kernels were smaller and lighter, 4.2 vs. 6.1 mg. The grain yields of BW144 were less than 1/3 that of Bowman. Plants of the Bowman backcross-derived line for *cer-zv.268* (BW177) were phenotypically similar to BW144 (*cer-ym.753*) plants, but they were weaker (2, 3). Detached leaves of the mutant *cer-ym.753* lost 41.8% of their initial weight after 1 hour of dehydration under laboratory conditions, while that of the wild type Bowman plants lost only 7.1%. Stomata function was not affected by the mutation, but the mutant leaves showed increased cuticular permeability to water, suggesting a defective leaf cuticle, which was confirmed by toluidine blue staining (3, 4). The *cer-ym.753* leaf blades showed a substantial reduction in the amounts of the major cutin monomers and a slight increase in the main wax component, suggesting that the enhanced cuticle permeability was a consequence of cutin deficiency (3, 4). Allelism tests demonstrated that mutants at the *cer-ym* and *cer-zv* loci are not alleles (5).

Origin of mutant:

An ethylene oxide induced mutant in Bonus (NGB 14657, PI 189763) (5).

Mutational events:

cer-ym.130, (NGB 111015, GSHO 1546), *-ym.753* (NGB 111641) in Bonus (NGB 14657, PI 189763) (7); *cer-zv.268* (NGB 111155, GSHO 1529), *-zv.342* (NGB 111229), *-zv.576* (NGB 111464) in Foma (NGB 14659, CIho 11333) (6); *cer-zv.1256* (NGB 112144) in Kristina (NGB 14661, NGB 1500) (8).

Mutant used for description and seed stocks:

cer-ym.753 (NGB 111641, GSHO 1546) in Bonus; *cer-ym.753* in Bowman (PI 483237)*5 (GSHO 2217); *cer-ym.753* in Bowman*7 (BW144, NGB 20550).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S.

- Radovic, F. Shahinnia, V. Vendrarnin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Li, C., C. Liu, X. Ma, A. Wang, R. Duan, C. Nawrath, T. Komatsuda, and G. Chen. 2015. Characterization and genetic mapping of *eceriferum-ym* (*cer-ym*), a cutin deficient barley mutant with impaired leaf water retention capacity. *Breed. Sci.* 65:327-332.
4. Li, C., A. Wang, X. Ma, M. Pourkheirandish, S. Sakuma, N. Wang, S. Ning, E. Nevo, C. Nawrath, T. Komatsuda, and G. Chen. 2013. An *eceriferum* locus, *cer-zv*, is associated with a defect in cutin responsible for water retention in barley (*Hordeum vulgare*) leaves. *Theor. Appl. Genet.* 126:637-646.
5. Lundqvist, U. (Unpublished).
6. Lundqvist, U., and D. von Wettstein. 1973. Stock list for the *eceriferum* mutants II. *Barley Genet. Newsl.* 3:110-112.
7. Lundqvist, U., and D. von Wettstein. 1977. Stock list for the *eceriferum* mutants IV. *Barley Genet. Newsl.* 7:92-96.
8. Lundqvist, U., and D. von Wettstein. 1979. Stock list for the *eceriferum* mutants V. *Barley Genet. Newsl.* 9:135-137.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:453.

Revised:

U. Lundqvist and J.D. Franckowiak. 2016. *Barley Genet. Newsl.* 46:118-119.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:173-174.

Stock number: BGS 554
Locus name: Breviaristatum-m
Locus symbol: *ari-m*

Revised locus symbol:

The allelism based similarities in phenotypes of the Bowman-backcross-derived lines (3) and retained donor parent SNP markers (2) for the *ari-m.28* and several *brh1* mutants (2) was confirmed by Braumann et al. (1). Therefore, to find more information about this locus, see BGS 001, the barley genetic stock (BGS) description for the Brachytic 1 (*brh1*) locus.

Previous nomenclature and gene symbolization:

Brachytic = *br* (1, 8, 9).
Breviaristatum-i = *ari-i* (5, 6).
Breviaristatum-28 = *ari-28* (7).

Inheritance:

Monofactorial recessive (6).
Location in chromosome 7HS (3); *ari-m.28* is associated with SNP markers 2_0710 to 2_0307 (positions 4.74 to 9.55 cM) in 7H bin 01 of the Bowman backcrossed-derived line BW051 (3); the genomic position of the *Brh1* locus at 12.74 cM in 7HS (1), in 1H bin 01 near the border with 1H bin 02.

Description:

Plants are semidwarf (2/3 normal height) and can be placed in the brachytic class of semidwarfs (4). Kernels are globe-shaped and awns are about 1/2 normal length (4). Spikes are slightly more lax than normal (4). Plants of the Bowman backcross-derived line for *ari-m.28* (BW051) are phenotypically similar to other Bowman backcross-derivates with mutant alleles at the *Brh1* (Brachytic 1) locus (1). The *ari-m* mutants occur in the *Brh1* locus and are deficient in the Gα subunit of a heterotrimeric G protein, which is an important regulator of culm length in barley (1).

Origin of mutant:

A gamma-ray induced mutant in Bonus (NGB14657, PI 189763) (6).

Mutational events:

ari-m.12 (NGB 115858, GSHO 1661), *-m.28* (NGB 115876) in Bonus (NGB14657, PI 189763), *-m.141* (NGB 115951), *-m.177* (NGB 115987) in Foma, (NGB 14659, Clho 11333), *-m.251* (NGB 116059) in Kristina (NGB 14661, NGB 1500) (6); *ari-m.269* (NGB 116081) in Kristina (6, 7).

Mutant used for description and seed stocks:

ari-m.28 (GSHO 1661, NGB 115876) in Bonus; *ari-m.28* in Bowman (PI 483237)*6 (GSHO 2161); *ari-m.28* in Bowman*8 (BW051, NGB 20459).

References:

1. Braumann, I., C. Dockter, S. Beier, A. Himmelbach, F. Lok, U. Lundqvist, B. Skadhauge, N. Stein, S. Zakhrabekova, R. Zhou, and M. Hansson. 2018. Mutations in the gene of the Gα subunit of the heterotrimeric G protein are the cause for the *brachytic1* semi-dwarf phenotype in barley and applicable for practical breeding. *Hereditas* 155:10.
2. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011.

Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.

3. Franckowiak, J.D. (Unpublished).

4. Franckowiak, J.D. 1995. The brachytic class of semidwarf mutants in barley. *Barley Genet. Newsl.* 24:56-59.

5. Gustafsson, Å., A. Hagberg, U. Lundqvist, and G. Persson. 1969. A proposed system of symbols for the collection of barley mutants at Svalöv. *Hereditas* 62:409-414.

6. Kucera, J., U. Lundqvist, and Å. Gustafsson. 1975. Inheritance of brevistaristatum mutants in barley. *Hereditas* 80:263-278.

7. Lundqvist, U. (Unpublished).

8. Powers, L. 1936. The nature of the interactions of genes affecting four quantitative characters in a cross between *Hordeum deficiens* and *vulgare*. *Genetics* 21:398-420.

9. Swenson, S.P. 1940. Genetic and cytological studies on a brachytic mutant in barley. *J. Agric. Res.* 60:687-713.

Prepared:

U. Lundqvist and J.D. Franckowiak. 1997. *Barley Genet. Newsl.* 26:480.

Revised:

U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:184.

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:175-176.

Stock number: BGS 614
Locus name: Zeocriton 2
Locus symbol: Zeo2

Revised locus symbol:

Since the zeocriton spike phenotype was first described by Hayes and Harlan (7), *Zeo1* became the recommended locus symbol for various mutants at this locus. The semi-dominant dense spike variants at the complex *Zeo1* locus exhibit a range of phenotypes associated with restricted elongation of the rachis internodes. Based on DNA sequencing, three distinct phenotypic groups of dense spike variants are associated with the *Hordeum vulgare* *APELATA2* (*AP2*)-like transcription factor (*HvAP2*) locus (9, 10). The phenotypic expressions were historically assigned different locus symbols, but each group is associated with specific molecular changes in the *HvAP2* transcript (9). Alleles at this locus are assigned locus symbols: The *Zeo1* mutants exhibit the most extreme phenotypes, the *Zeo2* variants have intermediate phenotypes, and the *Zeo3* variants may show a lesser degree in shortening of rachis internodes (9). Molecular markers and/or pedigree information are necessary to separate variants assigned to the *Zeo1* (BGS 082) and *Zeo3* (BGS 184) groups (8). See BGS 082 for more information of the alleles at the *Zeo1* locus (9).

Previous nomenclature and gene symbolization:

Zeocriton with no gene symbol (4, 7).

Zeocriton 2 = *Zeo2* (6, 9).

Rachis internode length QTL in 2HL = *qSIL.ak-2H* (11).

Cleistogamy 1 = *cly1.b* (8, 9, 13, 14).

Hordeum vulgare *APELATA2* (*AP2*)-like transcription factor = *HvAP2* (9, 10).

Inheritance:

Monofactorial semi-dominant (2, 11).

Located in chromosome 2HL (2, 11, 17); *Zeo2* from Haruna Nijo is in 2H bin 13 (2); *qSIL.ak* (*Zeo2*) from Kanto Nakate Gold is near SRS marker ABG613 (11); *Zeo2* is closely to the *Cleistogamy 1* (*cly1.b*) locus (10, 13, 14); *Zeo2.c* is associated with SNP markers 1_0404 to 1_0072 (positions 186.61 to 239.78 cM) in 2HL bins 12 to 14 of the Bowman backcross-derived line BW939 (3); the dense spike traits in the *dsp1.a* stock (likely *Zeo2.c*) is associated with SNP markers 1_0376 to 2_0561 (positions 209.87 to 247.86 cM) in 2HL bins 13 to 14 of the Bowman backcross-derived line BW277 (3); *Zeo2* with the *eog1.e* gene from GSHO 285 (Ab 1380) is associated with SNP markers 1_1480 to 2_0895 (positions 173.50 to 209.91 cM) in 2HL bins 11 to 13 of the Bowman backcross derived line BW302 (3); *Zeo2.ax* (previously named *dsp.ax*) is associated with SNP markers 2_0064 to 2_0175 (positions 179.99 to 213.08 cM) in 2H bins 11 to 13 of the Bowman backcross-derived line BW270 (3); *Zeo2* from Haruna Nijo in the *ant22.1508* stock is associated with SNP markers 1_1346 to 2_0895 (positions about 165 to 209.87 cM) in 2H bins 11 to 13 of the Bowman backcross-derived line BW022 (3); the *HvAP2* locus is near marker BOPA2_12_10579 (9), in 2H bin 13.

Description:

Many dense spike phenotypes with shortened rachis internodes have been described in the barley literature (12). Hayes and Harlan (7) identified a strong pyramid shaped spike with short internodes as zeocriton (little barley), but they reported that three genes

controlled this trait in their cross to Zeocriton (8). Accession Clho 6238 is phenotypically to the Zeocriton cultivar. The progeny cross to the cultivar Svanhals (NGB 1482, PI 5474) best matched the inheritance pattern expected for cross segregating for the *Zeo2* gene (7). Spikes of plants with the *Zeo2* mutant are compact and remain strap-shaped because all rachis internodes are about the same length. Plants are slightly shorter and spikes contain 2 to 4 more fertile spikelets than those of normal sibs (2). Plants of the Bowman backcross-derived lines with *Zeo2* alleles in the *Zeo2.c* and *Zeo2.ax* group have small anthers (2/3 of normal length) (5) and are associated with small lodicules (closed flowering or cleistogamy) (11, 14). The rachis internode length of plants in Bowman backcross lines for *Zeo2* alleles averaged 3.3 mm compared to 4.5 mm for Bowman. In some field environments, the BW lines for *Zeo2* gene had 1 to 2 more kernels per spike than Bowman, and kernel weights were slightly lower. No effects of the *Zeo2* gene on plant height and grain yield were observed (5). Cleistogamy or closed flowering was strongly associated with a gene for reduced rachis internode length (13), which was later identified as *Zeo2* (9). The *Zeo2* region of 2HL was associated with tolerance to low temperature injury (1) and Fusarium head blight infection (10, 16, 17). Cultivars with the *Zeo2* mutant were demonstrated to have cleistogamous flowering pattern (11). The cleistogamous flower sheds its pollen before opening, forcing plants with this habit to be almost entirely autogamous. The lodicule in cleistogamous plants is atrophied (8). DNA sequencing showed that the *Zeo1* mutants occur in a *Hordeum vulgare* APELATA2 (*AP2*)-like transcription factor, *HvAP2*. The dense spike and cleistogamous (small lodicules) phenotypes are a consequence of a perturbed interaction between microRNA 172 (*Hv-miR172*) and its corresponding binding site on the mRNA from the *HvAP2* gene, which acts early in spike development to regulate turnover of *HvAP2* mRNA (9, 10). The *Zeo1* mutants occur in the last intron of *HvAP2*, the binding site of *Hv-miR172*, and prevent cleavage of the *HvAP2* mRNA (9, 10).

Origin of mutant:

A naturally occurring variant in the fourth exon of the *HvAP2* locus (9), likely of occidental origin in two-rowed barley cultivars from Europe (9, 13); this variant was likely evaluated in Svanhals (NGB 1482, PI 5474) by Hayes and Harlan (7).

Mutational events:

Phenotypically similar variants were isolated from several different accessions (2, 4, 7, 12). However, based on identical DNA haplotypes in the first exon of the *HvAP2* gene, all of the variants have the same gene identified as *Zeo2* (9, 10). Several cultivars from Northern Europe were identified as having the *Zeo2* mutant including Golden Promise (PI 343079), Imperial (OUU352, PI 61340), and Plumage (OUU351, Clho 2511) (9, 10). The variants identified initially as having different origins include *Zeo2.c* from line 36Ab51 (GSHO 637), which was called “good zeocriton” (15), and *Zeo2.ax* from Clho 6880 and Haruna Nijo (3, 9); which likely is the *Zeo2* allele from Golden Melon (OUJ808, PI 263410) as identified in Kanto Nakato Gold (OUJ 518) (8).

Mutant used for description and seed stocks:

Zeo2.c in 36Ab51 (GSHO 637); *Zeo2.c* in Bowman (PI 483237)*4 (GSHO 3433); *Zeo2.c* in Bowman*4 (BW939, NGB 22368); *Zeo2.ax* from Clho 6880 (Ahang/Twisted Flag/Triple-awn Lemma) in Bowman*5 (BW270, NGB 22095); *Zeo2.ax* with *ant22.1508* (Proanthocyanidin-free 22, GSHO 1635) from Haruna Nijo (NGB 13705) in Bowman*4 (GSHO 1841); *Zeo2.ax* with *ant22.1508* in Bowman*6 (BW022, NGB 20430); *Zeo2.c* from 36Ab51 (GSHO 637, ‘good zeocriton’) in Bowman*4 (GSHO 3433, BW939, NGB 22368); *Zeo2.c* from a naked two-rowed accession from China [originally identified as

*dsp1.a (3)] in Bowman*7 (GSHO 1833, BW277, NGB 20561).*

References:

1. Chen, A., L.V. Gusta, A. Brûlé-Babel, R. Leach, U. Baumann, G.B. Fincher , and N.C. Collins. 2009. Varietal and chromosome 2H locus-specific frost tolerance in reproductive tissues of barley (*Hordeum vulgare* L.) detected using a frost simulation chamber. Theor. Appl. Genet. 119:685-694.
2. Chen, A., J. Reinheimer, A. Brûlé-Babel, U. Baumann, M. Pallotta, G.B. Fincher, and N.C. Collins. 2009. Genes and traits associated with chromosome 2H and 5H regions controlling sensitivity of reproductive tissues to frost in barley. Theor. Appl. Genet. 118:1465-1476.
3. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. Plant Physiol. 155:617-627.
4. Engledow, F.L. 1924. Inheritance in barley. III. The awn and the lateral floret (cont'd): fluctuation: a linkage: multiple allelomorphs. J. Genet. 14:49-87.
5. Franckowiak, J.D. (Unpublished).
6. Franckowiak, J.D. 1999. Coordinator's report: Semidwarf genes. Barley Genet. Newsl. 29:74-79.
7. Hayes, H.K., and H.V. Harlan. 1920. The inheritance of the length of internode in the rachis of the barley spike. U.S. Dept. Agr., Bull. 869. 26 pp.
8. Honda, I., Y. Turuspekov, T. Komatsuda, and Y. Watanabe. 2005. Morphological and physiological analysis of cleistogamy in barley (*Hordeum vulgare*). Physiol. Plant 124:524-531.
9. Houston, K., S.M. McKim, J. Comadran, N. Bonar, I. Druka, N. Uzrek, E. Cirillo, J. Guzy-Wrobelska, N.C. Collins, C. Halpin, M. Hansson, C. Dockter, A. Druka, and R. Waugh. 2013. Variation in the interaction between alleles of *HvAPETALA2* and microRNA172 determines the density of grains on the barley inflorescence. Proc. Natl. Acad. Sci. USA 110:16675-16680.
10. Nair, S.K., N. Wang, Y. Turuspekov, M. Pourkheirandish, S. Sinsuwongwat, G. Chen, M. Sameri, A. Tagiri, I. Honda, Y. Watanabe, H. Kanamori, T. Wicker, N. Stein, Y. Nagamura, T. Matsumoto, and T. Komatsuda. 2010. Cleistogamous flowering in barley arises from the suppression of microRNA-guided *HvAP2* mRNA cleavage. Proc. Natl. Acad. Sci. USA 107:490-495.
11. Sameri, M., K. Takeda, and T. Komatsuda. 2006. Quantitative trait loci controlling agronomic traits in recombinant inbred lines from a cross of oriental- and occidental-type barley cultivars. Breed. Sci. 56:243-252.
12. Smith, L. 1951. Cytology and genetics of barley. Bot. Rev. 17:1-51, 133-202, 285-355.
13. Turuspekov, Y., N. Kawada, I. Honda, Y. Watanabe, and T. Komatsuda. 2005. Identification and mapping of a QTL for rachis internode length associated with cleistogamy in barley. Plant Breed. 124:542-545.
14. Turuspekov, Y., Y. Mano, I. Honda, N. Kawada, Y. Watanabe, and T. Komatsuda. 2004. Identification and mapping of cleistogamy gene in barley. Theor. Appl. Genet. 109:480-487.
15. USDA-ARS. No date. List of Genetic Stocks in Barley. USDA-ARS National Small Grains Germplasm Research Facility, Aberdeen, Idaho.
16. Yu, G.T., J.D. Franckowiak, S.M. Neate, B. Zhang and R.D. Horsley. 2010. A native QTL for Fusarium head blight resistance in North American barley (*Hordeum vulgare* L.)

Barley Genetics Newsletter (2017): 46:50-187.

independent of height, maturity, and spike type loci. *Genome* 53:111-116.

17. Yu, G.T., R.D. Horsley, B. Zhang, and J.D. Franckowiak. 2010. A new semi-dwarfing gene identified by molecular mapping of quantitative trait loci in barley. *Theor. Appl. Genet.* 120:853-861.

Prepared:

J.D. Franckowiak. 2002. *Barley Genet. Newsl.* 32:115.

Revised:

J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:193-194.

J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:177-180.

Stock number: BGS 679
Locus name: Accordion rachis 4
Locus symbol: *acr4*

Previous nomenclature and gene symbolization:

Accordion rachis 3 = *acr-3* (3).

Inheritance:

Monofactorial recessive (2, 3).

Located in chromosome 2H or/and 6HL (1); *acr4.3* is primarily associated with SNP markers 1_0891 to 2_1533 (positions 54.47 to 141.56 cM) in 2H bins 05 to 10 of the Bowman backcross-derived line BW004 and is associated with SNP markers 1_0015 to 2_0733 (positions 160.38 to 180.69 cM) in 6HL bins 10 to 11 of BW004 (1).

Description:

The *acr4.3* mutant plants have multiple changes from normal besides elongated rachis internodes (3). Plant height is reduced, maturity is delayed, awns are short, the basal rachis internode is elongated, and partial fertility occurs (3). Plants of the Bowman backcross-derived line for *acr4.3*, BW004, showed similar alterations in plant morphology (2). The two largest DNA segments of BW004 in which donor parent SNP markers are retained overlap regions where lax spike loci have been mapped: 2HL (*acr1*, accordion rachis 1) and 6HL (*lax-c*, *laxatum-c*) (1, 2).

Origin of mutant:

An ethylene imine induced mutant in Bonus (NGB 14657, PI 189763) (3).

Mutational events:

acr4.3 (NGB 115043) in Bonus (NGB 14657, PI 189763) (2, 3).

Mutant used for description and seed stocks:

acr4.3 (NGB 115043) in Bonus; *acr4.3* in Bowman (PI 483237)*3 (BW004, NGB 20412).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Lundqvist, U. (Unpublished).

Prepared:

U. Lundqvist and J.D. Franckowiak. 2011. *Barley Genet. Newsl.* 41:201.

Revised:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:181.

Barley Genetics Newsletter (2017): 46:50-187.
BGS 740, Pubescence on glume 1, *Pbg1*

Stock number: BGS 740
Locus name: Pubescence on outer glume 1
Locus symbol: *Pbg1*

Previous nomenclature:

Pubescence on the outer glume = *S* (1).
Pubescence on outer glume = *Pbg* (2).

Inheritance:

Monofactorial recessive (1).
Located in chromosome 7H (1); *pbg1.b* is about 25.3 cM from the naked caryopsis 1 (*nud1*) locus based on coupling data (1); *pdg1.b* is closer to the dense spike 1 (*dsp1*) locus than the *nud1* locus (1).

Description:

Plants with the incomplete dominant allele, *Pbg1.a*, have the outside surface of the outer glumes covered with pubescence on short hairs. Plants with the recessive allele, *pbg1.b*, pubescence restricted to a central ridge on the outer glume (1). The short rachilla hair 1 (*srh1.a*) allele reduces the length of the hairs on the outer glumes, but not their distribution (1). Other studies in which inheritance of glume hairs is more complex were reviewed by Smith (2). Wiebe and Reid (3) classified the distribution of hairs on the outer surface of glumes into four groups: fully covered, narrow band, midline only, and absent.

Origin of mutant:

Natural occurrence with naked caryopsis in line No. 154 (1); hairs confined to bands in Montcalm (Clho 7149) (3); hairs restricted to the midline in Otis (Clho 7557) (3).

Mutant used for description:

pbg1.b in line No. 154, possibly GSHO 173 (1).

Mutant used for description and seed stocks:

pbg1.b in line No. 154 and likely in a number of six-rowed cultivars from East Asia.

References:

1. Hor, K.S. 1924. Interrelations of genetic factors in barley. *Genetics* 9:151-180.
2. Smith, L. 1951. Cytology and genetics of barley. *Bot. Rev.* 17:1-51, 133-202, 285-355.
3. Wiebe, G.A., and D. A. Reid. 1961. Classification of barley varieties grown in the United States and Canada 1958. U. S. Dept. Agric. Technical Bulletin No 154. 234 pp.

Prepared:

J.D. Franckowiak 2017. *Barley Genet. Newsl.* 47:182.

BGS 741, Stubble 1, *stb1*

Stock number: BGS 741
Locus name: Stubble 1
Locus symbol: *stb1*

Previous nomenclature:

Gillette (2, 4).
Stubble = *stb* (3, 4).
Short rachilla hair 2 = *srh2* (1).

Inheritance:

Monofactorial recessive (3, 4).
Location is unknown; *stb1.a* is associated with several groups of retained SNP markers in 1H (positions 61.55 to 157.02 cM in bins 06 to 12), 3H (52.41 to 119.10 cM in bins 04 to 07), and 5H (26.06 to 98.42 cM in bins 02 to 10) in the Bowman backcross-derived line BW874 (1).

Description:

Gillette or stubble 1, *stb1.a*, is expressed on the rachilla, glume, and rachis margin hairs. The effect looks as if a mower had been run over these areas leaving a "stubble" of very short hairs. This expression can readily be seen in both lines with long rachilla hair (*Srh1*) and short rachilla hair (*srh1*) (3, 4). The stubble trait cannot be scored easily in certain smooth-awned barleys where most, and sometimes all, hairs on the glumes and rachis margins are missing. In an F₂ population, segregation of the stubble trait appeared independent of the short rachilla hair trait (4). The stubble trait was found in the world collection barleys from Korea and adjacent China (4). Isogenic lines with "stubble" (*stb1*) and short rachilla hair 1 (*srh1*) were selected from a Manchuria (CIho 2330) X Gillette (CIho 4485) cross (4). Both parents have rough awns (*Raw1* plus *Raw2*).

Origin of mutant:

Natural occurrence in US 85 [GSHO 656, selected from CIho 4551 (PI 69686) introduced from Jilin, China] and US 86 [GSHO 659, selected from CIho 4485 (PI 69621) from Shanxi, China] (2, 4).

Mutational events:

stb1.a in GSHO 659 (T12-34) selected from CIho 4485 (PI 69621) (2).

Mutant used for description and seed stocks:

stb1.a in GSHO 656 and GSHO 659; *stb1.a* from GSHO 656 in Bowman (PI 483237)*2 (BW874, NGB 22308). The isogenic lines for *stb1.a* were selected in the F₂₅ generation from a Manchuria (CIho 2330) X Gillette (CIho 4485) cross: CIho 16005 (*Srh1 Stb1*), CIho 16006 (*Srh1 stb1.a*), CIho 16007 (*srh1.a Stb1*), and CIho *srh1.a stb1.a*.

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Hang, A. 1995. List of genetic stocks in barley arranged in numerical order of BGS (Barley Genetic Stock) numbers. USDA-ARS National Small Grains Germplasm Research Facility, Aberdeen, ID 83210. (Notes from T. Tsuchiya on the collection of barley genetic stocks at Colorado State University.)
3. Reid, D.A., and G.A. Wiebe. 1968. Taxonomy, botany, classification, and world

- Barley Genetics Newsletter (2017): 46:50-187.*
collection. p. 61-84. *In* Barley: Origin, Botany, Culture, Winterhardiness, Genetics, Utilization, Pests. USDA Agric. Handb. No. 338.
4. Wiebe, G.A. 1972. "Stubble" gene. *Barley Genet. Newsl.* 2:109.
- Prepared:
J.D. Franckowiak 2017. *Barley Genet. Newsl.* 47:183-184.

BGS 743, Tweaky spike 1, *twk1*

Stock number: BGS 743
Locus name: Tweaky spike 1
Locus symbol: *twk1*

Previous nomenclature:

Tweaky = *tw* (1, 5).

Tweaked spike (3).

Inheritance:

Monofactorial recessive (5).

Located in chromosome 7H (5).

Description:

Tweaky is a spike development abnormality in which one or more rachis internodes in the upper third of the spike are elongated and rounded in shape giving the appearance of a short stem. As a result, the spike is misshapen or twisted and appears to have missing kernels (6). Often rachis internodes just above the tweaked area are foreshortened and give a bunched appearance of spikelets. The tweaky characteristic has been reported only in the cultivar Wisconsin Barbless (CIho 5105) where it occurs in only a small percentage of the spikes (6). See Reid and Wiebe (3) for a photo of the *twk1.a* mutant. Chemically induced mutants in barley that exhibit instability in inflorescence and floral organ development have been isolated (4, 7). These mutants resemble some of the traits associated with *twk1.a* mutant. Partial rescue of specific phenotypic features of tweaky mutants following treatment with auxin inhibitors and the synthetic auxin 2,4-dichloroacetic acid (2,4-D) has been reported (4).

Origin of mutant:

Natural occurrence in Wisconsin Barbless (CIho 5105) (1, 6).

Mutational events:

twk1.a in Wisconsin Barbless (CIho 5105) (1, 6); a tweaky-like spike phenotype might be present in the two-rowed cultivar ND Genesis (PI 677345) (2). Chemically induced mutants in Auksiniai II have somewhat similar, but more extreme, phenotypes (4, 7); however, they have not been tested for allelism with the *twk1.a* mutant.

Mutant used for description and seed stocks:

twk1.a in Wisconsin Barbless.

References:

1. Åberg, E., and G.A. Wiebe. 1948. Taxonomic Value of Characters in Cultivated Barley. U. S. Dept. Agr. Tech, Bul. 942. 88 pp.
2. Franckowiak, J.D. (Unpublished).
3. Reid, D.A., and G.A. Wiebe. 1968. Taxonomy, botany, classification, and world collection. p. 61-84. *In* Barley: Origin, Botany, Culture, Winterhardiness, Genetics, Utilization, Pests. USDA Agric. Handb. No. 338.
4. Šiukštė, R., V. Vaitkūnienė, G. Kaselytė, V. Okockytė, J. Žukauskaitė, D. Žvingila, and V. Rancelis. 2015. Inherited phenotype instability of inflorescence and floral organ development in homeotic barley double mutants and its specific modification by auxin inhibitors and 2,4-D. *Ann. Bot.* 115:651-663.
5. Smith, L. 1951. Cytology and genetics of barley. *Bot. Rev.* 17:1-51, 133-202, 285-355.
6. Wiebe, G.A., and D.A. Reid 1961. Classification of Barley Varieties Grown in the United States and Canada in 1958. U. S. Dept. Agr. Tech, Bul. 1224. 234 pp.

Barley Genetics Newsletter (2017): 46:50-187.

7. Žvingila, D., V. Vaitkūnienė, J. Patamsytė, A. Leistrumaitė, M. Staniūtė, L. Balčiūnienė, T. Cėsnienė, V. Kleizaitė, R. Šiukšta, and V. Rančelis. 2012. DNA polymorphism and agronomic traits of revertants from barley (*Hordeum vulgare* L.) mutant *tw*. *Žemdirbystė = Agriculture* 99:138-148.

Prepared:

J.D. Franckowiak 2017. *Barley Genet. Newsl.* 47:185-186.

Stock number: BGS 744
Locus name: Accordion rachis 5
Locus symbol: *acr5*

Previous nomenclature and gene symbolization:

Accordion rachis 2 = *acr-2* (3).

Inheritance:

Monofactorial recessive (2, 3).

Located in chromosome 7H (1); *acr5.2* is associated with SNP markers 1_0721 to 2_0103 (positions 82.82 to 139.96 cM) in 7H bins 06 to 08 of the Bowman backcross-derived line BW003 (1).

Description:

The *acr5.2* mutant plants exhibited low plant vigor, elongated rachis internodes, and a reduced number of kernels per spike (3). Plants of the Bowman backcross-derived line for *acr5.2*, BW003, showed reduced plant vigor (2). Compared to Bowman, rachis internodes of BW003 were slightly longer (5.1 vs. 4.3 mm), plants and awns were slightly shorter, and peduncles were slightly longer. Kernels of BW003 were elongated (11.3 vs. 9.7 mm) and thin (3.4 vs. 3.9 mm) and weighed less (3.5 vs. 5.9 mg). Grain yields of BW003 were highly variable, but never more than half those of Bowman (2).

Origin of mutant:

A neutron induced mutant in Bonus (NGB 14657, PI 189763) (1, 3).

Mutational events:

acr5.2 (NGB 115042) in Bonus (NGB 14657, PI 189763) (2, 3).

Mutant used for description and seed stocks:

acr5.2 (NGB 115042, GSHO 2153) in Bonus; *acr5.2* in Bowman (PI 483237)*6 (BW003, NGB 20411).

References:

1. Druka, A., J. Franckowiak, U. Lundqvist, N. Bonar, J. Alexander, K. Houston, S. Radovic, F. Shahinnia, V. Vendramin, M. Morgante, N. Stein, and R. Waugh. 2011. Genetic dissection of barley morphology and development. *Plant Physiol.* 155:617-627.
2. Franckowiak, J.D. (Unpublished).
3. Lundqvist, U. (Unpublished).

Prepared:

U. Lundqvist and J.D. Franckowiak. 2017. *Barley Genet. Newsl.* 47:187.